

BRD EXT

1

August 12, 2004

Environment Canada Nunavut Regional Office **Environmental Protection Branch** P.O. Box 1870 Igaluit, NU XOA OHO

Phyllis
Sid Bruinsma, Enforcement / Emergencies Officer, Nunavut Northern Division

Dear Sir:

Polaris Mine - Garrow Lake Final Discharge Point Exceedance of TSS Sample Under Schedule 4 of Re: the MMER

In accordance with section 24 (2) of the Metal Mining Effluent Regulations ("MMER"), please be advised that on August 9, 2004 we received laboratory results indicating that a single weekly effluent sample taken July 7th, 2004 from the final discharge point of Garrow Lake (a single grab sample) had a Total Suspended Solids (TSS) level of 117 mg/L which exceeded the TSS limits in Schedule 4 of the MMER (limit of 30 mg/L for a single grab sample). Concurrently on July 7th we also took our monthly acute lethality samples. The results were compliant with the MMER and showed no acute toxicity to either the Rainbow Trout or the Daphnia magna (i.e. LC50s in both cases were > 100% effluent). I have attached copies of both of these laboratory results. The date on the laboratory record for the metals analysis states that the sample was taken on July 6th but this is incorrect. The metals samples were originally scheduled to be taken on the 6th and the bottles were labeled as such by our consultant but fog delayed the plane that ships the samples to the laboratory so the sample was actually taken on the 7th at the same time as the acute lethality samples (and the dates on the bottles and associated chain of custody papers were not updated).

The July 7th sample was taken during the first week of effluent discharge from Garrow Lake. The TSS level was the result of Garrow Lake being ice covered which blocks the creek inlet (Garrow Creek) until the ice melted sufficiently to uncork the lake, and then the flows increase to a peak very quickly causing turbidity in the creek channel. The peak flows last a day or two and then the flows gradually subside with the TSS levels decreasing concurrently. As stated previously, the turbidity is not from tailings being mobilized as evidenced by the metals concentrations in the July 7th sample being in compliance (i.e. Zn concentrations were 0.198 mg/L). The TSS levels in the subsequent weekly water sample (July 13th) was in compliance (TSS was 5.7 mg/L) and metals concentrations continued to be in compliance (Zn was 0.106 mg/L). I have also attached the final results for the July 20th weekly water samples (TSS was <3.0 mg/L and the Zn was 0.0435 mg/L). The July 27th weekly water sample interim laboratory results report TSS at 3.7 mg/L and the Zn results are not available yet. Samples for the acute lethality tests were also taken on July 27th and compliant results were again achieved with the LC50 being >100 for both

the Rainbow Trout and the Daphnia magna (attached). I will forward the final July 27th laboratory results once they are received.

If you have any other questions in the interim, please contact me at (250) 427-8405 or by email at bruce.donald@teckcominco.com

Yours truk

Bruce Donald

Attachments:

July 7th Monthly Water Sample Results July 7th Acute Lethality Sample Results July 13th Weekly Water Sample Results July 20th Weekly Water Sample Results July 27th Acute Lethality Sample Results

Walter Kuit (Teck Cominco) Cc:

> Bob Hutchinson (Teck Cominco) Patrick Allard (Azimuth Consulting) Phyllis Beaulieu (Nunavut Water Board)

ALS Environmental



CHEMICAL ANALYSIS REPORT

Date:

August 9, 2004

ALS File No.

U5572

Report On:

Polaris Water Analysis

Report To:

Azimuth Consulting Group Inc.

218 - 2902 West Broadway

Vancouver, BC

V6K 2G8

Attention:

Mr. Randy Baker

Received:

July 12, 2004

ALS ENVIRONMENTAL

per:

