



QUALITY ASSURANCE (QA)
AND QUALITY CONTROL (QC) PLAN
FOR THE COLLECTION OF EFFLUENT SAMPLES
AT THE
CAM-2 (GLADMAN POINT)
DEW LINE SITE

Revised September 2005

Prepared by:
ENVIRONMENTAL SCIENCES GROUP
for
DEFENCE CONSTRUCTION CANADA
&
UMA ENGINEERING LTD.



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1. Introduction

During the clean-up of the CAM-2 DEW Line site at Gladman Point, Nunavut the collection of sewage effluent samples was undertaken as required by the Water Use License. The collection of wastewater samples is similar to the collection of other types of water samples.

As stated in the Water Use License, the contents of the Sewage Disposal Facility shall be analyzed once at 30 days after its establishment and a second time prior to discharge. All waste discharged from this Facility will be analyzed for the following parameters:

- Mineral oil and grease (observations of presence/absence of sheen will be made – no samples will be collected or analyzed);
- Total suspended solids (TSS);
- Biological oxygen demand (BOD₅);
- Faecal coliforms; and
- pH.

2. Sample Collection

2.1. Location

GPS coordinates of sample locations are collected and recorded. Photographs of the sample location are also taken.

2.2. Sampling Equipment

The following table summarizes the equipment and storage requirements for each water sample type collected. New bottles were used in all cases for the collection of the water samples.

Contaminant	Container	Amount	Rinse	Storage	Special Treatment
TSS, pH	1L Plastic Bottle	Full	No	Cool	Do not filter
BOD ₅	250 mL amber glass bottle	Full – no headspace	No	Cool	Do not filter
Bacteria and coliforms	Bacti bottles (Accutest)	Full	No	Cool	Analyze within 48 hours of collection



2.3. Sampling Methods

Sample bottles will be filled completely at the time of sampling. Bottles are not to be filled progressively over the course of days. If there is not sufficient water to completely fill the bottle(s), then no water sample will be collected. The bottles are to be filled with no headspace remaining to guard against volatilization of dissolved phases. Generally, the samples will be collected immediately prior to departure from the site and submitted for analysis within 48 hours.

3. Sample Handling

3.1. Preservation

The water samples will be kept cool (approximately 4⁰ C) prior to and during shipping. In general, water samples will be collected when transportation from the site will be available almost immediately after, as many types of the required analyses should be performed as quickly as possible after collection.

Ideally, samples collected for inorganic analyses should be acidified in the field, at the time of collection. However, regulations concerning the transportation of dangerous goods make supplying concentrated nitric acid in the field difficult. Where samples cannot be acidified in the field, it will be requested that the samples are acidified immediately upon receipt in the lab, *prior* to decanting or sample extraction. When acidifying in the lab, the container will be rinsed with 35% HNO₃ and included with the sample.

Samples are not to be filtered at any time. If samples contain excessive sediment, the samples will simply be decanted in the southern laboratory (*following* acidification, for metal analyses) prior to analysis.

3.2. Sample Identification

Each water sample will be given a blind number that was the only number provided on the labels of samples submitted for analysis. This sample number corresponds to the number assigned to that specific sample location which will be recorded on a map and in the field notebook.



3.3. Transportation

Samples are to be shipped by guaranteed airfreight in coolers from the site to their respective accredited laboratory for analysis. Chain-of-custody forms will be filled out and checked for each sample before shipment from the North, and the contents of shipments will be verified upon receipt in the laboratory.

4. Lab Analysis

4.1. Lab Accreditation

All laboratory analysis is carried out at accredited labs. The following laboratories are the ones primarily responsible for the analysis of water samples collected at CAM-2 (Gladman Point):

- 1) Analytical Services Unit, Queen's University, Kingston ON;
- 2) Analytical Sciences Group, Royal Military College of Canada, Kingston ON;
- 3) Taiga Laboratories, Yellowknife NWT.

Proof of accreditation from these laboratories is located in Appendix A. The standard methods used by the laboratories for each of these analyses are listed in the laboratory's scope of accreditation.

4.2. Detection Limits

The following table provides a summary of the detection limits for the analysis to be performed on water samples collected at CAM-2 (Gladman Point).

