Appendix 48

2022 Wildlife and HHRA Country Foods Screening Level Risk Assessment Plan Version 8



MEADOWBANK COMPLEX

Wildlife and HHRA Country Foods Screening Level Risk Assessment Plan

In Accordance with NIRB Project Certificate No.004

Version 8 January, 2023

IMPLEMENTATION SCHEDULE

This Plan will be implemented immediately subject to any modifications proposed by the NIRB as a result of the review and approval process.

DISTRIBUTION LIST

Agnico Eagle – Environment and Critical Infrastructures Superintendent

Agnico Eagle - Environment General Supervisor

Agnico Eagle – Environmental Coordinator

Agnico Eagle – Environmental Technician

DOCUMENT CONTROL

Version	Date (YMD)	Section	Revision	
1	2016- 03-31	All	Comprehensive plan for Meadowbank Project	
2	2018- 06-01	2.5	ROC based on a commitment made during the Whale Tail Final Hearing through discussions with Environment and Climate Change Canada	
3	2019- 04-25	Throughout 3.4	Version 2 text expanded to describe assessment of risks to Semi-Palmated Sandpiper from exposure to contaminants in the TSF. Updated time-in-area for caribou from 33% to 12% based on collaring data.	
		2.4	Introduced screening of potential COPCs in soil and water samples using soil and water quality guidelines or baseline +10%, according to Golder, 2016/Agnico Eagle, 2019.	
4	2021-	3.2	Reduction of sampling effort from 5 samples per monitoring station to 3 samples	
4	04-12	3.4	dated time-in-area factors according to Golder, 2019.	
		Throughout	Added analysis of risk to all receptors of concern from consumption of tailings material.	
5	2021- 08-27	Appendix B	Updated SOP to reflect V4 sampling requirements (3 locations, additional tailings samples) and corrected sample site coordinates.	
		Section 2.1	Conceptual model added.	
		Section 2.1.3	Clarified pathway for shorebirds (TSF only). Previously this section erroneously suggested tundra pathways would also be assessed, which was not planned, as indicated in Section 3.4.	
6	2022- 04-11	Section 2.2	EDI approach updated to align with Golder (2019b) – adopted method of calculating EDI for assessment location only, rather than assessment location + external reference combined.	
	I Section 222	Mammoth Lake added as an assessment location for water quality, due to discharge of effluent in this location		
		Section 2.2.2.2	Dietary preferences – percentage of benthic invertebrates obtained from the TSF adjusted to 13% from 100%, based on results of 2021 field study (further described in Section 3.3)	

		Section 2.3	TRVs – removed allometric scaling factor for mammalian TRVs to align with Golder (2019a,b) methods, as recommended in Allard et al. (2010).
		Former Appendix A	Appendix A (example calculation) removed for brevity since the components are clearly presented in main text.
		Appendix A	Updated SOP with selected 2021 Whale Tail site UTM coordinates and tailings beach sediment sample collection (for consideration).
		Section 3 and throughout	HHRAcountry foods Assessment Plan added
7	2022-	Section 2.2.1	Minor correction to soil intake values to properly reflect diet % moisture value indicated.
/	07-14	Section 2.2.2.2	Proportion of shorebird diet obtained from TSF adjusted from 13% to 100% (more conservative) per ECCC recommendation.
8	2023- 01-25	Section 2.2.2.1 & Appendix A	Methods for collection and analysis of tailings sediment samples updated (collection of beach sediment rather than mill effluent)

Prepared By: Meadowbank Environment Department

Approved by: Eric Haley

Environment and Critical Infrastructures Superintendent Interim

TABLE OF CONTENTS

1	INTRO	DUCTION	1
1.1	Backgr	ound	1
1.2	Genera	al Approach	2
1.3	Locatio	on Description	5
1.4	Site Fa	ncilities	5
1.5	Source	es of Contaminants	5
2	WILDL	IFE ASSESSMENT	6
2.1	Problei	m Formulation	6
	2.1.1	Receptors of Concern (ROCs)	8
	2.1.2	Protection Goals and Endpoints	8
	2.1.3	Exposure Pathways	8
	2.1.4	Contaminants of Potential Concern (COPCs)	9
2.2	Exposi	ure Assessment	10
	2.2.1	I _{w,s,f} - Intake of Water, Soil/Sediment, and Food	11
	2.2.2	C _{w,s,f} - Dietary Concentrations of COPCs	12
	2.2.3	Biotransfer Factor	15
	2.2.4	Time in Area	15
2.3	Toxicity	y Assessment	16
2.4	Risk C	haracterization	17
2.5	Uncert	ainty Assessment	18
3	HUMA	N HEALTH – COUNTRY FOODS ASSESSMENT	19
3.1	Problei	m Formulation	19
	3.1.1	Exposure Pathways	21
	3.1.2	Potential Receptors	21
	3.1.3	Contaminants of Potential Concern	21
3.2	Exposi	ure Assessment	22
	3.2.1	C _F - COPC Concentrations in Country Food Items	23
	3.2.2	IR _F – Intake (Consumption) Rate of Food Items	24
	3.2.3	BW – Receptor Body Weight	27
3.3	Toxicity	y Assessment	27

3.4	Risk Ch	aracterization
	3.4.1	Non-Cancer Risks
	3.4.2	Incremental Lifetime Cancer Risk
3.5	Uncerta	inty Assessment
4	REPOR	TING AND ADAPTIVE MANAGEMENT28
REFE	RENCES	30
1	SOIL A	ND VEGETATIONI
1.1	Materia	li
1.2	Sample	Locationsi
1.3	Sample	Collection Procedurev
	1.3.1	Vegetation Sample Collectionvii
	1.3.2	Soil Sample Collectionvii
1.4	Tracking	g, Storage, Transportation and Laboratory Analysisvii
2	WATER	XIII
3	TAILIN	GS ANALYSISXIV
3.1	Benthic	Invertebrates xiv
3.2	Sedime	nt Chemistryxiv
3.3	Tailings	Water Chemistryxv
		APPENDICES
Appe	ndix A: Si	andard Operating Procedure for Field Sample Collection
		oxicity Reference Values, Bio-Transfer Factors, Tolerable Daily Intake Values
		LIST OF TABLES
Table	-	veight (BW), water intake (I _{water}), soil intake (I _{soil}), and wet and dry (I _{food} ; FI) food or the identified ROCs11
Table		cumulation factors (BAFs) from USEPA (1999) used for estimating whole-body trations of COPCs in benthic invertebrates

receptors of concern	
Table 4. Time-in-area assumption for each study area and ROC	. 16
Table 5. Food items, consumption scenarios, and estimated consumption rates. Toddler values are 43% of adult values (Richardson, 1997) unless otherwise indicated	. 26
LIST OF FIGURES	
Figure 1. Meadowbank site WSLRA sampling locations	3
Figure 2. Whale Tail site WSLRA assessment locations.	4
Figure 3. Wildlife risk assessment conceptual model	7
Figure 4. Human health (country foods) risk assessment conceptual model	. 20

INTRODUCTION

1.1 BACKGROUND

In 2006, Azimuth Consulting Group Inc. conducted a pre-construction wildlife screening level risk assessment (WSLRA) for the Meadowbank site to assess potential risks to wildlife via dietary uptake of mine-related contaminants (Azimuth, 2006). Specifically, the pre-construction SLRA focused on determining the contaminants of potential concern (COPCs) from predicted minesite activities, and evaluating potential risks to wildlife from exposure to contaminants under baseline conditions as well as predicted conditions during mine operation.

Under baseline conditions, negligible risks were found for all COPCs except chromium, which was determined to pose an improbable but potential risk for songbirds due to naturally elevated concentrations in the region. COPC exposure concentrations were not expected to increase during mine operation, so potential risks were not expected to change from baseline conditions.

In addition, a pre-construction risk assessment for consumption of country foods (HHRA_{country foods}) was completed (Wilson Scientific, 2006) to assess risk to human receptors associated with ingestion of various country foods under baseline and operational conditions. No incremental risks from consumption of country foods were predicted as a result of mine operations.

In 2016, Agnico Eagle submitted a Final Environmental Impact Statement (EIS) to NIRB for the Whale Tail Mine satellite deposit (Agnico Eagle, 2016). This FEIS included assessments of risk to wildlife and country foods consumers in the Whale Tail Mine area under baseline and operational conditions. Concentrations of contaminants in soil were not predicted to change or met screening criteria, so no residual impacts to soil and subsequently, vegetation and prey quality were predicted as a result of mine operation, and no quantitative risk characterization (hazard quotient calculation) was required. Results of an updated assessment conducted in support of the Whale Tail Mine Expansion Project in 2018 – 2019 (Golder, 2019a) indicated the same conclusion.

Further, in 2019, as part of the Whale Tail Mine Expansion Project permitting (Technical Meeting Commitment 45), Agnico Eagle submitted an updated assessment for exposure of wildlife receptors of concern to contaminants in ingested tailings water and sediment (Golder, 2019b). Results indicated that while screening values were exceeded for a number of parameters, risks were acceptable (HQ <1) for all receptors.

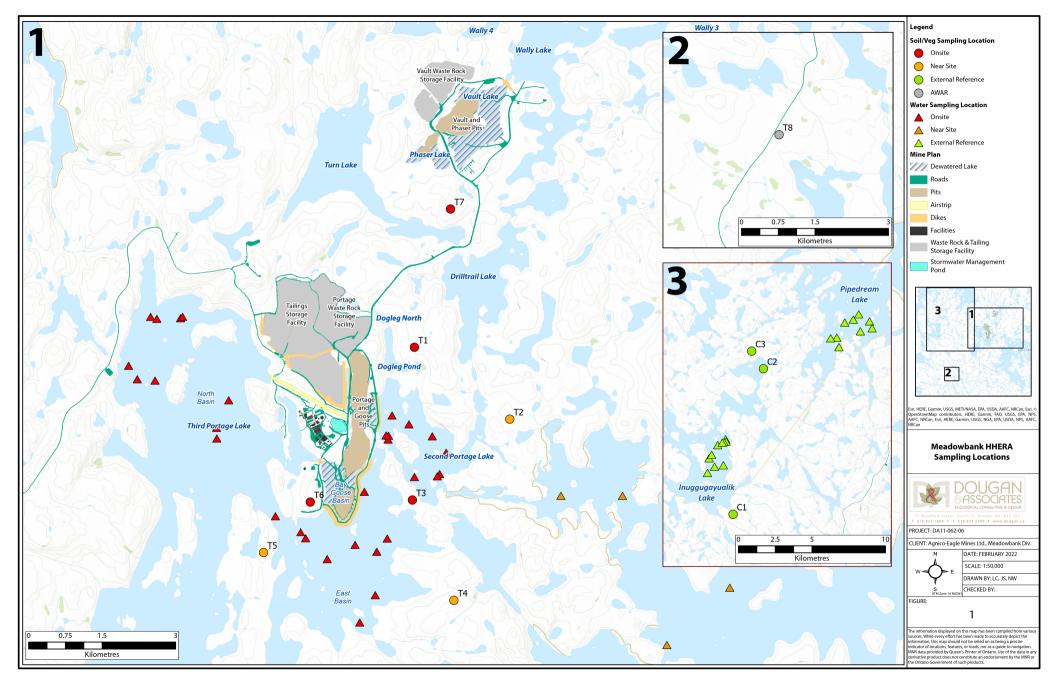
While no incremental risks to wildlife or country foods consumers from minesite operations has been predicted, this Wildlife and Country Foods Risk Assessment Plan presents the assessment approach and methodology that will continue to be used to assess risk to these receptors from chemical contaminants as a result of operations at the Meadowbank Complex. As required under the Meadowbank Complex Terrestrial Ecosystem Management Plan (TEMP) and Nunavut Impact Review Board Project Certificate No.004 - Condition 67, this WSLRA and HHRA_{country foods} is scheduled to be completed every 3 years during mine operation.