Heather A. Ross-Easton, B.Sc. - Project Chemist

Andre Langlais, M.Sc. - Project Chemist

RESULTS OF ANALYSIS - Water



Sample ID			G Creek	Dup	F Blank	T Blank
Sample Date Sample Time ALS ID			04-07-06 11:00 1	04-07-06 11:00 2	04-07-06 11:00 3	04-07-06 11:00 4
Physical Tests Hardness pH Salinity o/d Total Suspended	CaCO3		1400 8.05 7.0 117	1400 8.06 6.9 120	<0.54	<0.54
Dissolved Anion Alkalinity-Total	<u>1S</u>	CaCO3	138	132	-	-
Nutrients Ammonia Nitrogen	en	N N	0.071 0.277	0.069 0.284	<u>.</u>	-
<u>Cyanides</u> Total Cyanide	CN		<0.0050	<0.0050	-	-
Arsenic 1 Cadmium 1 Calcium 1	Γ-AI Γ-As Γ-Cd Γ-Ca Γ-Cu		0.34 <0.0010 0.000588 140 0.00265	0.26 <0.0010 0.000582 139 0.00252	<0.10 <0.0010 <0.000020 <0.050 0.00012	<0.10 <0.0010 <0.000020 <0.050 0.00012
Lead 1 Magnesium 1 Mercury 1	Г-Fe Г-Pb Г-Mg Г-Hg Г-Mo		0.487 0.00269 256 <0.000010 <0.0050	0.441 0.00240 256 <0.000010 <0.0050	<0.010 0.00017 <0.10 <0.000010 <0.0050	<0.010 0.00021 <0.10 <0.000010 <0.0050
	Γ-Ni Γ-Zn		0.00442 0.198	0.00438 0.196	<0.00050 0.0012	<0.00050 <0.0010
Radiological Par Radium-226 1.2	rameters		0.0200	0.0200	<0.0050	0.0060

Results are expressed as milligrams per litre except where noted. < = Less than the detection limit indicated.

¹Results are expressed as Becquerels per litre (Bq/L). This analysis is
²subcontracted to SRC from Saskatoon.

Appendix 1 - METHODOLOGY



Outlines of the methodologies utilized for the analysis of the samples submitted are as follows

Conventional Parameters in Water

These analyses are carried out in accordance with procedures described in "Methods for Chemical Analysis of Water and Wastes" (USEPA), "Manual for the Chemical Analysis of Water, Wastewaters, Sediments and Biological Tissues" (BCMOE), and/or "Standard Methods for the Examination of Water and Wastewater" (APHA). Further details are available on request.

pH in Water

This analysis is carried out using procedures adapted from APHA Method 4500-H "pH Value". The pH is determined in the laboratory using a pH electrode.

Recommended Holding Time:

Sample: 2 hours Reference: APHA

For more detail see ALS Environmental "Collection & Sampling Guide"

Solids in Water

This analysis is carried out using procedures adapted from APHA Method 2540 "Solids". Solids are determined gravimetrically. Total dissolved solids (TDS) and total suspended solids (TSS) are determined by filtering a sample through a glass fibre filter, TDS is determined by evaporating the filtrate to dryness at 180 degrees celsius, TSS is determined by drying the filter at 104 degrees celsius. Total solids are determined by evaporating a sample to dryness at 104 degrees celsius. Fixed and volatile solids are determined by igniting a dried sample residue at 550 degrees celsius.

Recommended Holding Time:

Sample: 7 days Reference: APHA

For more detail see ALS Environmental "Collection & Sampling Guide"

Alkalinity in Water by Titration

This analysis is carried out using procedures adapted from APHA Method 2320 "Alkalinity". Total alkalinity is determined by potentiometric titration to a pH 4.5 endpoint. Bicarbonate, carbonate and hydroxide alkalinity are calculated from phenolphthalein alkalinity and total alkalinity values.

Recommended Holding Time:

Sample: 14 days Reference: APHA

Appendix 1 - METHODOLOGY - Continued



For more detail see ALS Environmental "Collection & Sampling Guide"

Ammonia in Water by Selective Ion Electrode

This analysis is carried out, on sulphuric acid preserved samples, using procedures adapted from APHA Method 4500-NH3 "Nitrogen (Ammonia)". Ammonia is determined using an ammonia selective electrode.

Recommended Holding Time:

Sample: 28 days Reference: APHA

For more detail see ALS Environmental "Collection & Sampling Guide"

Dissolved Anions in Water by Ion Chromatography

This analysis is carried out using procedures adapted from APHA Method 4110 "Determination of Anions by Ion Chromatography" and EPA Method 300.0 "Determination of Inorganic Anions by Ion Chromatography". Anions are determined by filtering the sample through a 0.45 micron membrane filter and injecting the filtrate onto a Dionex IonPac AG17 anion exchange column with a hydroxide eluent stream. Anions routinely determined by this method include: bromide, chloride, fluoride, nitrate, nitrite and sulphate.