Parameter	Detection Limit
Biological oxygen demand (BOD ₅)	3 mg/L
Total suspended solids (TSS)	1 mg/L
Faecal coliforms	0 counts/100 mL

4.3. Methodology



The following is a summary of the methods to be used in the analysis of the water samples collected from CAM-2 (Gladman Point).

4.3.1. *Total Suspended Solids*

Analyses were conducted by the Analytical Services Group, Royal Military College, Kingston, Ontario. Each sample was clearly labelled and stored at low temperatures in a secured area before and after analysis.

Total suspended solids (TSS) in water were determined by filtering a sample through a 0.45-micron glass fibre filter. The filter was first dried in an oven at 105 °C for 4 hours, allowed to cool, and weighed. A measured volume of water (usually 500 mL) was poured through the filter, which was then oven-dried for 12-18 hours, cooled, and reweighed. The TSS were reported as the weight of suspended material divided by the volume of water (units of mg/L).

4.3.2. *Coliforms in Water*

Analyses were conducted by Taiga Laboratories in Yellowknife NT.

The coliform group comprises many facultative anaerobic, gram-negative, non-spore forming, rod-shaped bacteria. Total coliforms (TC), fecal coliforms (FC), and the specific coliform, *E. coli* (EC), were determined according to standard methods of the American Public Health Association,¹ using variations of the Membrane Filter Technique.²

Dilutions of the original sample were filtered through a 0.45 µ filter. Each filter was placed on an agar plate containing an indicator medium specific for the organisms being assayed. For total coliform (TC),³ LES Endo Agar was used. This indicator medium contains lactose, which is fermented by all coliforms, producing aldehydes to form red colonies with a metallic sheen. Agar plates were incubated 24 hours at 35 °C +/- 0.5 °C. For fecal coliform (FC), detecting only those coliforms from the feces of warm-blooded animals,⁴ the lactose-enriched medium, M-FC, which contains an aniline blue indicator, was used. Fecal coliforms produce blue colonies after 24 hours incubation at 44.5 °C +/- 0.2 °C. Note the difference in incubation temperature between total and fecal coliforms.

¹ American Public Health Association, Washington, DC. 1995. Published by the American Water Works Association (AWWA). Standard Methods for the Examination of Water and Wastewater. 19th Edition.

² Standard Methods for the Examination of Water and Wastewater, 18th Edition. 1992. Method # 9222 Membrane Filter Technique for Members of the Coliform Group.

³ Standard Methods 9222:B

⁴ Standard Methods 9222:D



To distinguish the coliform species, *E. coli* (EC), the indicator medium, mColiBlue24 (Millipore⁵), was used. Only *E. coli* forms blue colonies on this medium.

4.3.3. *Biological Oxygen Demand (BOD)*

Analyses were conducted by Taiga Laboratories in Yellowknife NT.

The Biological Oxygen Demand 5-day (BOD5) test was used.⁶ An airtight bottle filled to overflowing with sample was incubated for 5 days at 20 °C +/-1 °C. Dissolved oxygen (DO) was measured initially and after incubation, and BOD was calculated as the difference between initial and final DO (mg oxygen/L). The test measures the molecular oxygen used for the biochemical degradation of organic material, and the oxygen used to oxidize inorganic material such as sulphides and ferrous iron, over the incubation period.

4.3.4. *pH Measurement*

Measurements on water and soil samples were conducted by the Analytical Services Group, Royal Military College, Kingston, Ontario.

Water samples were measured directly using a 50-mL sample; the pH of soil was determined by mixing 10 g of soil with 10 mL of distilled water, allowing the mixture to settle, and measuring the pH of the supernatant. The pH was measured with a Denver Instruments model 220/300729.1 pH meter and probe, relative to buffered reference standards and control.

4.4. Reporting Requirements

The following types of QA/QC samples will also be collected as part of the water sampling program. Note that if more than one type of bottle is used for each water sample, QA/QC samples will be submitted in each type of bottle used for the collection of the samples.

Field duplicates: Approximately 10% of the samples were collected as field duplicates. That is, two samples were collected from one sample location. These samples were handled in the same way and submitted blindly to the laboratories for analysis.

⁵ Contact Millipore for more information.

⁶ Standard Methods 5210:B



Field blanks: Field blanks consisted of distilled water and were collected to ensure that there is no corruption of samples from the sampling method. The distilled water was poured from its container into the sample container at the same time and using the same techniques as used to collect the regular water samples.