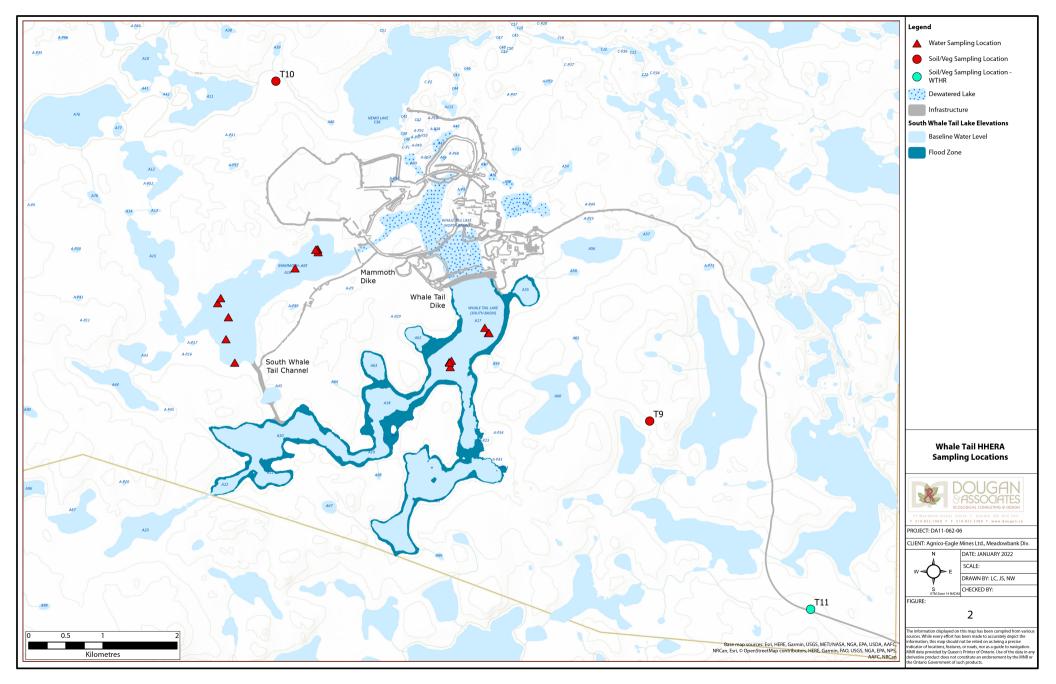
This plan is intended to guide the WSLRA and HHRA_{country foods} risk assessment process, and methods in each reporting year will be reviewed and adjusted as required, based on current practices at the time.

1.2 GENERAL APPROACH

The goal of the Meadowbank Complex WSLRA and HHRA_{country foods} is to determine potential risks to wildlife and general public consumers of country foods from ingestion of chemical contaminants under operational conditions. The general approach includes the common risk assessment components of problem formulation, exposure assessment, toxicity assessment and risk characterization. In particular, assessments will aim to distinguish risk due to operation of the mine from risk due to background conditions by collecting samples of environmental media from a variety of potentially impacted and reference locations in each assessment year (Figures 1 and 2; further described in Section 2.2.2).

Risk assessments will follow a hazard quotient approach, and are based on food-chain modeling developed for the baseline screening level risk assessments at the Meadowbank site (Azimuth, 2006; Wilson Scientific, 2006) and updated using methods from the Whale Tail site assessments (Golder 2019a,b) as described below. The risk assessment framework follows various standard Canadian and American sources (e.g. CCME, 1996; USEPA, 1992, 1998; Health Canada, 2012).





1.3 LOCATION DESCRIPTION

The main Meadowbank site is located 70 km north of the hamlet of Baker Lake, Nunavut, near the border of the Northern and Southern Arctic ecozones. The Whale Tail site is located a further 50 km north-west. Collectively these sites are referred to as the Meadowbank Complex. Terrain in the Meadowbank area is typical barren-ground subarctic, with low-growing vegetation in poorly developed soil with continuous permafrost. The landscape is dominated by many interconnected lakes and isolated ponds with indistinct drainage patterns. Topography consists of rolling hills, boulder fields and bedrock outcrops. The main mine site is located at the headwaters of the Quioch River system, which flows southeast through Chesterfield Inlet into Hudson Bay. Lakes in this region are ultra-oligotrophic, with low productivity levels. This region supports few terrestrial mammals (15 species) and birds (62 species) (Azimuth, 2006). Migratory species (primarily caribou and Canada geese) are present.

1.4 SITE FACILITIES

The Meadowbank Complex consists of several gold-bearing deposits, primarily mined as open pits (Portage, Goose, Vault, Phaser, Whale Tail, and IVR Pits). Much of the original infrastructure is located in close proximity to the Meadowbank mill, with the exception of the Vault Pit which is approximately 10 km northeast of the main site. The Whale Tail Mine, which was permitted in 2018, is approximately 50 km northwest of the site. Operations at the Portage, Vault and Phaser Pits ceased in 2019, and simultaneously operations at the Whale Tail Mine began.

Waste rock from the pits is stored in the Portage Waste Rock Storage Facility, Vault Waste Rock Storage Facility, Whale Tail and IVR Waste Rock Storage Facilities (RSFs). Rock Storage Facilities are constructed to minimize the disturbed area and will be capped with a layer of non-potentially acid-generating rock (NPAG). During the construction period, NPAG is also used for construction of dikes and roads. Mined ore is either processed in the mill or stockpiled for eventual processing.

Tailings are stored in the Tailings Storage Facility (TSF) at the main minesite. The TSF is defined by the series of dikes built around and across the basin of the dewatered northwest arm of Second Portage Lake. In-pit disposal is also occurring in Portage and Goose Pits since 2019. Tailings water is reclaimed for use in ore processing.

An onsite airstrip supports transportation of goods and personnel to and from the Meadowbank site by jet. A 110-km All Weather Access Road (AWAR) runs between the main minesite and the hamlet of Baker Lake, where Agnico Eagle maintains a bulk fuel storage and barge facility. The Vault Pit is connected to the main minesite by a 10-km haul road, and the Whale Tail site is connected by a 62-km haul road.

1.5 SOURCES OF CONTAMINANTS

Major mine site operations and their potential to contribute to COPCs (based on Azimuth, 2006 and Golder, 2019a) are summarized here.

Open pits – Along with ore, pits produce waste rock, which may contribute to COPCs through dust emissions.

Rock storage facilities – Waste rock (not containing ore) is moved to these areas. Dust may be blown from the rock piles during dumping and vehicle traffic during transport of material. Seepage from rock storage facilities is controlled in sumps and pumped back to attenuation ponds or the TSF.

Borrow pits and quarries – Borrow pits and quarries are used as necessary for the construction of mine site roads and road maintenance. The COPCs for borrow pits and quarries are similar to open pits.

Tailings Storage Facilities (TSF) – The northwest arm of Second Portage Lake was partitioned off by the East Dike and de-watered from 2009 to 2012. The northwestern portion of this area was further partitioned by the Stormwater Dike to create the North and South Cell TSF. Although permafrost is expected to freeze the tailings, the material is fine-grained and could be a source of dust emissions during dry periods. Tailings are also disposed in Portage and Goose Pit.

Roads and airstrip – Frequently used gravel haul roads run throughout the mine site to connect pits, waste rock storage and processing facilities. An airstrip, receiving approximately five planes per week, was built at the mine site to receive deliveries and personnel. Dust from these sources could be a potential source of contaminants. A 110 km long all weather access road (AWAR) was constructed between the mine and the Hamlet of Baker Lake, using gravel from quarries along the road. Whale Tail Mine satellite deposit is connected by a 62-km haul road to the Meadowbank Site

Effluent discharge – Dewatering of lakes for pit development or TSF construction is considered effluent discharge and is regulated under the current NWB Water License and MDMER. Lake water is treated for suspended solids removal before discharge, if needed, and since it is an existing surface water source, it is not likely to be a source of contaminants in the receiving water. Effluent is also periodically discharged from attenuation ponds into adjacent lakes, under NWB Water License and MDMER requirements. As a result, metals regulated under MDMER are evaluated for potential as COPCs.

Diesel generating plant, mine mill plant and associated facilities — Diesel generating plants provide power for the mine. The Air Quality Impact Assessment (2005) determined emission of PAHs was "very low" and did not require modeling. The milling of rock in the processing plant takes place under wet conditions, and is not a source of particulate emissions. All health and safety-related requirements to reduce particulate emissions during handling of the ore at the mine plant before processing are met, so these are not expected to be a significant source of contaminants.

Overall, roads, waste rock and tailings were determined to be the main sources potentially contributing to COPCs through dust emissions. Dewatering effluent discharge may potentially contribute to COPCs in water sources, along with physical changes to water bodies for the Whale Tail site (flooding). In addition, risks to wildlife from exposure to contaminants within the tailings storage facility are now considered, following discussions with Environment and Climate Change Canada during the Final Hearing for the Whale Tail Mine project (2017).

WILDLIFE ASSESSMENT

1.6 PROBLEM FORMULATION

The WSLRA evaluates risks to wildlife receptors of concern (ROCs) from ingestion of COPCs measured in environmental media for specified exposure pathways. Receptors, protection goals, exposure pathways, and methods for identification of COPCs are summarized below, and the conceptual model is shown in Figure 3.

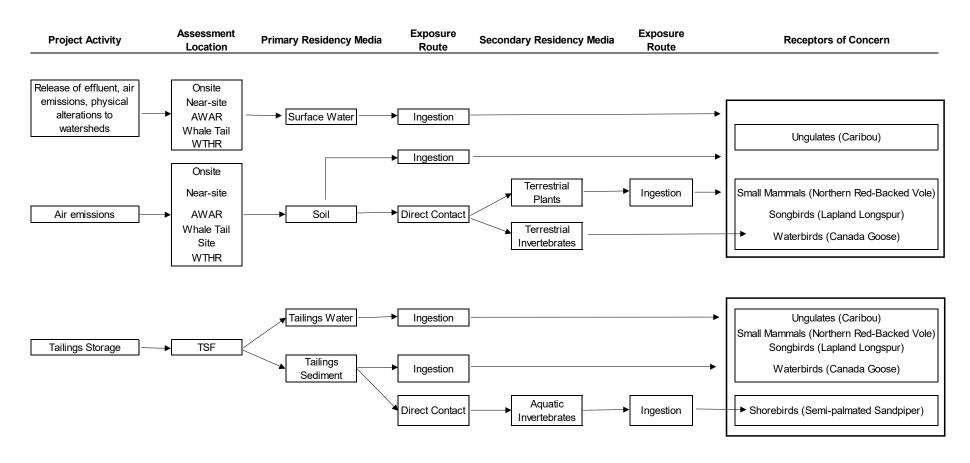


Figure 3. Wildlife risk assessment conceptual model.

1.6.1 Receptors of Concern (ROCs)

The original Meadowbank WSLRA (Azimuth, 2006) considered four groups of ROCs: ungulates, small mammals, waterfowl and songbirds. These choices were determined from the project's initial FEIS (Cumberland, 2005), which included discussions with stakeholders, public meetings, traditional knowledge and experience from other mines. Specifically, the WSLRA focussed on Caribou, Canada Goose, Lapland Longspur and Northern Red-Backed Vole as representative species. An ecological description of the area and detailed descriptions of the biology of each of these receptors can be found in Azimuth (2006). This updated assessment framework also includes an assessment of risks to all original receptors, plus shorebirds (as represented by Semi-Palmated Sandpiper) from contaminants within the TSF, based on a commitment made during the Whale Tail Mine project Final Hearing, following discussions with Environment and Climate Change Canada (ECCC) (Technical Meeting Commitment 45, as described in Golder, 2019b.