Recommended Holding Time:

Sample: 28 days (bromide, chloride, fluoride, sulphate)

Sample: 2 days (nitrate, nitrite) Reference: APHA and EPA

For more detail see ALS Environmental "Collection & Sampling Guide"

Cyanide Species in Water

This analysis is carried out using procedures adapted from APHA Method 4500-CN "Cyanide". Total or strong acid dissociable (SAD) cyanide and weak acid dissociable (WAD) cyanide are determined by sample distillation and analysis using the chloramine-T colourimetric method. Cyanate is determined by the cyanate hydrolysis method using an ammonia selective electrode. Thiocyanate is determined by the ferric nitrate colourimetric method.

Recommended Holding Time:

Sample: 14 days Reference: APHA

For more detail see ALS Environmental "Collection & Sampling Guide"

Metals in Seawater

This analysis is carried out using procedures adapted from "Recommended Guidelines for Measuring Metals in Puget Sound Marine Water, Sediment, and Tissue Samples" prepared for the United States Environmental Protection Agency and the Puget Sound Water Quality Authority, 1995. The procedures

Appendix 1 - METHODOLOGY - Continued



may involve preliminary sample treatment by acid digestion or filtration (EPA Method 3005A). Instrumental analysis of the seawater is by atomic absorption/emission spectrophotometry (EPA Method 7000 series), inductively coupled plasma - optical emission spectrophotometry (EPA Method 6010B), and/or inductively coupled plasma - mass spectrometry (EPA Method 6020).

Recommended Holding Time:

Sample: 6 months Reference: Puget

For more detail see ALS Environmental "Collection & Sampling Guide"

Trace Metals in Seawater by SPR-IDA Chelation

This analysis is carried out using procedures adapted from "Recommended Guidelines for Measuring Metals in Puget Sound Marine Water, Sediment, and Tissue Samples" prepared for the United States Environmental Protection Agency and the Puget Sound Water Quality Authority, 1995, and with procedures adapted from Cetac Technologies Incorporated. A suspended particulate resin (SPR), consisting of immobilized iminodiacetate (IDA) on a divinylbenzene polymer, is used to chelate and preconcentrate metals in seawater. Instrumental analysis is by inductively coupled plasma mass spectrometry (ICPMS) and/or routine atomic absorption spectrophotometry techniques (EPA 7000 series).

Recommended Holding Time:

Sample/Extract: 6 months Reference: Puget

For more detail see ALS Environmental "Collection & Sampling Guide"

Metals in Water

This analysis is carried out using procedures adapted from "Standard Methods for the Examination of Water and Wastewater" 20th Edition 1998 published by the American Public Health Association, and with procedures adapted from "Test Methods for Evaluating Solid Waste" SW-846 published by the United States Environmental Protection Agency (EPA). The procedures may involve preliminary sample treatment by acid digestion, using either hotplate or microwave oven, or filtration (EPA Method 3005A). Instrumental analysis is by atomic absorption/emission spectrophotometry (EPA Method 7000 series), inductively coupled plasma - optical emission spectrophotometry (EPA Method 6010B), and/or inductively coupled plasma - mass spectrometry (EPA Method 6020).

Recommended Holding Time:

Sample: 6 months Reference: EPA

For more detail see: ALS "Collection & Sampling Guide"

Appendix 1 - METHODOLOGY - Continued



Mercury in Seawater

This analysis is carried out using procedures adapted from "Recommended Guidelines for Measuring Metals in Puget Sound Marine Water, Sediment, and Tissue Samples" prepared for the United States Environmental Protection Agency and the Puget Sound Water Quality Authority, 1995. The procedure involves a cold-oxidation of the acidified seawater sample using bromine monochloride prior to reduction of the sample with stannous chloride. Instrumental analysis is by cold vapour atomic fluorescence spectrophotometry (EPA Method 245.7).

Recommended Holding Time:

Sample: 28 days Reference: Puget

For more detail see ALS Environmental "Collection & Sampling Guide"

Mercury in Water

This analysis is carried out using procedures adapted from "Standard Methods for the Examination of Water and Wastewater" 20th Edition 1998 published by the American Public Health Association, and with procedures adapted from "Test Methods for Evaluating Solid Waste" SW-846 published by the United States Environmental Protection Agency (EPA). The procedure involves a cold-oxidation of the acidified sample using bromine monochloride prior to reduction of the sample with stannous chloride. Instrumental analysis is by cold vapour atomic fluorescence spectrophotometry (EPA Method 245.7).