Travel blanks: The purpose of travel blanks is to ensure that there is no corruption of the sample or sample container during travel. Ideally, a full set of travel blanks should accompany each shipment of water samples. However, in cases where very few samples are shipped at a time, this guideline can be extended to a more reasonable number. Travel blanks were filled at ESG prior to leaving for the field. They were shipped with the sample bottles, stored with the sample bottles on site, brought out to the sampling location in the field, returned to the lab, and shipped to the labs with the water samples. They should not be opened unless the other bottles or water samples are opened for some reason during shipping.



CAM-2 QA/QC Plan
Revised September 2005

Appendix A: Proof of Laboratory Accreditation



CAEAL Directory of Laboratories

Login	
Member:	
Password:	
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Membership Number: 2709

Laboratory Name: **Queen's Analytical Services Unit**

Parent Institution: Queen's University

Address: Environmental Studies Biosciences Complex Kingston, Ontario K7L 3N6

Contact: Dr. Allison Rutter

Phone: (613) 533-2642

Fax: (613) 533-2897

Email: rutterra@biology.queensu.ca

Standard: Conforms with requirements of CAN-P-4D (ISO/IEC 17025)

Clients Served:

Revised On: February 21, 2005

Valid To: February 16, 2008

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Showing Accreditation Directory

Scope of Accreditation

Search Criteria - results highlighted in yellow
Laboratory Name contains "analytical services unit"

Oil

PCB - Oil (004)

ASU 04; based on EPA 8081

GC/ECD - EXTRACTION

Total PCB

Soil/Sediment

Metals - Soil (007)

ASU007; based on EPA 200.7

ICP/OES - DIGESTION

Arsenic

Cadmium

Chromium

Cobalt

Copper

Lead

Nickel

Zinc

Water (Inorganic)

Ammonia - Water (009)

ASU09; based on TECHNICON METHOD

AUTO COLOR

Ammonia

Water (Inorganic)

Dissolved Metals - Water (008)

ASU08; based on EPA 200.7

ICP/AES

Dissolved Arsenic
Dissolved Cadmium
Dissolved Chromium (High)
Dissolved Cobalt (High)
Dissolved Copper (High)
Dissolved Lead (High)
Dissolved Manganese (High)
Dissolved Nickel (High)
Dissolved Zinc (High)

Water (Inorganic)

Major Ions - Water (003)

ASU 03; based on DIONEX MANUAL
ION CHROMATOGRAPHY
Chloride
Nitrate
Nitrate plus Nitrite
Sulfate

Water (Inorganic)

Oil and Grease - Water (010)

ASU10; based on SM 5520 (20TH EDITION)
GRAVIMETRIC - EXTRACTION
Oil and Grease

Water (Inorganic)

Total Metals - Water (012)

ASU08; based on EPA 200.7
ICP/AES
Total Arsenic
Total Cadmium
Total Chromium
Total Cobalt
Total Copper
Total Lead
Total Manganese
Total Nickel
Total Zinc

Water (Organic)

Biphenyl - Water (005)

ASU 05; IN HOUSE METHOD
GC/FID - EXTRACTION
Biphenyl
Biphenyl Ether

Water (Organic)

Phenols - Water (001)

ASU 01; based on MOE METHOD 1983
AUTOANALYZER/COLORIMETRY
Phenols



CAEAL Directory of Laboratories

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**Membership
Number:** 2965

Laboratory Name: **RMC Analytical Services Group**

Parent Institution: The Royal Military College of Canada

Address: Dept. of Chemistry & Chemical Engineering RMC, PO Box 17000 Stn. Forces Kingston, Ontario K7K 7B4

Contact: Dr. David Kelly

Phone: (613) 541-6000 Ext. 6921

Fax: (613) 545-8341

Email: david.kelly@rmc.ca

Standard: Conforms with requirements of CAN-P-4D (ISO/IEC 17025)

Clients Served: Specified Clients

Revised On: January 03, 2005

Valid To: January 03, 2008

[Click for PT Directory](#)

Showing Accreditation Directory

Scope of Accreditation

Search Criteria - results highlighted in yellow
Laboratory Name contains "analytical services group"

Oil

Total Chlorine - Oil (003)

ASG 003; based on J. RADIONAL CHEM., 50, 229-234 (1979) AND based on ANAL. CHIM. ACTA., 108, 137-147 (1979).