1.6.2 Protection Goals and Endpoints

Since the ROCs identified are not rare or endangered species, protection at the population level was determined to be appropriate (Azimuth, 2006). The assessment endpoint is no adverse effect of COPCs on populations of Caribou, Canada Goose, Lapland Longspur, Northern Red-Backed Vole, and Semi-Palmated Sandpiper.

As a result, ecotoxicological benchmarks used in the risk characterization will be lowest observable adverse effect levels (LOAELs), which are generally considered to be appropriate for determining risk at the population level (Azimuth, 2006). Toxicity reference value (TRV) selection is further described in Section 2.3.

1.6.3 Exposure Pathways

The following exposure pathways will be investigated, as shown in Figure 3. The term "tundra pathways" is used throughout to refer to evaluations conducted for samples collected at onsite, near-site, AWAR, Whale Tail Site and WTHR locations (Figures 1 and 2), as opposed to the "TSF pathway" which assesses exposure to contaminants directly from the TSF.

Small mammals (Northern Red-Backed Vole):

- TSF pathway: ingestion of tailings water
- Tundra pathways: ingestion of insects, plants, water; incidental ingestion of soil

Ungulates (Caribou):

- TSF pathway: ingestion of tailings water
- Tundra pathways: ingestion of plants, water; incidental ingestion of soil

Songbirds (Lapland Longspur):

- TSF pathway: ingestion of tailings water
- Tundra pathways: ingestion of insects, plants, water; incidental ingestion of soil

Waterfowl (Canada Goose):

- TSF pathway: ingestion of tailings water
- Tundra pathways: ingestion of insects, plants, water; incidental ingestion of soil

Shorebirds (Semi-Palmated Sandpiper):

- TSF pathway: ingestion of tailings water and tailings benthic invertebrates; incidental ingestion of tailings sediment
- Tundra pathways: not evaluated1

Inhalation and dermal absorption of metals are generally considered to be insignificant in comparison to exposures through ingestion (USEPA, 2005), so they are not considered here.

1.6.4 Contaminants of Potential Concern (COPCs)

Potential sources of contaminants are indicated in Section 1.5. In the baseline WSLRA, Azimuth (2006) identified COPCs for risk characterization based on the chemical composition of the identified dust sources, the predicted effects of effluent on water quality in Third Portage Lake, and a review of metals regulated under MDMER (see Azimuth, 2006, Section 2.5 for details). Projected concentrations of metals in four dust sources (roads, waste rock and tailings) that exceeded the 90th centile of baseline soil concentrations or CCME Soil Quality Guidelines for the Protection of Environmental and Human Health were included as COPCs for the main Meadowbank minesite. For water, manganese and cadmium were included because they were predicted to exceed CCME Water Quality Guidelines for the Protection of Aquatic Life (cadmium) or aesthetic drinking water objectives (manganese). Although not predicted to exceed guidelines, five metals regulated under MDMER (arsenic, copper, lead, nickel and zinc) and mercury were included in the assessment because they were found to be of concern to the general public in the Arctic.

For the Whale Tail site assessments (Golder, 2016 & 2019a), wildlife COPCs were identified by screening predicted concentrations in primary residency media (soil and water; tailings sediment and water) against established guideline values (primarily CCME sources), or maximum measured baseline concentrations +10% (where baseline was greater than the guideline value).

For Meadowbank WSLRAs completed in 2011, 2014 and 2017, quantitative risk characterization (hazard quotient calculation) was performed for all COPCs identified historically in Azimuth (2006). However, beginning in 2021, measured concentrations in primary residency media (soil and lake water; and TSF sediment and water - Figure 3) collected through the WSLRA field program at sites around the Meadowbank Complex (Section 2.2.2 and Appendix A) will be initially screened against established regulatory guideline values (described below) and/or maximum measured baseline values + 10%, as in Golder (2019a,b) to identify COPCs.

Parameters selected for screening will include all metals with CCME Soil Quality Guidelines for the Protection of Environmental and Human Health (CCME, 2022a) or CCME Canadian Water Quality Guidelines for the Protection of Agriculture (livestock watering - CCME, 2022b), plus COPCs identified in previous Meadowbank WSLRAs (manganese and strontium were identified as COPCs in Azimuth

¹ Based on discussions with ECCC during the permitting process for the Whale Tail Mine Expansion Project, assessment of risk to shorebirds was required to be added for the TSF pathway only (as in Golder, 2019b).

(2006) because predicted concentrations in mine rock dust sources exceeded baseline soil concentrations; cyanide was identified as a COPC for the TSF pathway in Golder (2019b)). Certain chemicals which are controlled through best management practices and which were not addressed in the baseline SLRA include petroleum hydrocarbons, dioxins, nitrates, ammonia and PAHs. For each source of these chemicals, best management practices are in place and environmental exposures are not expected to occur.

The corresponding primary soil and water quality guidelines selected for this screening are the same as those used in the Whale Tail Mine Expansion Project HHERA (Golder, 2019a):

Soil/Sediment:

- Canadian Council of Ministers of the Environment Soil Quality Guidelines for the Protection of Environmental and Human Health (SQG_E – Residential Land Use) (CCME, 2022a)

Water:

- Livestock Watering Guidelines from the CCME Water Quality Guidelines for the Protection of Agriculture (CCME, 2022b)
- Livestock Watering Guidelines from the British Columbia Ministry of Environment (BC MOE, 2021)

Where guideline values are unavailable from these sources for the screening parameters, alternate appropriate sources will be referenced (e.g. CCME, 2022c; BC CSR, 2021a).

Any parameters with measured concentrations exceeding screening values are retained as COPCs for quantitative risk characterization. This approach is consistent with the recent ecological risk assessments for the Whale Tail site (Golder, 2016 & 2019a,b).

1.7 EXPOSURE ASSESSMENT

Exposure assessment is used to calculate the dose of each identified COPC received by each ROC for each complete exposure pathway. The exposure assessment uses the food chain model developed by Azimuth (2006) with updates as in Golder (2019b), as described below. The model was developed to calculate daily exposure to contaminants in the various study locations based on COPC concentrations in exposure media, dietary preferences, ingestion rates and dose-adjustment factors.

Estimated daily intake of each COPC is calculated separately for each study location as:

EDI =
$$\sum_{w,s,f} (I_{w,s,f} \times C_{w,s,f}) \times BF \times T$$

Where:

EDI = estimated daily intake (mg/kg body weight·d)

lws,f = intake of water, soil/sediment and food items (L/kg ww·d; kg dw/kg ww·d; kg dw/kg ww·d)

 $C_{w,s,f}$ = concentration of COPC in water, soil/sediment and food items (L/kg ww·d; kg dw/kg ww·d; kg dw/kg ww·d)

BF = biotransfer factor (absorption factor) – assumed to be 100%

T = proportion of time in area (%)

Each component is described below.

1.7.1 I_{w.s.f} - Intake of Water, Soil/Sediment, and Food

Water, food and soil/sediment ingestion rates used in the assessments are shown in Table 1. All intake parameters are considered to be conservative. Water and food ingestion rates were derived from species profiles or allometric equations in USEPA (1993), as described in Table 1. Soil ingestion rates for Canada Goose and Northern Red-Backed Vole are also from USEPA (1993). Sediment ingestion rates for Semi-Palmated Sandpiper are from Beyer et al. (1994). Although Beyer et al. (1994) was referenced as the source of most soil ingestion rates in the Meadowbank baseline assessment, the species chosen to represent Caribou and Lapland Longspur were not indicated. The soil consumption rate for Caribou was increased in subsequent Meadowbank assessments and here from 2% of dry food consumption to 5%, which is the general rate for mammals in Beyer et al. (1994), as used in (Senes, 2008). The soil ingestion rate for Lapland Longspur was increased from 2% to 7%, based on Hansen et al. (2011). This study identified a rate of 0.7% for Swainson's thrush, a ground-dwelling songbird that primarily feeds on flying insects and berries. A 10x safety factor was applied because Swainson's thrush is a foliage-gleaner, while Lapland Longspur is considered a ground-forager (Cornell University, 2011). This factor is considered to be conservative however, because Lapland Longspur does not scratch the ground to uncover food items as other ground foragers do (Harrison, 1967).

Table 1. Body weight (BW), water intake (I_{water}), soil intake (I_{soil}), and wet and dry (I_{food} ; FI) food intake for the identified ROCs.

Parameter	Units	Value	Reference	Notes			
Northern Re	Northern Red-backed Vole						
BW	kg wet	0.02	Nagorsen (2005)	Smallest body weight used			
I _{water}	L/kg wet/day	0.253	USEPA (1993)	Species profile data for the Prairie Vole			
I _{soil}	kg dry/kg wet/day	0.0012	USEPA (1993)	Assumed 2.4% of dry food ingestion rate (similar to Meadow Vole)			
I _{food}	kg wet/kg wet/day	0.135	USEPA (1993)	Species profile data for the Prairie Vole			
FI	kg dry/kg wet/day	0.049	Not available	Moisture in food assumed to be 64% as per diet moisture calculation			
Caribou		•					
BW	kg wet	75	Dauphine (1976)	Smallest body weight used			
I _{water}	L/kg wet/day	0.064	USEPA (1993)	Based on allometric equation for all mammals (L/day) (0.099*(BW) ^{0.90})			
I _{soil}	kg dry/kg wet/day	0.0013	Beyer et al. (1994)	Assumed 5% of dry food ingestion rate (general rate for mammals)			
I _{food}	kg wet/kg wet/day	0.047	Not available	Moisture in food assumed to be 43% as per diet moisture calculation			
FI	kg dry/kg wet/day	0.027	USEPA (1993)	Based on total dry food intake for herbivorous mammals (g/day) (0.577*(BW) ^{0.727})			

Parameter	Units	Value	Reference	Notes		
Lapland Longspur						
BW	kg wet	0.023	Cornell University (2011)	Smallest body weight used		
I _{water}	L/kg wet/day	0.205	USEPA (1993)	Based on allometric equation for all birds (L/day) (0.059*(BW) ^{0.67})		
Isoil	kg dry/kg wet/day	0.0174	Hansen et al. (2011)	Assumed 7% of dry food ingestion rate (rate of Swainson's thrush +10x safety factor)		
I _{food}	kg wet/kg wet/day	0.654	USEPA (1993)	Moisture in food of insectivorous birds; assumed 62% as per diet moisture calculation		
FI	kg dry/kg wet/day	0.249	USEPA (1993)	Based on total dry food intake for passerine birds (g/day) (0.398*(BW) ^{0.850})		
Canada Goo	se					
BW	kg wet	2.000	Mowbray et al. (2002)	Smallest body weight used		
Iwater	L/kg wet/day	0.044	USEPA (1993)	Species profile data for Canada Goose		
I _{soil}	kg dry/kg wet/day	0.0009	USEPA (1993)	Assumed 8.2% of dry food ingestion rate		
I _{food}	kg wet/kg wet/day	0.032	USEPA (1993)	Species profile data for Canada Goose		
FI	kg dry/kg wet/day	0.011	Not available	Moisture in food assumed to be 66% as per diet moisture calculation		
Semi-Palma	ted Sandpiper					
BW	kg wet	0.0235	ECCC (2019)	Site specific body weight provided from ECCC in teleconference July 10, 2019		
Iwater	L/kg wet/day	0.188	USEPA (1993)	Allometric scaling for birds (L/day) = 0.059 Wt ^{0.67} (kg)		
I _{sediment}	kg dry/kg wet/day	0.059	Beyer et al. (1994)	30% of dry food ingestion rate		
I _{food}	kg wet/kg wet/day	1.0	USEPA (1993)	Allometric scaling for birds (g (dw)/day) = 0.648 Wt ^{0.651} (g); assumed moisture content of 80% per Senes (2008)		
FI	kg dry/kg wet/day	0.197	USEPA (1993)	Allometric scaling for birds (g (dw)/day) = 0.648 Wt ^{0.651} (g); assumed moisture content of 80% per Senes (2008)		

1.7.2 $C_{w,s,f}$ - Dietary Concentrations of COPCs

Dietary concentrations of COPCs in drinking water, soil/sediment, and food items are calculated as:

Cs = [soil]; (measured directly)

Cw = [drinking water]; (measured directly)

 $C_{\text{food items}} = \sum_{\text{food items}} ([\text{food item}] \times DP(\%)); \text{ (measured or modeled - Section 2.2.2.1)}$

Where:

DP = dietary preference (% of food item in diet of the ROC – Section 2.2.2.2)

Each component is described below.