Recommended Holding Time:

Sample: 28 days Reference: EPA

For more detail see ALS Environmental "Collection & Sampling Guide"

Results contained within this report relate only to the samples as submitted.

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End of Report



195 Pemberton Avenue North Vancouver, BC Canada V7P 2R4 Fax: 604-662-8548

Tel: 604-986-4331

Info@evsenvironment.com www.evsenvironment.com

FAX TRANSMITTAL SHEET

TO:	Cheryl Mackintosh	DATE:	July 16, 2004
	Azimuth Consulting Group	PROJECT No.:	09-0302-54
	218 – 2902 West Broadway	W.O. No.:	0400304,307
	Vancouver, BC	FAX No.:	604-739-9070
	V6K 2G8	TEL No.:	604-730-1220
SENT BY:	Robert Harrison	# PAGES (incl. o	cover): One (1)

This message is intended only for the use of the individual or entity to which it is addressed and may contain information that is privileged confidential and exempt from disclosure under applicable law. If you have received this communication in error please notify us by telephone (collect) and return the original transmission to us by mail without making a copy. Thank you.

Re: Interim data for the 96-h rainbow trout and 48-h Daphnia magna LC50 toxicity tests performed on the sample identified as G-Creek acute 070704 (collected July 7, 2004).

Sample ID	Sample Collection	LC50 (95% Confidence Limits) [%vol/vol]		
	Date	96-h Rainbow Trout	48-h Daphnia magna	
G-Creek acute 070704	July 7, 2004 (2200h)	> 100	> 100	

Please note that these are draft results and are subject to a QA/QC review. A complete report will follow by mail. Should you have any questions, please contact Edmund Canaria, Manager of Laboratory Services, or myself at 604-986-4331.

Thank you,

Robert Harrison, B.Sc. Hons. Assistant Bioassay Test Supervisor – Fish Team <u>rharrison@evsenvironment.com</u>

REH

ALS Environmental



CHEMICAL ANALYSIS REPORT

Date: August 9, 2004

ALS File No. U5829

Report On: Polaris Water Analysis

Report To: Azimuth Consulting Group Inc.

218 - 2902 West Broadway

Vancouver, BC V6K 2G8

Attention: Mr. Randy Baker

Received: July 16, 2004

ALS ENVIRONMENTAL

per:

Heather A. Ross-Easton, B.Sc. - Project Chemist Andre Langlais, M.Sc. - Project Chemist

L-East

REMARKS



The detection limits for some of the metals have been increased for the sample "G Creek" due to sample matrix interferences.

RESULTS OF ANALYSIS - Water



Sample ID	G Creek
Sample Date ALS ID	04-07-13 1
Physical Tests pH Salinity o/oo Total Suspended Solids	7.90 4.5 5.7
Cyanides Total Cyanide CN	<0.0050
Total Metals Aluminum T-Al Arsenic T-As Cadmium T-Cd Copper T-Cu Iron T-Fe	<0.40 <0.0020 0.000332 0.000703 0.019
Lead T-Pb Molybdenum T-Mo Nickel T-Ni Zinc T-Zn	0.000318 <0.0050 0.00204 0.106
Radiological Parameters Radium-226 1.2	0.0070

Remarks regarding the analyses appear at the beginning of this report. Results are expressed as milligrams per litre except where noted. < = Less than the detection limit indicated.
¹Result is expressed as Becquerels per litre (Bq/L). This analysis is ²subcontracted to SRC, Saskatoon.

Appendix 1 - METHODOLOGY



Outlines of the methodologies utilized for the analysis of the samples submitted are as follows

pH in Water

This analysis is carried out using procedures adapted from APHA Method 4500-H "pH Value". The pH is determined in the laboratory using a pH electrode.