NAA

Total Chlorine

Plant Tissue

Polychlorinated Biphenyls (PCB) - Plants (020)

ASG026; based on EPA 8082, EPA 680, EPA 8082, EPA 3545, EPA 3640C, FRAME, ET. AL. J. HIGH RESOL. CHROMATOGR., 19:657-668, 1996.

Arochlors

PCB Conegeners

Radio Chemistry

Radionuclide Activity - Biota (008)

ASG 031; based on SM AMERICAN WATER ASSN. METHOD 7120, SM FOR EXAM. OF WATER/WASTEWATER, 20TH ED., & USEPA METHOD 901.1.

GAMMA SPECTROSCOPY

Barium-140

Cerium-144

Cesium-134

Cesium-136

Cesium-137

Cesium-138

Cobalt-60
 Iodine-131
 Iodine-132
 Iodine-133
 Iodine-134
 Iodine-135
 Lanthanum-140
 Molybdenum-99
 Niobium-95
 Rubidium-86
 Rubidium-88
 Ruthenium-103
 Ruthenium/Rhodium-106
 Strontium-91
 Tellurium-129m
 Tellurium-131m
 Tellurium-132
 Yttrium-90m
 Yttrium-91m
 Zirconium-95

Soil

BTEX/Petroleum Hydrocarbons (PHC) - Soil (025)

ASG 053; based on CCME REFERENCE METHOD FOR THE CANADA-WIDE STANDARD FOR PETROLEUM HYDROCARBONS IN SOIL - TIER 1 METHOD 2001

GC/MS - EXTRACTION

Benzene
 Ethylbenzene
 F1: C6-C10
 F2: C10-C16
 F3: C16-C34
 F4: C34-C50
 m/p-xylene
 o-xylene
 Toluene

Soil/Sediment

Activity of Radionuclide - Soil (007)

ASG 030; based on SM AMERICAN WATER ASSN. METHOD 7120, SM FOR EXAM. OF WATER/WASTEWATER, 20TH ED., & USEPA METHOD 901.1.

GAMMA SPECTROSCOPY

Barium-140
 Cerium-144
 Cesium-134
 Cesium-136
 Cesium-137
 Cesium-138
 Iodine-131
 Iodine-132
 Iodine-133
 Iodine-134
 Iodine-135
 Lanthanum-140
 Molybdenum-99
 Niobium-95
 Rubidium-86
 Rubidium-88
 Ruthenium-103
 Ruthenium/Rhodium-106
 Strontium-91
 Tellurium-129m
 Tellurium-131m
 Tellurium-132
 Yttrium-90m
 Yttrium-91m
 Zirconium-95

Soil/Sediment

PCBs - Soil (005)

ASG 005; based on EPA 8081

GC/ECD - SOXHLET/PSE

PCBs

Soil/Sediment

Polycyclic Aromatic Hydrocarbons (PAH) - Soil (001)

ASG 002; based on EPA 8100

GC/MS - EXTRACTION
 Acenaphthene
 Acenaphthylene
 Anthracene
 Benzo (a) anthracene
 Benzo (a) pyrene
 Benzo (b) fluoranthene
 Benzo (g,h,i) perylene
 Benzo (k) fluoranthene
 Chrysene
 Dibenzo (a,h) anthracene
 Fluoranthene
 Fluorene
 Indeno (1,2,3 - cd) pyrene
 Naphthalene
 Phenanthrene
 Pyrene

Soil/Sediment

Total Petroleum Hydrocarbons (TPH) - Soil (010)

ASG010; based on EPA 3550B, EPA 8015C

GC/FID - EXTRACTION
 Total Petroleum Hydrocarbons

Water (Inorganic)

Alkalinity - Water (Drinking/Surface/Sewage/Ground) (013)

ASG035; based on NAQUADAT NO. 10101, EPA 310.1, SM 20TH ED. 2320

POTENTIOMETRIC
 Alkalinity (pH 4.5)

Water (Inorganic)

Biochemical Oxygen Demand (BOD) - Water (Surface/Sewage/Ground) (019)

ASG042; based on NAQUADAT NO. 08201, EPA 405.1, SM 20TH ED. 5210

D.O. METER
 BOD (5 day)