1.7.2.1 Concentrations in Ingested Media ([soil], [drinking water], [food items])

For tundra exposure pathways, concentrations of COPCs in lake water, soil, and plant tissue (food items: sedges, lichens, berries) are measured directly in samples collected at specified locations around the Meadowbank Complex in assessment years (Figures 1 & 2). For each location, three samples of each media type will be collected. Historically (through 2017), five samples of each media type were collected from each location. Beginning in 2021, three of the five stations will be randomly selected and sampled in any monitoring year. This slightly reduced sampling effort is warranted since to date, no unacceptable mine-related risk has been identified (2011, 2014, 2017, 2021 WSLRA assessments), and recent risk assessments for the Whale Tail site (Golder, 2016; Golder, 2019a; Golder, 2019b) have not identified any anticipated unacceptable risk.

Lake water samples from the Core Receiving Environment Monitoring Program (CREMP) will be used in the WSLRA analyses for drinking water in tundra pathways (assessment lakes and 2021 sites shown in Figure 1 – specific sites within each lake change annually). Onsite (main Meadowbank minesite) concentrations will be from samples collected in Second Portage Lake (SPL) and the east and north basins of Third Portage Lake (TPE, TPN). AWAR concentrations will be from samples collected in TPN. Near-site concentrations will be from samples collected in Tehek Lake (TE). Whale Tail Mine concentrations will be from samples collected in Whale Tail Lake South and Mammoth Lake. Whale Tail Haul Road concentrations will be from samples collected in Pipedream Lake (PDL). External reference samples are from Inuggugayualik Lake (INUG) and PDL. Exact coordinates are subject to slight changes each year – see CREMP Plan for details.

For the TSF exposure pathway, water quality samples (ponded North and South Cell Tailings water) are collected monthly during the open water season (typically June – October) under regular mine-site monitoring programs. Results for the WSLRA monitoring year are used in the assessment. Historically (2021 WSLRA Report and Golder (2019b)) tailings sediment exposures were assessed using monthly analysis results for direct mill effluent. Moving forward, tailings beach sediment will be preferentially sampled in the WSLRA monitoring year, as it is more representative of exposure media.

An SOP for sample collection along with UTM coordinates for sample locations is provided in Appendix A.

For the identified COPCs in tundra pathways (see Section 2.1.4), the upper 95% confidence limit of the mean (UCLM) concentrations in soil and plant tissue for each assessment location will be used for food chain modeling. If measured values are below the laboratory detection limit, a value of one half the detection limit will be used in calculations. Based on published literature, methyl mercury is assumed to comprise 1% of total mercury in water and soil, and 34% of total mercury in plant tissue, and inorganic mercury = total – methyl mercury (Azimuth, 2006).

For the TSF pathway, maximum measured values in tailings sediment and ponded TSF water were initially planned to be used in EDI calculations, as in Golder (2019b). This approach is more conservative than the UCLM method and was adopted because of the relatively low sample size for these media (generally one sample per month). However, since exposure concentrations for sediment

will now be measured in a suite of tailings beach sediment locations (10+) during the in-migration period in each WSLRA monitoring year, rather than a single monthly sample of mill effluent, a measure of central tendency (e.g. mean measured concentrations, or UCLM) will also be used in EDI calculations for the TSF pathway, as for the tundra pathway.

Concentrations of COPCs in terrestrial insects consumed by certain ROCs (Section 2.2.2.2) are not measured, but are modeled from soil concentrations using published bioaccumulation models for arsenic, cadmium, copper, lead, and zinc (Sample and Arenal, 2001; as described in Azimuth, 2006):

In[insect] = B0 + B1(In[soil]); concentrations are expressed in mg/kg dry weight

This method is particularly conservative, because the modeled factors are for ground insects whereas the songbird population in this assessment consumes primarily flying insects. A bioaccumulation factor (BAF) of 1 is assumed for all remaining COPCs, which is also considered to be very conservative.

For benthic invertebrates, BAFs from USEPA (1999) will be used to estimate whole-body tissue concentrations based on measured sediment concentrations as:

[benthic invertebrate] = BAF x [sediment]; concentrations are expressed in mg/kg dry weight. Available BAFs are shown in Table 2. For all other COPCs, a BAF of 1 is assumed.

Table 2. Bioaccumulation factors (BAFs) from USEPA (1999) used for estimating whole-body concentrations of COPCs in benthic invertebrates.

Parameter	BAF
Aluminum	0.90
Arsenic	0.90
Barium	0.90
Chromium	0.39
Copper	0.30
Lead	0.63
Mercury	0.068
Nickel	0.90
Selenium	0.90
Silver	0.90
Thallium	0.90
Zinc	0.57
Cyanide	0.90

1.7.2.2 Dietary Preferences (DP)

For tundra pathways, the proportions of food items (sedge, lichen, berries, insects, benthic invertebrates – Table 3) contributing to the diet of each receptor for each assessment location were determined using literature reviews in Azimuth (2006). Sedges, lichens and berries are considered surrogates for all plant matter ingested by the ROCs.

For the TSF pathway, as in Golder (2019b), Semi-Palmated Sandpiper is assumed to consume drinking water and benthic invertebrates from the TSF. All other receptors are assumed to consume water only.

For informational purposes, in 2021, sampling for benthic invertebrates was conducted within the TSF to determine whether any significant populations are established in this area. Since the TSF is not intended as aquatic habitat, it was considered unlikely that any resident populations of invertebrates would be substantial enough to fulfill the dietary needs of shorebirds. Results of this exploratory work are presented in the 2021 WSLRA Report (Agnico Eagle, 2022). Briefly, average abundance among

the five samples collected was approximately 147 organisms/m², which is significantly less than historical average abundance in area reference systems (1172 organisms/m² – as reported in the 2020 CREMP Report (Azimuth, 2021), Table 4-10, historical average for Inuggugayualik and Pipedream Lakes). Nevertheless, shorebirds are conservatively assumed to obtain 100% of their food intake as benthic invertebrates from the TSF.

Table 3. Proportion of dietary items obtained from the WSLRA assessment locations for the receptors of concern.

Dietary Item	Northern Red- Backed Vole	Caribou	Lapland Longspur	Canada Goose	Semi-Palmated Sandpiper
Tundra Pathways					
Sedges	55%	30%	25%	50%	-
Lichens	0%	65%	0%	0%	-
Berries	40%	5%	5%	45%	-
Insects	5%	0%	70%	5%	-
Total Food	100%	100%	100%	100%	-
Drinking Water	100%	100%	100%	100%	-
TSF Pathway					
Benthic Invertebrates	-	-	-	-	100%
Drinking Water	100%	100%	100%	100%	100%

1.7.3 Biotransfer Factor

The uptake efficiency factor (biotransfer or absorption factor) describes the proportion of the COPC that is absorbed into the animal from any ingested sources. Uptake efficiency was conservatively assumed to be 100% for all COPC/receptor combinations. This is likely an extremely conservative assumption; for example, chromium compounds were found to have a maximum absorption efficiency of 10% in the GI tract (Outridge and Scheuhammer, 1993).

1.7.4 Time in Area

Territory size (foraging range) affects the proportion of an animal's diet that could be affected by mine-related contaminants. In the baseline assessment for Meadowbank (Azimuth, 2006), an adjustment factor for foraging range was not applied (animals were assumed to spend 100% of time in the study areas). For subsequent assessments, the only ROC assumed to spend 100% of its time in any study area is the Northern Red-Backed Vole, because of its small territory size. Canada Geese, and Lapland Longspur are migratory species, and the fraction of time spent in any study area for those species (onsite, near-site, AWAR, Whale Tail site, Whale Tail Haul Road) is estimated at 33%, based on the 2008 Screening Level Environmental Effects Assessment for the Kiggavik Project (Senes, 2008).

The time caribou spend in any study area (12%) was determined through an examination of collared caribou from the Meadowbank region, which found that any one animal spent no more than a maximum of 12% of the year within 25 km of the minesite (Martin Gebauer and Jason Shaw, personal communication, March 2012). This is similar to the estimate of 10% used in the 2004 assessment of the Lupin minesite (Golder, 2004).

For the TSF, time-in-area factors from Golder (2019b) were used. While Semi-Palmated Sandpiper are migratory and estimated to spend 33% of the year in the minesite area (Senes, 2008), they are not expected to obtain 100% of their food and water from the TSF, since best management practices are in place to actively discourage wildlife from this area. During breeding season, inspections are performed at least once per day, and birds are deterred from the open-water areas of the TSF through the use of personnel presence, decoys, noise cannons, and flares as necessary. While bird presence around the TSF occurs for up to 2 weeks in the very early spring, prior to ice-off on natural lakes, very few birds are observed in this area after that time. Therefore, the proportion of Semi-Palmated Sandpiper exposure to COPCs originating from the TSF is conservatively over-estimated at 8% (i.e. 4 weeks per year). The same factor was applied for all other receptors except vole, based on the ice-free season and that voles are not actively deterred from the TSF as the other receptor types are.

Risk will be characterized for small-territory ROCs (Northern Red-Backed vole, Canada Geese and Lapland Longspur) for main Meadowbank minesite (onsite), near-site, AWAR, Whale Tail Pit, Whale Tail Haul Road, and external reference locations separately, in order to determine whether those animals choosing territories at any mine-related location are at increased risk compared to those choosing territories at nearby reference locations. Exposure data for main minesite and near-site locations will be combined for Caribou because Caribou can readily roam between the onsite and near-site locations in the course of a day. Risk for Semi-Palmated Sandpiper is determined due to exposure to contaminants in the TSF only, based on recommendations from ECCC and commitments made during the 2018 Whale Tail Final Hearings.

Time-in-area adjustment factors are summarized in Table 4.

Table 4. Time-in-area assumption for each study area and ROC.

Study Area	Northern Red-Backed Vole	Caribou	Lapland Longspur	Canada Goose	Semi-Palmated Sandpiper
Onsite	100%	12%	33%	33%	-
Near-site	100%	1270	33%	33%	-
AWAR	100%	12%	33%	33%	-
Whale Tail Pit Area	100%	12%	33%	33%	-
Whale Tail Haul Road	100%	12%	33%	33%	-
External Reference	100%	12%	33%	33%	-
TSF	25%	8%	8%	8%	8%

1.8 TOXICITY ASSESSMENT

Toxicity reference values (TRVs) used to date in the Meadowbank assessments are provided in Appendix B. These were collated in Azimuth (2006) unless otherwise indicated, from a review of the literature; mainly from Sample et al. (1996). This still represents one of the most comprehensive and commonly used sources available for wildlife toxicity reference values and has been used in other similar assessments for this region (e.g. Senes, 2008). In order to ensure the selected TRVs were relevant to the Meadowbank site and the conditions of that risk assessment, several criteria were used in the baseline assessment in screening toxicity studies. These included selecting values from studies

conducted on species of similar phylogeny (i.e. bird or mammal), and selecting studies that examined individual or population-level effects over chronic time periods. The following describes TRV selection, as performed by Azimuth (2006):

The TRVs chosen for use in the risk characterization include both no observable adverse effect levels (NOAELs) and lowest observable adverse effect levels (LOAELs) when available. If effects concentrations were reported in terms of food concentrations, these were converted to dose. If a LOAEL was reported but no NOAEL could be determined, it was estimated as 1% of the LOAEL (as in Sample et al. 1996, Chapman et al. 1998). LOAELs cannot be estimated if only a NOAEL is available. Since the protection goal of this risk assessment no adverse effect of COPCs on populations of the ROCs, LOAELs are the most relevant TRV, and are used in the final risk estimate.