Recommended Holding Time:

Sample: 2 hours Reference: APHA

For more detail see ALS Environmental "Collection & Sampling Guide"

Conventional Parameters in Water

These analyses are carried out in accordance with procedures described in "Methods for Chemical Analysis of Water and Wastes" (USEPA), "Manual for the Chemical Analysis of Water, Wastewaters, Sediments and Biological Tissues" (BCMOE), and/or "Standard Methods for the Examination of Water and Wastewater" (APHA). Further details are available on request.

Solids in Water

This analysis is carried out using procedures adapted from APHA Method 2540 "Solids". Solids are determined gravimetrically. Total dissolved solids (TDS) and total suspended solids (TSS) are determined by filtering a sample through a glass fibre filter, TDS is determined by evaporating the filtrate to dryness at 180 degrees celsius, TSS is determined by drying the filter at 104 degrees celsius. Total solids are determined by evaporating a sample to dryness at 104 degrees celsius. Fixed and volatile solids are determined by igniting a dried sample residue at 550 degrees celsius.

Recommended Holding Time:

Sample: 7 days Reference: APHA

For more detail see ALS Environmental "Collection & Sampling Guide"

Cyanide Species in Water

This analysis is carried out using procedures adapted from APHA Method 4500-CN "Cyanide". Total or strong acid dissociable (SAD) cyanide and weak acid dissociable (WAD) cyanide are determined by sample distillation and analysis using the chloramine-T colourimetric method. Cyanate is determined by the cyanate hydrolysis method using an ammonia selective electrode. Thiocyanate is determined by the ferric nitrate colourimetric method.

Recommended Holding Time:

Appendix 1 - METHODOLOGY - Continued



Sample: 14 days Reference: APHA

For more detail see ALS Environmental "Collection & Sampling Guide"

Metals in Water

This analysis is carried out using procedures adapted from "Standard Methods for the Examination of Water and Wastewater" 20th Edition 1998 published by the American Public Health Association, and with procedures adapted from "Test Methods for Evaluating Solid Waste" SW-846 published by the United States Environmental Protection Agency (EPA). The procedures may involve preliminary sample treatment by acid digestion, using either hotplate or microwave oven, or filtration (EPA Method 3005A). Instrumental analysis is by atomic absorption/emission spectrophotometry (EPA Method 7000 series), inductively coupled plasma - optical emission spectrophotometry (EPA Method 6010B), and/or inductively coupled plasma - mass spectrometry (EPA Method 6020).

Recommended Holding Time:

Sample: 6 months Reference: EPA

For more detail see: ALS "Collection & Sampling Guide"

Metals in Seawater

This analysis is carried out using procedures adapted from "Recommended Guidelines for Measuring Metals in Puget Sound Marine Water, Sediment, and Tissue Samples" prepared for the United States Environmental Protection Agency and the Puget Sound Water Quality Authority, 1995. The procedures may involve preliminary sample treatment by acid digestion or filtration (EPA Method 3005A). Instrumental analysis of the seawater is by atomic absorption/emission spectrophotometry (EPA Method 7000 series), inductively coupled plasma - optical emission spectrophotometry (EPA Method 6010B), and/or inductively coupled plasma - mass spectrometry (EPA Method 6020).

Recommended Holding Time:

Sample: 6 months Reference: Puget

For more detail see ALS Environmental "Collection & Sampling Guide"

Trace Metals in Seawater by SPR-IDA Chelation

This analysis is carried out using procedures adapted from "Recommended Guidelines for Measuring Metals in Puget Sound Marine Water, Sediment, and Tissue Samples" prepared for the United States Environmental Protection Agency and the Puget Sound Water Quality Authority, 1995, and with procedures adapted from Cetac Technologies Incorporated. A suspended particulate resin (SPR), consisting of immobilized iminodiacetate (IDA) on a divinylbenzene polymer, is used to chelate and preconcentrate metals in seawater. Instrumental analysis is by inductively coupled plasma mass spectrometry (ICPMS) and/or routine atomic absorption spectrophotometry techniques (EPA 7000 series).

Appendix 1 - METHODOLOGY - Continued



Recommended Holding Time:

Sample/Extract: 6 months Reference: Puget

For more detail see ALS Environmental "Collection & Sampling Guide"

Results contained within this report relate only to the samples as submitted.

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End of Report

CHEMICAL ANALYSIS REPORT

Date: August 11, 2004

ALS File No. U6161

Report On: Polaris Water Analysis

Report To: Azimuth Consulting Group Inc.