Water (Inorganic)

Conductivity - Water (Drinking/Surface/Sewage/Ground) (016)

ASG038; based on SM 20TH. ED. 2510, EPA 120.1

CONDUCTIVITY METER
 Conductivity (25Å°C)

Water (Inorganic)

Mercury - Water (Ground/Surface/Drinking) (011)

ASG021; based on EPA 7470A

FLOW INJECTION MERCURY SPEC
 Mercury

Water (Inorganic)

Metals - Water (024)

ASG 049; based on EPA 200.8 AND SM 20TH ED. 3125

ICP/MS
 Dissolved Aluminum
 Dissolved Beryllium
 Dissolved Boron
 Dissolved Cadmium
 Dissolved Chromium
 Dissolved Cobalt
 Dissolved Copper
 Dissolved Lead
 Dissolved Manganese
 Dissolved Molybdenum
 Dissolved Nickel
 Dissolved Silver
 Dissolved Strontium
 Dissolved Thallium
 Dissolved Tin
 Dissolved Uranium
 Dissolved Vanadium Parameter suspended on 5/18/2005
 Total Antimony
 Total Arsenic

Water (Inorganic)

pH - Water (Drinking/Surface/Sewage/Ground) (015)

ASG037; based on SM 20TH. ED. 4500-H+, EPA 150.1

pH METER

pH

Water (Inorganic)

Total Dissolved Solids (TDS) - Water (Drinking/Surface/Sewage/Ground) (018)

ASG040; based on EPA 160.1, SM 18TH ED. 2540C

GRAVIMETRIC

Total Dissolved Solids

Water (Inorganic)

Total Suspended Solids (TSS) - Water (Drinking/Surface/Sewage/Ground) (017)

ASG039; based on EPA 160.2, SM 18TH ED. 2540D

GRAVIMETRIC

Total Suspended Solids

Water (Microbiology)

Coliforms - Water (Drinking/Surface/Sewage/Ground) (014)

ASG036; based on MOE MICROMEFD-EC3407, SM 20TH ED. 9225

MEMBRANE FILTRATION (DC)

Escherichia coli (E. coli)

Total Coliforms

Water (Microbiology)

Fecal Coliforms - Water (Drinking/Surface/Sewage/Ground) (022)

ASG044; based on MOE MICROMEFD-EC3407, SM 20TH ED. 9222 D

MEMBRANE FILTRATION (m FC)

Fecal Coliforms

Water (Microbiology)

Heterotrophic Plate Count (HPC) - Water (Drinking/Surface/Sewage/Ground) (023)

ASG041; SM 20TH ED. 9215 D

MEMBRANE FILTRATION

Heterotrophic Plate Count (HPC)

Water (Organic)

Total PCB - Water (Surface/Sewage/Ground) (009)

ASG015, ASG022; based on EPA 8082, EPA 617, FRAME, ET. AL. J. HIGH RESOL. CHROMATOGR., 19: 657-668, 1996

GC/MS - EXTRACTION

Total PCB

Water (Organic)

Total PCB - Water (021)

ASG006, ASG008; based on EPA 8082, EPA 617, FRAME, ET. AL. J. HIGH RESOL. CHROMATOGR., 19: 657-668, 1996.

GC/ECD - EXTRACTION

Total PCB

Water (Organic)

Volatile Organic Compounds (VOC) - Water (Drinking/Surface/Ground) (012)