Where toxicity information was found for multiple forms of a contaminant, the one with the greatest toxic potency was chosen. TRVs for chromium-VI were available for mammals, but only chromium-III was available for birds. No NOAELs or LOAELs were available for total mercury. Mammalian LOAELs were not available for inorganic mercury or beryllium. Avian LOAELs were not available for uranium or vanadium. Avian NOAELs were not available for antimony and beryllium and were extrapolated from the mammalian values. The avian LOAEL for antimony was extrapolated from the mammalian value.

While previous versions of this plan and WSLRAs in 2011 – 2017 included allometric scaling to adjust mammalian TRVs from test species to ROCs, those were removed beginning in Version 5, as recommended in Allard et al. (2010), and to align with Golder (2019a,b) methods.

The TRV for cyanide used for the TSF pathway was obtained from Ma and Pritsos (1997), as applied for another Northern shorebird, the Common Snipe, in Golder (2004).

TRVs presented here will be reviewed and updated as necessary in future assessments.

1.9 RISK CHARACTERIZATION

Risk characterization compares predicted exposure concentrations with the toxicity reference values from the literature, using the hazard quotient approach. Hazard quotients for all study areas (main Meadowbank minesite (onsite), near-site, AWAR, Whale Tail Pit, Whale Tail Haul Road, TSF, and external reference) will be calculated as:

HQ = EDI / TRV

Where:

EDI = estimated daily intake (mg/kg body weight·d)

TRV = toxicity reference value (mg/kg body weight·d)

Because of the conservative assumptions included at this level of assessment, there is generally considered to be a high degree of certainty associated with results indicating negligible risk. A hazard quotient > 1 indicates the possible need for more in-depth assessment, including analysis of assumptions used. However, when HQ values exceed 1 for both the external reference (or baseline) and study areas, and are of similar magnitude, it may be assumed that the receptor is adapted to the measured exposure level, or that the assumptions used in calculating the HQ have resulted in an overestimation of risk (Dominion Diamond, 2015).

HQ values and a characterization of risk for each ROC will be provided in the assessment report.

1.10 UNCERTAINTY ASSESSMENT

The assumptions included in each section of the assessment will be discussed, along with implications for over- or under-estimating risk.

HUMAN HEALTH - COUNTRY FOODS ASSESSMENT

1.11 PROBLEM FORMULATION

The HHRA_{country foods} assessment will re-evaluate risk to human receptors for country foods exposure pathways identified in the pre-construction HHRA (Wilson, 2006) and follow-up assessments (HHRAs conducted in 2011, 2014, 2017, and 2021), making use of environmental samples collected through the Wildlife Assessment (Section 2.2.2). The conceptual model for country foods consumption is shown in Figure 4.

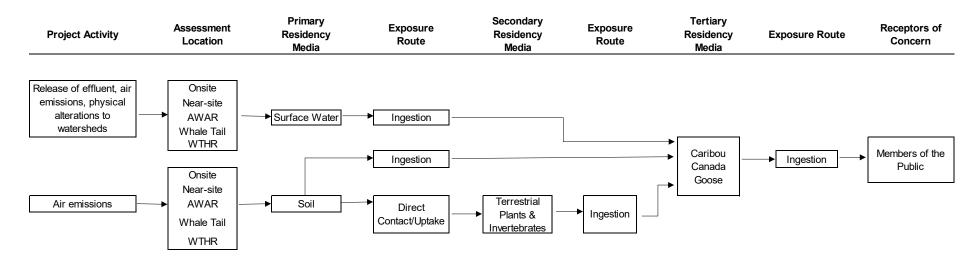


Figure 4. Human health (country foods) risk assessment conceptual model.

1.11.1 Exposure Pathways

In the baseline HHRA (Wilson, 2006), the consumption of terrestrial mammals, waterfowl, fish and plants was assessed for people residing in the hamlet of Baker Lake. From reviews of oral testimony collected in 2005 (Traditional Knowledge Report as part of the Meadowbank FEIS - Cumberland, 2005), Caribou meat, kidney and liver, Canada Goose meat and Lake Trout were found to be the food items most representative of local consumption patterns. Lake Trout was included in the baseline assessment because they represent the majority of fish in the project lakes. However, since a no-fishing policy was put in place for workers and fish from project lakes are non-migratory, consumption of fish impacted by the mine site is expected to be negligible. Analyses of risk from fish consumption were therefore excluded from subsequent assessments. Although local residents may consume wild berries, it was found to be unlikely that they would be harvested from the mine site area due to distance, the fact that public access is prohibited past km 85 on the AWAR, and abundance of this food source closer to Baker Lake. Consumption of plants was therefore not evaluated in the baseline assessment or subsequent updates. Finally, although risk analyses for consumption of Canada Goose are maintained in this Plan, it is noted that only 7% of the population of Baker Lake was found to consume this item, at a frequency of less than 1 day per month (Areva, 2011).

The following food items are therefore included in the HHRA_{country foods} evaluation:

- Caribou meat (muscle)
- Caribou kidney
- Caribou liver
- Canada Goose meat (muscle)

Assessed exposure pathways for these country food items are identified in Figure 4.

1.11.2 Potential Receptors

For consistency with Wilson (2006), potential receptors are considered to be a young child or toddler (age 7 months – 4 years) and an adult consumer of country foods. These receptors are considered to be representative and protective of the general population.

1.11.3 Contaminants of Potential Concern

Potential sources of contaminants are identified in Section 1.5.

As described in Section 2.1.4, pre-construction wildlife risk assessments for the Meadowbank site identified COPCs according to predicted concentrations in dust sources, effluent, and a review of metals regulated under MDMER (see Azimuth 2006, Section 2.5 for details). These parameters were also assessed as COPCs in the pre-construction HHRA_{country foods} (Wilson, 2006), and subsequent 2011, 2014, and 2017 assessments under operational conditions.

For the Whale Tail site HHERAs (Golder, 2016 & 2019a), COPCs were identified by screening predicted concentrations in primary residency media (soil and water) against established regulatory guideline values that are protective of human health, or maximum measured baseline values +10% (where baseline was greater than the guideline value).

Beginning in the 2021 assessment, human health COPCs will be identified for risk characterization using the screening approach described in Golder (2019a), which is consistent with the wildlife

assessment approach (Section 2.1.4). Measured concentrations in primary residency media (e.g. soil and lake water; Figure 4) collected through the WSLRA field program at sites around the Meadowbank Complex (Section 2.2.2 and Appendix A) will be initially screened against established regulatory guideline values (described below) and/or maximum measured baseline values + 10%, as in Golder (2019a) to identify COPCs.

Parameters selected for screening will include all metals with CCME Soil Quality Guidelines for the Protection of Environmental and Human Health (CCME, 2022a) or Health Canada Canadian Water Quality Guidelines for Drinking Water (Health Canada, 2020), plus any COPCs identified in previous assessments (manganese and strontium were identified as COPCs in 2006 because concentrations in onsite dust sources were predicted to exceed baseline soil concentrations).

The corresponding primary soil and water quality guidelines selected for this screening are the same as those used in the Whale Tail Mine Expansion Project HHERA (Golder, 2019a):

Soil/Sediment:

- Canadian Council of Ministers of the Environment Soil Quality Guidelines for the Protection of Environmental and Human Health (SQG_H – Residential Land Use) (CCME, 2022a)

Water:

Health Canada Canadian Water Quality Guidelines for Drinking Water (Health Canada, 2020)

Where guideline values are unavailable from these sources for the screening parameters, alternate appropriate sources will be referenced (e.g. BC CSR, 2021a,b).

Any parameters with measured concentrations exceeding screening values will be retained as COPCs for quantitative risk characterization.

1.12 EXPOSURE ASSESSMENT

Exposure assessment is used to calculate the dose of each COPC received by country foods consumers. For consistency, the exposure assessment follows methods from Wilson (2006), with updates as described below, and is based on the food chain model for Caribou and Canada Goose developed by Azimuth (2006).

Daily exposure to COPCs from consumption of country foods is calculated based on Health Canada (2012) as:

Dose = C_F x IR_F x RAF_{ORAL} / BW

Where:

Dose = estimated daily intake of COPC from consumption of food item; µg/kg bw·d

C_F = concentration of COPC in food items (caribou meat, kidney, liver, goose meat); mg/kg ww

IR_F = consumption rate of food item; g/day

RAF_{ORAL} = relative absorption factor (assumed to be 1)

BW = body weight of person; kg

Each component is described below

1.12.1 C_F - COPC Concentrations in Country Food Items

To estimate risk from consumption of country foods, concentrations in each food item (Caribou meat, Caribou organs, Canada Goose meat) are modelled from the soil, lake water, sedge, lichen, and berry samples collected as described in Section 2.2.2.

Estimated daily intake (EDI) for Caribou and Canada Goose is calculated as described in Section 2.2, except source concentrations used for HHRAcountry foods food chain modeling are maximum measured values for each location, as recommended by Health Canada (2012), as opposed to the 95% UCLM values used in the baseline assessment and WSLRA.

Tissue concentrations for each food item are then calculated as follows:

1.12.1.1 Caribou Muscle

To estimate concentrations of COPCs in Caribou muscle, EDIs for Caribou are multiplied by body weight and feed-to-muscle biotransfer factors from the literature.

Caribou muscle concentrations (C_M) are calculated as:

Where:

C_M = Concentration in muscle tissue (meat); mg/kg ww

EDIc = estimated daily intake of COPC by caribou; mg/kg ww·d

W_C = caribou weight; kg (75 kg; Dauphine, 1976)

BTF = biotransfer factor (feed to muscle; beef); d/kg ww (Appendix B; or other appropriate referenced source)

1.12.1.2 Caribou Kidney and Liver

Concentrations of COPCs in caribou kidney and liver have historically been estimated from muscle concentrations using muscle-to-kidney and muscle-to-liver transfer factors for Caribou provided by Gamberg (2012) (Appendix B). While these factors were unpublished, they are from a large scale and long-term study that is part of the Northern Contaminants Program. These values differ from the 2006 assessment, in which only kidney transfer factors calculated from mean concentrations were available (mainly for moose), and these were assumed to be representative of both organs in caribou. Factors will be reviewed and sources updated as available at the time of the next assessment.

Kidney concentrations (C_K) and liver concentrations (C_L) were calculated as:

$$C_K$$
 or $C_L = C_M \times BTF$

Where:

C_K = concentration of COPC in caribou kidney; mg/kg ww

C_L = concentration of COPC in caribou liver; mg/kg ww

C_M = concentration of COPC in caribou muscle; mg/kg ww

BTF = transfer factor (muscle to organ; caribou) (Gamberg, 2012, as in Appendix B; or other appropriate referenced source)

1.12.1.3 Canada Goose Muscle

To estimate concentrations in Canada goose muscle, estimated daily intake rates were multiplied by Goose weight and feed-to-muscle biotransfer factors from the literature.