218 - 2902 West Broadway

Vancouver, BC

V6K 2G8

Attention: Mr. Randy Baker

Received: July 24, 2004

ALS ENVIRONMENTAL

per:

Andre Langlais, M.Sc. - Project Chemist Heather A. Ross-Easton, B.Sc. - Project Chemist

RESULTS OF ANALYSIS - Water

Sample ID Sample Date ALS ID	G-Creek_ Gen_ 072004 04-07-20	Tracer Dye 04-07-20 2	Injectn. Dye 04-07-20	G-Creek_ Rhod_ 072004 04-07-20
Physical Tests pH Salinity o/oo Total Suspended Solids	7.86 3.3 <3.0	-	:	- - -
Cyanides Total Cyanide CN Total Metals	<0.0050	-	-	-
Aluminum T-Al Arsenic T-As Cadmium T-Cd Copper T-Cu Iron T-Fe	<0.20 <0.0020 0.000109 0.000427 <0.010	- - - -		
Lead T-Pb Molybdenum T-Mo Nickel T-Ni Zinc T-Zn	0.000837 <0.0050 0.000876 0.0435	- - -	- - - -	- - -
Miscellaneous Rhodamine		see below ¹	76900	0.0380
Radiological Parameters Radium-226 2.3	<0.0050	-	-	_

Results are expressed as milligrams per litre except where noted. <= Less than the detection limit indicated.

¹This sample was used to prepare calibration standards
²Result is expressed as Becquerels per litre (Bq/L). This analysis is
³subcontracted to SRC, Saskatoon.

Appendix 1 - METHODOLOGY

Outlines of the methodologies utilized for the analysis of the samples submitted are as follows

pH in Water

This analysis is carried out using procedures adapted from APHA Method 4500-H "pH Value". The pH is determined in the laboratory using a pH electrode.

Recommended Holding Time:

Sample: 2 hours Reference: APHA

For more detail see ALS Environmental "Collection & Sampling Guide"

Conventional Parameters in Water

These analyses are carried out in accordance with procedures described in "Methods for Chemical Analysis of Water and Wastes" (USEPA), "Manual for the Chemical Analysis of Water, Wastewaters, Sediments and Biological Tissues" (BCMOE), and/or "Standard Methods for the Examination of Water and Wastewater" (APHA). Further details are available on request.

Solids in Water

This analysis is carried out using procedures adapted from APHA Method 2540 "Solids". Solids are determined gravimetrically. Total dissolved solids (TDS) and total suspended solids (TSS) are determined by filtering a sample through a glass fibre filter, TDS is determined by evaporating the filtrate to dryness at 180 degrees celsius, TSS is determined by drying the filter at 104 degrees celsius. Total solids are determined by evaporating a sample to dryness at 104 degrees celsius. Fixed and volatile solids are determined by igniting a dried sample residue at 550 degrees celsius.

Recommended Holding Time:

Sample: 7 days Reference: APHA

For more detail see ALS Environmental "Collection & Sampling Guide"

Cyanide Species in Water

This analysis is carried out using procedures adapted from APHA Method 4500-CN "Cyanide". Total or strong acid dissociable (SAD) cyanide and weak acid dissociable (WAD) cyanide are determined by sample distillation and analysis using the chloramine-T colourimetric method. Cyanate is determined by the cyanate hydrolysis method using an ammonia selective electrode. Thiocyanate is determined by the ferric nitrate colourimetric method.

Recommended Holding Time:

Appendix 1 - METHODOLOGY - Continued

Sample: 14 days Reference: APHA

For more detail see ALS Environmental "Collection & Sampling Guide"

Metals in Water

This analysis is carried out using procedures adapted from "Standard Methods for the Examination of Water and Wastewater" 20th Edition 1998 published by the American Public Health Association, and with procedures adapted from "Test Methods for Evaluating Solid Waste" SW-846 published by the United States Environmental Protection Agency (EPA). The procedures may involve preliminary sample treatment by acid digestion, using either hotplate or microwave oven, or filtration (EPA Method 3005A). Instrumental analysis is by atomic absorption/emission spectrophotometry (EPA Method 7000 series), inductively coupled plasma - optical emission spectrophotometry (EPA Method 6010B), and/or inductively coupled plasma - mass spectrometry (EPA Method 6020).