ASG023; based on EPA 624, EPA 8260B

GC/MS - PURGE AND TRAP

1,1-Dichloroethane

1,1-dichloroethylene

1,1-Dichloropropene

1,1,1-Trichloroethane

1,1,1,2-Tetrachloroethane

1,1,2-Trichloroethane

1,1,2,2-Tetrachloroethane

1,2-Dibromo-3-chloropropane

1,2-Dibromoethane

1,2-dichlorobenzene

1,2-dichloroethane

1,2-Dichloropropane

1,2,3-Trichlorobenzene

1,2,3-Trichloropropane

1,2,4-Trichlorobenzene

1,2,4-Trimethylbenzene

1,3-Dichlorobenzene

1,3-Dichloropropane

1,3,5-Trimethylbenzene

1,4-dichlorobenzene

2-Chlorotoluene

2,2-Dichloropropane

4-Chlorotoluene

Benzene

Bromobenzene

Bromochloromethane

Bromodichloromethane

Bromoform

Bromomethane
 Carbon Tetrachloride
 Chlorobenzene
 Chlorodibromomethane
 Chloroethane
 Chloroform
 Chloromethane
 cis-1,2-Dichloroethylene
 cis-1,3-Dichloropropene
 Dibromomethane
 Dichlorodifluoromethane
 Dichloromethane Parameter suspended on 8/17/2005
 Ethylbenzene
 Hexachlorobutadiene
 Isopropylbenzene
 Isopropyltoluene
 m/p-xylene
 n-Butylbenzene
 n-Propylbenzene
 Naphthalene
 o-xylene
 sec-Butylbenzene
 Styrene
 tert-Butylbenzene
 Tetrachloroethylene
 Toluene
 trans-1,2-Dichloroethylene
 trans-1,3-Dichloropropene
 Trichloroethylene
 Trichlorofluoromethane
 Vinyl Chloride

Water (Radiochemistry)

Alpha/Beta Radiation (Swab) (004)

ASG 004; RMC-CMR LSC PROCEDURES MANUAL VER. 1.0

LIQUID SCINTILLATION COUNTING

Alpha radiation

Beta radiation

Water (Radiochemistry)

Radionuclide (Activity) - Water (006)

ASG 024; based on SM AMERICAN WATER ASSN. METHOD 7120, SM FOR EXAM. OF WATER/WASTEWATER, 20TH ED., & USEPA METHOD 901.1.

GAMMA SPECTROSCOPY

Barium-140

Cerium-144

Cesium-134

Cesium-136

Cesium-137

Cesium-138

Cobalt-60

Iodine-131

Iodine-132

Iodine-133

Iodine-134

Iodine-135

Lanthanum-140

Molybdenum-99

Niobium-95

Rubidium-86

Rubidium-88

Ruthenium-103

Ruthenium/Rhodium-106

Strontium-91

Tellurium-129m

Tellurium-131m

Tellurium-132

Yttrium-90m

Yttrium-91m

Zirconium-95

SCOPE OF ACCREDITATION

The Royal Military College of Canada
RMC ANALYTICAL SERVICES GROUP
Department of Chemistry and Chemical Engineering P.O. Box 17000, Station Forces
Kingston, ON
K7K 7B4

Accredited Laboratory No. 276
(Conforms with requirements of CAN-P-4D (ISO/IEC 17025), and CAN-P-1598)

CONTACT: Dr. David Kelly
TEL: (613) 541-6000
x6921
FAX: (613) 545-8341
EMAIL: david.kelly@rmc.ca

CLIENTS SERVED: All interested parties

FIELDS OF TESTING: Chemical/Physical

PROGRAM SPECIALTY Environmental
AREA:

ISSUED ON: 2005-06-22

VALID TO: 2006-11-02

ENVIRONMENTAL AND OCCUPATIONAL HEALTH AND SAFETY

Environmental

Water (Microbiology)

(Coliforms – Water [014]) (OSDWA)

ASG036; based on MOE
MICROMEFDCE3407, SM 20TH ED. 9225

MEMBRANE FILTRATION (DC)
Escherichia coli (E. coli)
Total Coliforms

(Fecal Coliforms – Water [022]) (OSDWA)

ASG044; based on MOE
MICROMEFDC-E3407, SM 20TH ED. 9222
D

MEMBRANE FILTRATION (m FC)
Fecal Coliforms

(Heterotrophic Plate Count (HPC) – Water [023]) (OSDWA)

ASG041; SM 20TH ED. 9215 D

MEMBRANE FILTRATION
Heterotrophic Plate Count (HPC)

Notes:

CAN-P-4D (ISO/IEC 17025): General Requirements for the Competence of Testing and Calibration Laboratories ISO/IEC 17025-1999)

CAN-P-1598: Guidelines for Accreditation of Environmental Testing Laboratories

OSDWA: Indicates the appendix is used for the analysis of Ontario drinking water samples, which is subject to the rules and related regulations under the Ontario "Safe Drinking Water Act" (2002)

P. Paladino, P. Eng., Director, Conformity Assessment

Date: 2005-06-22

SCC 1003-15/358
CAEAL #2965
Partner: CAEAL
MOE License No.: 2264