Goose muscle concentrations (C_{GM}) were calculated as:

C_{GM} = EDI_G x W_G x BTF

Where:

C_G = concentration of COPC in goose muscle; mg/kg ww

EDIG = estimated daily intake of COPC by goose; mg/kg ww·d

W_G = weight of goose; 2 kg (Mowbray et al. 2002)

BTF = biotransfer factor (feed to muscle; chicken); d/kg ww (Appendix B)

1.12.2 IR_F – Intake (Consumption) Rate of Food Items

Based on a review of oral testimony and professional judgement, Wilson (2006) considered three scenarios (heavy, moderate and low consumption) for each food item, which will be carried forward to subsequent assessments unless otherwise indicated:

Caribou Muscle

- heavy consumption: 2 meals per day, 365 days per year
- moderate consumption: 3 meals per week, 52 weeks per year
- low consumption: 1 meal per month, 12 months per year

Caribou Organs

- heavy consumption: 1 meal per week, 52 weeks per year
- moderate consumption: 2 meals per month, 12 months per year
- low consumption: 1 meal per month, 12 months per year

Waterfowl Muscle

- heavy consumption: 3 meals per week, 52 weeks per year
- moderate consumption: 1 meal per week, 52 weeks per year
- low consumption: 1 meal per month, 12 months per year

Using a serving size of 200 g/serving for adults and 86 g/serving for toddlers (Richardson, 1997), Wilson (2006) estimated average daily consumption values as presented in Table 5. Since the derivation of consumption rates used in Wilson (2006) could not be traced to quantitative survey data, a review of the literature was performed in 2014 to verify that values were consistent with those available from published sources. This review indicated that some consumption rates for Caribou meat may have been underestimated, while consumption rates for Caribou organs and Canada Goose may have been overestimated compared to values identified in local or regional surveys. As a result, consumption rates were updated to reflect data from these published sources, as described in Table 5. Scenarios of heavy,

moderate and low consumption were maintained to reflect variety in preferences for country foods and the range of consumption rates identified in the literature.

Table 5. Food items, consumption scenarios, and estimated consumption rates. Toddler values are 43% of adult values (Richardson, 1997) unless otherwise indicated.

Food Item	Consumptio n Scenario	Ra	mption ite (d)	Reference/Rationale	
	Toddler A		Adult		
	Heavy	189.2	440	Highest daily intake in Kivalliq survey, as shown in Senes (2008), Table C-1	
Caribou meat (muscle)	Moderate	89.4	208	Average daily consumption in Nunavut survey (IHS, 2012); similar to value recommended by Health Canada (2012) for wild game consumption by Canadian Aboriginal Populations (270 g/d)	
	Low	15	65	Average consumption for men and maximum consumption for toddlers in Baker Lake survey using ¹³⁷ Cs body burdens in 1989-90 (Tracy and Kramer, 2000)	
Caribou organ (kidney, liver)	Heavy	1.2	2.9	Harvest survey estimate: In 2010, there were 5020 caribou harvested by Baker Lake hunters (Areva, 2011) and an adult population of 1779 (GNBS, 2014). At a kidney weight of 187 g (Crete and Nault, 1989), maximum consumption would be of 2.9 g/d if adults consume all kidneys. This value is consistent with a Yukon survey by Schuster et al. (2011) indicating 3.2 g/d consumption of kidney, and 2.5 g/d consumption of liver. Larter and Nagy (2000) indicate 2.1 g/d for kidney. Chan et al. (2012) indicate 2.7 g/d for all ungulate organs combined.	
	Moderate	0.6	1.3	Based on proportion of "heavy" in Wilson (2006)	
	Low	0.3	0.7	Based on proportion of "heavy" in Wilson (2006)	
0	Heavy	9.9	23	Average daily consumption in Nunavut survey (IHS, 2012); also 95 th centile of consumption in Chan et al. (2012) for 2 northern Manitoba reserves	
Canada goose meat (muscle)	Moderate	5.6	13	Average daily consumption in Chan et al. (2012); also value used in Senes (2008) for ptarmigan consumption in Baker Lake	
	Low	0.8	1.8	Based on proportion of "heavy" in Wilson (2006)	

1.12.3 BW - Receptor Body Weight

The following body weight values from Richardson (1997; as recommended in Health Canada, 2012) are used in the HHRA_{country foods} assessments, unless otherwise indicated:

Toddler: 16.5 kg
Adult: 70.7 kg

1.13 TOXICITY ASSESSMENT

Toxicity reference values (TRVs) used in all previous HHRA_{country foods} assessments are presented in Appendix B. TRVs will be reviewed and updated as appropriate at the time of future assessments.

TRVs for metals are expressed as Tolerable Daily Intakes (TDIs) for non-cancer endpoints, and cancer slope factors for cancer endpoints. Inorganic arsenic was the only COPC identified as a potential carcinogen through the oral ingestion route.

1.14 RISK CHARACTERIZATION

1.14.1 Non-Cancer Risks

The risk characterization stage compares predicted exposure concentrations with published tolerable daily intake (TDI) values from the literature. Non-cancer risks were classified using hazard quotients (HQs), which are calculated as:

HQ = Dose / TDI

Where:

Dose = estimated daily intake from country foods; µg/kg bw·d

TDI = toxicity reference value; μg/kg bw·d

HQ values and a characterization of risk for each receptor and consumption scenario will be provided in the assessment report.

Based on recommendations in Health Canada (2012) for single-substance exposure in preliminary quantitative risk assessment, a hazard quotient \leq 0.2 indicates negligible risk when exposure from one pathway (i.e. country foods) is considered. This is different from the baseline HHRA_{country foods} assessment in which negligible risk was associated with an HQ \leq 1.

Because of the conservative assumptions included at this level of assessment, there is generally considered to be a high degree of certainty associated with results indicating negligible risk. For HQs greater than 0.2, adverse health effects will not necessarily occur. Rather, the assumptions and uncertainties associated with the risk analysis should be studied, and the possibility for more detailed or probabilistic assessment may be considered.

The main goal of this assessment is to determine potential effects of the Project over and above background concentrations. Therefore, when HQs exceeded the threshold of 0.2, HQ values will be compared with external reference values or historical data in order to comment on the incremental

effects of the Project on exposure to COPCs in country foods. Values are not directly compared to baseline HQs, because TDIs and some exposure parameters differ in certain cases, as described in the preceding sections.

It is noted that the magnitude of HQs is not necessarily proportional to risk, due to differences in underlying dose-response curves. While very large HQ values may indicate higher potential for risk, small differences in HQs cannot be considered to be significant (Ritter et al. 2002). The expectation of what represents a small difference will be explored individually for each food item/COPC where necessary, and expected significance of the incremental risk (difference in HQs) investigated through additional analysis of the underlying data where warranted.

1.14.2 Incremental Lifetime Cancer Risk

For carcinogenic substances (inorganic arsenic), risk is determined assuming lifetime exposure (no amortization) at adult consumption rates. Incremental lifetime cancer risk (ILCR), is calculated as:

ILCR = LADD x SL

Where:

ILCR = incremental lifetime cancer risk

LADD = estimated lifetime average daily dose from country foods; µg/kg bw·d

SL = slope factor; $(\mu g/kg \cdot d)^{-1}$

ILCR values and a characterization of risk for each receptor and consumption scenario will be provided in the assessment report.

Based on recommendations in Health Canada (2010a) for single-substance exposure, cancer risk is found to be "essentially negligible" (*de minimis*) when ILCR \leq 1 x 10⁻⁵.

1.15 UNCERTAINTY ASSESSMENT

The assumptions included in each section of the assessment will be discussed, along with implications for over- or under-estimating risk.

REPORTING AND ADAPTIVE MANAGEMENT

The WSLRA and HHRA_{country foods} for the Meadowbank Complex will evaluate risks to wildlife and consumers of country foods from contaminant exposure in and around the mine site every three years during operation. Results will be reported to NIRB in the context of Agnico Eagle's Annual Report for the Meadowbank Complex.

Because of the conservative assumptions included at this level of assessment, there is generally considered to be a high degree of certainty associated with results indicating negligible risk (i.e. HQ <1 for the wildlife assessment, or <0.2 for the HHRAcounty foods). When hazard quotients exceed these

targets and differ substantially (generally, by more than an order of magnitude) between mine-related and external reference sites for a certain COPC, incremental risk due to mine operation will be classified as potentially unacceptable and more detailed investigations will be initiated. This may include a desktop review and refining of the assessment parameters, and/or additional sampling in the subsequent year to confirm results. In the case that results of refined assessments continue to indicate unacceptable risk, adaptive management may include such interventions as capping of dust sources, increased road watering, delineation of contaminated areas, and deterrence methods pending reclamation.

REFERENCES

Agnico Eagle (Agnico Eagle Mines Ltd.) 2022. 2021 Wildlife and Country Foods Screening Level Risk Assessment – Appendix of the 2021 Wildlife Monitoring Summary Report. March, 2022.

Agnico Eagle (Agnico Eagle Mines Ltd.) 2018. FEIS Addendum for the Whale Tail Pit Expansion Project. December, 2018.

Agnico Eagle (Agnico Eagle Mines Ltd.) 2016. FEIS for the Whale Tail Pit Project. June, 2016.

Azimuth (Azimuth Consulting Group Ltd.) 2021. 2020 Meadowbank Complex Core Receiving Environment Monitoring Report. March, 2021.

Allard, P., A. Fairbrother, B.K. Hope, R.N. Hull, M.S. Johnson, L. Kapustka, G. Mann, B. McDonald, and B.E. Sample. 2010. Recommendations for the development and application of wildlife toxicity reference values. Integrated Environmental Assessment and Management 6(1):28-37.

Areva, 2011. Kiggavik Project Environmental Impact Statement – Technical Appendix 9A. Socio-Economic Baseline. Areva Resources Canada, Inc. December, 2011.

Azimuth (Azimuth Consulting Group Inc.). 2006. Wildlife Screening Level Risk Assessment for the Meadowbank Site. Prepared by Azimuth Consulting Ltd. for Cumberland Resources Ltd. 2006.

BC CSR (British Columbia Contaminated Sites Regulation), 2021a. BC Reg. 375/96 British Columbia Contaminated Sites Regulation Schedule 3.1 - Matrix Numerical Soil Standards for Human Health and Environmental Protection. Current to February 1, 2021.

BC CSR (British Columbia Contaminated Sites Regulation), 2021b. BC Reg. 375/96 British Columbia Contaminated Sites Regulation Schedule 3.2 Generic Numerical Water Standards - drinking water. Current to July 6, 2021.

BC MOE (British Columbia Ministry of Environment and Climate Change Strategy), 2021. British Columbia Approved Water Quality Guidelines: Aquatic Life, Wildlife, and Agriculture – Guideline Summary. Water Quality Guideline Series, WQG-20. Prov B.C., Victoria, B.C. Available online: https://www2.gov.bc.ca/gov/content/environment/air-land-water/water-quality/water-quality-guidelines/approved-water-quality-guidelines

Beyer, W.N., Connor, E.E., Gerould, S. 1994. Estimates of soil ingestion by wildlife. The Journal of Wildlife Management. 58(2):375-382.

CCME (Canadian Council of Ministers of the Environment). 2022a. Canadian Environmental Quality Guidelines – Soil Quality Guidelines for the Protection of Environmental and Human Health. Current to February 1, 2022. Online: https://ccme.ca/en/resources#

CCME (Canadian Council of Ministers of the Environment). 2022b. Canadian Water Quality Guidelines for the Protection of Agriculture. Livestock Watering. Current to February 1, 2022. Online: https://ccme.ca/en/resources#

CCME (Canadian Council of Ministers of the Environment). 2022c. Canadian Water Quality Guidelines for the Protection of Aquatic Life. Current to February 1, 2022. Available online: https://ccme.ca/en/resources#

CCME (Canadian Council of Ministers of the Environment). 1996. A framework for ecological risk assessment: General guidance. National Contaminated Site Remediation Program. Winnipeg, Manitoba Branch.

Chapman, P.M., A. Fairbrother and D. Brown. 1998. A critical evaluation of safety (uncertainty) factors for ecological risk assessment. Environmental Toxicology and Chemistry. 17:99-108.