Recommended Holding Time:

Sample: 6 months Reference: EPA

For more detail see: ALS "Collection & Sampling Guide"

Metals in Seawater

This analysis is carried out using procedures adapted from "Recommended Guidelines for Measuring Metals in Puget Sound Marine Water, Sediment, and Tissue Samples" prepared for the United States Environmental Protection Agency and the Puget Sound Water Quality Authority, 1995. The procedures may involve preliminary sample treatment by acid digestion or filtration (EPA Method 3005A). Instrumental analysis of the seawater is by atomic absorption/emission spectrophotometry (EPA Method 7000 series), inductively coupled plasma - optical emission spectrophotometry (EPA Method 6010B), and/or inductively coupled plasma - mass spectrometry (EPA Method 6020).

Recommended Holding Time:

Sample: 6 months Reference: Puget

For more detail see ALS Environmental "Collection & Sampling Guide"

Trace Metals in Seawater by SPR-IDA Chelation

This analysis is carried out using procedures adapted from "Recommended Guidelines for Measuring Metals in Puget Sound Marine Water, Sediment, and Tissue Samples" prepared for the United States Environmental Protection Agency and the Puget Sound Water Quality Authority, 1995, and with procedures adapted from Cetac Technologies Incorporated. A suspended particulate resin (SPR), consisting of immobilized iminodiacetate (IDA) on a divinylbenzene polymer, is used to chelate and preconcentrate metals in seawater. Instrumental analysis is by inductively coupled plasma mass spectrometry (ICPMS) and/or routine atomic absorption spectrophotometry techniques (EPA 7000 series).

Appendix 1 - METHODOLOGY - Continued

Recommended Holding Time:

Sample/Extract: 6 months Reference: Puget

For more detail see ALS Environmental "Collection & Sampling Guide"

Rhodamine WT in Water

This analysis is carried out using a Waters Model 470 Scanning Fluorescence Detector. Prior to analysis, samples are warmed to room temperature and then an aliquot of the sample is filtered through a 0.45 um cellulose acetate filter to remove some of the potential interferences. After filtration, a portion of the sample, typically 500 uL, is injected into the fluorescence detector. An excitation wavelength of 556 nm is used and fluorescence is measured at an emission wavelength of 580 nm.

Please note the following: This method is non-selective and subject to many interferences. Typical interferences may include; pH, salinity, temperature, suspended matter, photochemical degradation of Rhodamine WT, and some types of algae pigments. Detection for this analysis is not chromatographic; therefore, anything present in the sample that fluoresces at the same wavelength as Rhodamine WT will be detected and reported as Rhodamine WT. Results reported using this method should be considered semi-quantitative in nature.

Recommended Holding Time:

Sample: None Available Reference: None Available

For more detail see ALS Environmental "Collection & Sampling Guide"

Results contained within this report relate only to the samples as submitted.

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End of Report



195 Pemberton Avenue North Vancouver, BC Canada V7P 2R4 Fax: 604-662-8548 Tel: 604-986-4331

Info@evsenvironment.com www.evsenvironment.com

FAX TRANSMITTAL SHEET

TO:	Cheryl Mackintosh	DATE:	August 9, 2004
	Azimuth Consulting Group	PROJECT No.:	09-0302-54
	218 – 2902 West Broadway	W.O. No.:	0400342,348
	Vancouver, BC	FAX No.:	604-739-9070
	V6K 2G8	TEL No.:	604-730-1220
SENT BY:	Rachel DeWynter	# PAGES (incl. o	cover): One (1)

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Re: Interim data for the 96-h rainbow trout and 48-h Daphnia magna LC50 toxicity tests performed on the sample identified as G-Creek acute 270704 (collected July 27, 2004).

Sample ID	Sample Collection	LC50 (95% Confidence Limits) [%vol/vol]		
	Date	96-h Rainbow Trout	48-h Daphnia magna	
G-Creek acute 270704	July 27, 2004 (09:30h)	> 100	> 100	

Please note that these are draft results and are subject to a QA/QC review. A complete report will follow by mail. Should you have any questions, please contact Edmund Canaria or myself at 604-986-4331.

Thank you,

Jennifer Young, B.Sc. Bioassay Test Supervisor – Cladoceran Team iyoung@evsenvironment.com

RSD