Chan, L., Receveur, O., Sharp, D., Schwartz, H., Ing, A., Fediuk, K., Black, A., and Tikhonov, C., 2012. First Nations Food, Nutrition and Environment Study (FNFNES): Results from Manitoba (2010).

Cornell (The Cornell Lab of Ornithology), 2011. All About Birds – Lapland Longspur. Online: https://www.birds.cornell.edu/home/. Accessed January, 2011.

Crete, M. and Nault, R., 1989. Variation in cadmium content of caribou tissues from Northern Quebec. The Science of the Total Environment. 80: 103-112.

Cumberland (Cumberland Resources Ltd.) 2005. Meadowbank Gold Project, Final Environmental Impact Statement. October 2005.

CWS (Canadian Wildlife Service), 1991. Hinterland Who's Who. Semipalmated Sandpiper. Available at: http://www.hww.ca/hww2.asp?id=74

Dauphine, T.C. Jr. 1976. Biology of the Kaminuriak population of barren-ground caribou: Part 4. Report Series No. 38, Canadian Wildlife Service.

Dominion Diamond, 2015. Human and Wildlife Health Risk Assessment Report for the Jay Project. Prepared by Golder Associates Ltd. for Dominion Diamond Ekati Corporation. February, 2015.

GNBS, 2014. Government of Nunavut Bureau of Statistics. Population Estimates. Available at: http://www.stats.gov.nu.ca/en/Population%20estimate.aspx. Accessed February, 2015.

Golder (Golder Associates Ltd.), 2019a. Human Health and Ecological Risk Assessment (HHERA) from the FEIS Addendum (refer to NIRB Public Registry, NIRB Document ID 324905). Dated May, 2019.

Golder (Golder Associates Ltd.), 2019b. Technical Memorandum: Ecological Risk Assessment Calculations for Semi-Palmated Sandpiper and Other Ecological Receptors Exposed to Tailings Materials at the Meadowbank Tailings Storage Facility. Whale Tail Pit Expansion Project Technical Meeting Commitment 45. August 2, 2019.

Golder (Golder Associates Ltd.), 2016. Human Health and Ecological Risk Assessment Summary. Whale Tail Pit Project FEIS Appendix 3-B. June, 2016.

Golder (Golder Associates Ltd.), 2004. Ecological Risk Assessment for the Lupin Mine Tailings Containment Area. Final Report. Prepared by Golder Associates Ltd. For Kinross Gold Corporation, December, 2004.

Hansen, J.A., Audet, D., Spears, B.L., Healy, K.A., Brazzle, R.E., Hoffman, D.J., Dailey, A., Beyer, W.N. 2011. Lead exposure and poisoning of songbirds using the Coeur d'Alene River Basin, Idaho, USA. Integrated Environmental Assessment and Management. 7(4):587-595.

Harrison, C.J.O. 1967. The Double-Scratch as a Taxonomic Character in the Holarctic Emberizinae. The Wilson Bulletin. 79(1):22-27

Health Canada, 2020. Guidelines for Canadian Drinking Water Quality - Summary Table. Water and Air Quality Bureau, Healthy Environments and Consumer Safety Branch, Heath Canada. Ottawa, ON. Available online: https://www.canada.ca/en/health-canada/services/environmental-workplace-health/reports-publications/water-quality/guidelines-canadian-drinking-water-quality-summary-table.html

Health Canada, 2012. Federal Contaminated Site Risk Assessment in Canada – Part I: Guidance on Human Health Preliminary Quantitative Risk Assessment (PQRA). Revised 2012. Environmental Health Assessment Services, Safe Environments Programme, Health Canada, Ottawa, Ontario.

Health Canada, 2010. Federal Contaminated Site Risk Assessment in Canada - Part II: Toxicological Reference Values (TRVs). Environmental Health Assessment Services, Safe Environments Programme, Health Canada, Ottawa, Ontario.

IHS, 2012. 2007-2008 Inuit Health Survey – Contaminant Assessment in Nunavut. Prepared by Laurie Chan, UNBC Staff Members and Graduate Students, and the Nunavut Steering Committee of the International Polar Year Inuit Health Survey. August 2011. Revised and reprinted February, 2012.

Loos, M., Ragas, M.J., Tramper, J.J., Hendriks, A.J. 2009. Modeling zinc regulation in small mammals. Environmental Toxicology and Chemistry. 28(11):2378-2385.

Larter N.C. and J.A. Nagy. 2000. A comparison of heavy metals levels in the kidneys of High Arctic and mainland caribou populations in the Northwest Territories of Canada. Science of the Total Environment 246: 109-119.

Mowbray, T.B., C.R. Ely, J.S. Sedinger and R.E. Trost. 2002. Canada Goose *Branta canadensis*. In: A. Poole and F. Gill (eds.). The Birds of North America. No. 682. Philadelphia: The Academy of Natural Sciences; Washington, D.C.: The American Ornithologists' Union.

Nagorsen, D.W. 2005. Rodents & Lagomorphs of British Columbia. Volume 4, The Mammals of British Columbia. Royal British Columbia Museum Handbook.

Outridge, P.M. and Scheuhammer, A.M. 1993. Bioaccumulation and toxicology of chromium: implications for wildlife. Reviews in Environmental Contamination Toxicology. 130:31-77.

Richardson, G.M. 1997. Compendium of Canadian Human Exposure Factors for Risk Assessment. Ottawa: O'Connor Associates Environmental Inc.

Ritter, L., Solomon, K., Sibley, P., Hall, K., Keen, P., Mattu, G. and Linton, B. 2002. Sources, pathway, and relative risks of contaminants in surface water and groundwater: A perspective prepared for the Walkerton Inquiry. Journal of Toxicology and Environmental Health, Part A, 65:1-142.

Sample, B. and C.A. Arenal. 2001. Development of literature-based bioaccumulation models for terrestrial arthropods. Platform presentation at the 2001 Annual Meeting of the Society of Environmental Toxicology and Chemistry.

Schuster, R.C., Gamberg, M., Dickson, C. and Chan, H.M. Assessing risk of mercury exposure and nutritional benefits of consumption of caribou (*Rangifer tarandus*) in the Ventut Gwitchin First Nation community of Old Crow, Yukon, Canada. Environmental Research, 111:881-887.

Senes Consultants Ltd. 2008. Final Report – Screening Level Environmental Effects Assessment Proposed Kiggavik Project. Report prepared for Areva Resources Canada Inc.

Tracy, B.L. and Kramer, G.H. 2000. A method for estimating caribou consumption by Northern Canadians. Arctic. 53(1):42-52.

USEPA (United States Environmental Protection Agency). 1993. Wildlife exposure factors handbook, EPA/600/R-93/187. December 1993. Available: www.epa.gov/ncea/wefh.html.

USEPA (United States Environmental Protection Agency). 1999. Screening Level Ecological Risk Assessment Protocol for Hazardous Waste Combustion Facilities. Volume 3A and 3B. EPA530-D-99-001C.

USEPA (United States Environmental Protection Agency). 2005. Guidance for Developing Soil Screening Levels. OSWER Directive 9287.7-55. Washington, DC. November, 2003. Revised 2005.

Appendix A

Standard Operation Procedure for Field Sample Collection

Based on Azimuth (2006)

MEADOWBANK COMPLEX - STANDARD OPERATING PROCEDURE

Field Sampling for the Screening Level Risk Assessment Program

Update: March, 2022

SOIL AND VEGETATION

1.16 MATERIAL

- Nitrile gloves
- Stainless steel or plastic garden trowel/spoons
- Liquinox spray bottle
- DI water spray bottle
- Small brush (e.g. toothbrush) or paper towel
- WhirlPaks/Ziplocs (medium or sandwich size) 168 bags minimum
- Sharpie
- GPS
- Camera
- Field book & pencil

1.17 SAMPLE LOCATIONS

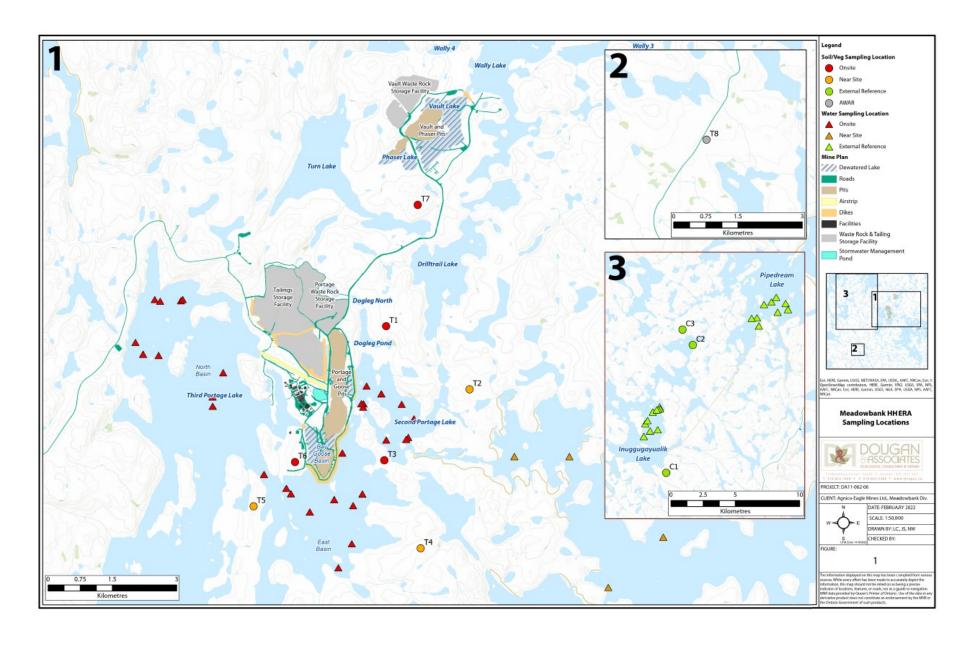
In total, three external reference (control) areas (C1 - C3) and eleven treatment areas (T1 - T11) are to be sampled.

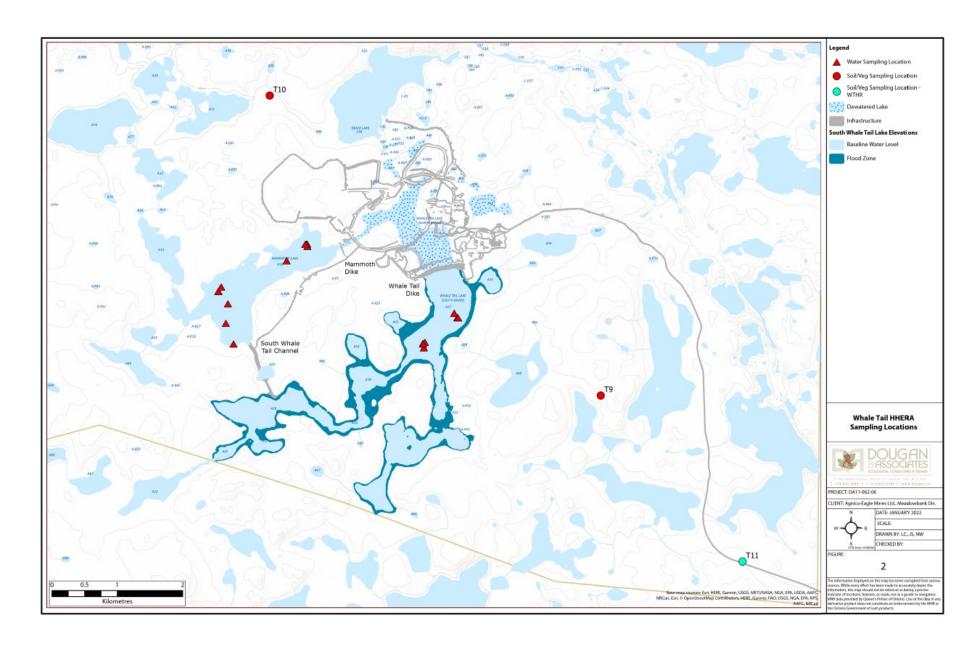
In each area, five sample sites (S1 - S5) are located within a 200 to 300 m radius, at least 150 m apart from one another. In a given year, **only three of the five sites need to be sampled** at random. In 2024, consider analysis of all five sites at T10 to better characterize soils in this area (elevated result at T10-2 in 2021, apparently not mine-related).

UTM coordinates for each sample site are presented in Table 1. Since the fourth and fifth Whale Tail sites have not yet been sampled, coordinates are provided for sites one, two, and three, with the other two to be determined in the field.

Table 1. UTM coordinates for soil and vegetation sampling locations (NAD 83). For each Sampling Area, choose 3 Sites at random. Coordinates for Whale Tail Sites 4 & 5 will be recorded in the field in 2024.

Sampling Area	General location	Site #1	Site #2	Site #3	Site #4	Site #5
T1 2006 location (no longer sampled)	MBK - west side of Dogleg	14N 0639238 7215692	14N 0639137 7215734	14N 0639061 7215668	14N 0639109 7215569	14N 0639010 7215459
T1 – 2011 + location	MBK - see Fig 1	14N 0640110 7215459	14N 0640010 7215458	14N 0640137 7215362	14N 0640090 7215555	14N 0640181 7215525
T2	MBK - see Fig 1	15N 0359410 7214020	15N 0359403 7214128	15N 0359507 7214072	15N 0359459 7213912	15N 0359391 7213816
Т3	MBK - see Fig 1	14N 0640069 7212342	14N 0640146 7212421	14N 0639967 7212281	14N 0639976 7212409	14N 0639991 7212541
T4	MBK - see Fig 1	14N 0640916 7210294	14N 0640994 7210201	14N 0641112 7210194	14N 0640890 7210137	14N 0640802 7210271
T5	MBK - see Fig 1	14N 0637020 7211270	14N 0636978 7211160	14N 0637013 7211394	14N 0637162 7211419	14N 0637057 7211513
T6 - 2006 location (no longer sampled)	MBK - near airstrip	14N 0638559 7213995	14N 0638651 7213953	14N 0638780 7214028	14N 0638515 7214226	14N 0638400 7214038
T6 – 2011+ location	MBK - see Fig 1	14N 0637985 7212300	14N 0638081 7212270	14N 0637887 7212318	14N 0637956 7212202	14N 0637991 7212401
T7	MBK - see Fig 1	14N 0640847 7218280	14N 0640872 7218395	14N 0640755 7218444	14N 0640719 7218338	14N 0640788 7218177
Т8	AWAR km 78, 100 m east side/downwind	14N 0626884 7200614	14N 0626837 7200520	14N 0626806 7200427	14N 0626746 7200306	14N 0626675 7200224
Т9	WT south east side – see Fig 2	14N 609867 7252815	14N 0610005 7252755	14N 0609825 7252981	TBD	TBD
T10	WT north west side – see Fig 2	14N 604817 7257393	14N 0605011 7257344	14N 0604771 7257154	TBD	TBD
T11	WT Haul Road – see Fig 2	14N 612035 7250280 (approx.) Move to 100 m downwind of road in 2024.	14N 0612189 7250361	14N 0612300 7250217	TBD	TBD
C1	Inug – see Fig 1	14N 0623453 7211586	14N 0623450 7211467	14N 0623416 7211345	14N 0623339 7211252	14N 0623217 7211558
C2	Inug – see Fig 1	14N 0625518 7221488	14N 0625569 7221607	14N 0625743 7221542	14N 0625790 7221388	14N 0625825 7221244
C3	Inug – see Fig 1	14N 0624717 7222685	14N 0624818 7222623	14N 0624850 7222504	14N 0624861 7222349	14N 0624636 7222313





1.18 SAMPLE COLLECTION PROCEDURE

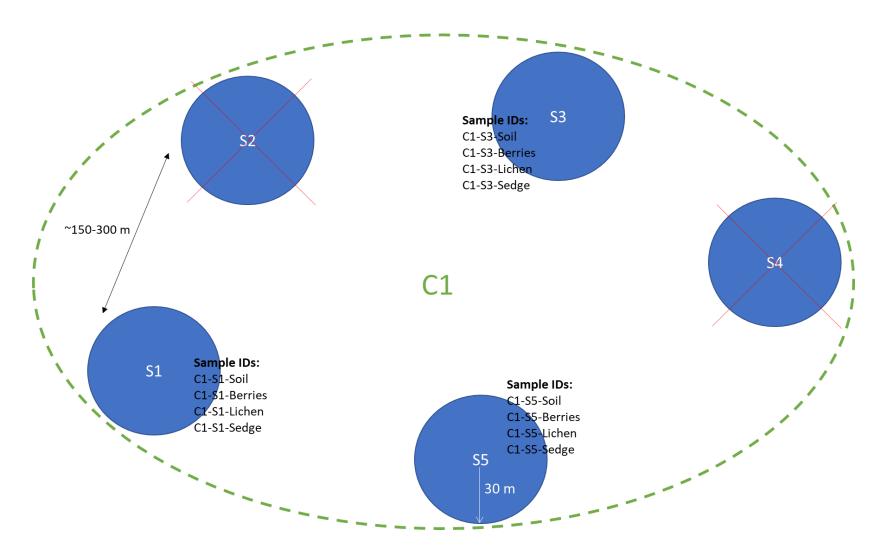
Locate your chosen sample site with the GPS and place your field bag/other marker on the ground in this location for a visual guide.

Take one or two photos of the ground surface at each site, showing an overview of the vegetation types.

At each site, four media are collected: one berry, one sedge/grasses, one lichen, one soil (procedures described below). Samples should be collected within approx. 30 m of the coordinates.

Following sample collection for each site, record date, time, personnel, sample IDs for collected samples (described below), and any general observations in the field book.

Example diagram of sampling sites (S) within a sampling area (C1). In this case, sites S1, S3, and S5 were randomly selected among the 5 sites for sampling.



1.18.1 Vegetation Sample Collection

Wear new nitrile gloves for each sample site.

Label three sample bags with the sample IDs (e.g. C1-S3-Lichen; C1-S3-Berries; C1-S3-Sedge), date, time, and personnel initials.

Handle vegetation gently & avoid shaking off dust. Each type of vegetation is placed in its own prelabelled bag. Collect approximately 20 g of each vegetation type. A min 5 g is required for analysis. Photos of sample media are provided at the end of this document.

Sedges/grasses should be collected by randomly selecting plants and tearing at the base of the aboveground growth. Place in labelled bag.

Reindeer lichen is collected by lifting from the ground surface. Place in labelled bag.

Berries (any variety) are collected by hand-picking, targeting ripe berries. Place in labelled bag.

Squeeze out air and seal bags after sample collection. Freeze upon return to site, prior to shipping.

1.18.2 Soil Sample Collection

Wear new nitrile gloves for soil sample collection.

Label sample bag with the sample ID (e.g. T1-S2-Soil), date, time, and personnel initials.

Remove leaves/organic matter and any significant debris from the ground surface. Using your trowel, collect a soil sample of approximately 50 g from the rooting zone (top 15 cm) and place it in the labelled plastic bag. Squeeze out air and seal the bag.

Refrigerate or freeze upon return to site, prior to shipping.

Clean the sampling trowel between sites by:

- Washing/wiping with a small brush (e.g. toothbrush) or paper towel and Liquinox detergent
- · A final thorough rinse with DI water

1.19 TRACKING, STORAGE, TRANSPORTATION AND LABORATORY ANALYSIS

Fill out chain-of-custody forms for transport. Care should be taken to ensure that the sample identification is clearly marked on each bag, and that the sample ID matches the COC.

Analysis request:

Sample Type	Min Weight Req'd	Analyses	Analysis Code	Total # samples
Soil	50 g	Total metals	Confirm with lab	42
	(target 100 g)	Total mercury	Confirm with lab	42
		pH	Confirm with lab	42
Tissue (use this	5 g	Moisture	Confirm with lab	126
designation for all 3 types of	(target 20 g)	Total metals	Confirm with lab	126
vegetation)		Total mercury	Confirm with lab	126

In total there should be 42 soil samples, and 126 tissue samples (42 berries, 42 sedge, and 42 lichen samples).

Samples should be placed on ice in coolers and shipped, along with the chain of custody records to the analytical laboratory (confirm account manager/lab location annually).

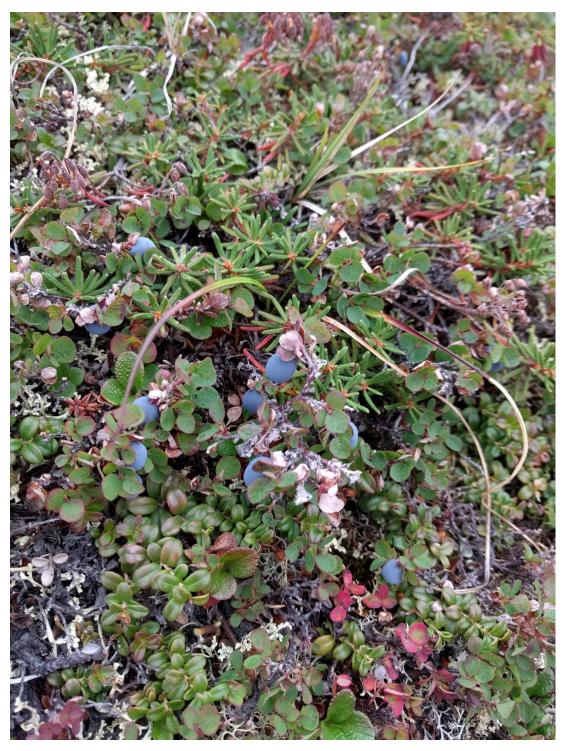


Photo 1: Blueberries (any type of berries can be sampled)



Photo 2: Bear berries (any type of berries can be sampled)



Photo 3: Sedge (any type of graminoid, or grass-like plant can be sampled)



Photo 4: Reindeer lichen (light grey-green tendrils)



Photo 5: Soil pit

WATER

Collected through the CREMP (2PL, 3PL, Tehek, PDL, Inug, WTS, MAM). No supplemental field sampling or additional analyses are required.

TAILINGS ANALYSIS

1.20 BENTHIC INVERTEBRATES

Benthic invertebrates are not analyzed in tailings sediment as part of the basic screening-level risk assessment. Intake of COPCs from ingestion of benthic invertebrate food sources is conservatively modeled from sediment concentrations. If refined risk characterization is required following the initial assessment, analyses of benthic invertebrates in TSF sediment may be conducted as part of supplemental or follow-up analyses to better understand exposure (e.g. density characterization in relation to sediment COPC concentrations, or tissue chemistry analysis).

A one-time analysis of the presence of benthic invertebrates in shoreline tailings sediment was conducted in 2021. Methods are reported in the 2021 WSLRA Report.

1.21 SEDIMENT CHEMISTRY

*See important note on hold time below prior to sampling.

Surface sediment chemistry (metals and CN) will be analyzed at five locations within each of the South Cell and North Cell to document exposure concentrations in areas where birds may be landing and feeding. Sample collection should proceed once during the regular in-migration period, following ice-off on the TSF (May-June).

- 1. Specific locations may be selected in the field, but should be representative of the variety of sediment beach conditions throughout the TSF.
- 2. For each selected location, take a photo of both the shoreline environment (macro scale) and sediment (micro scale), and record date, time, personnel, sample IDs for collected samples (described below), and any general observations (e.g. bird tracks, sediment disturbance, recent/aged depositional area) in the field book
- 3. Using Liquinox soap, wash sampling equipment (shovel, stainless-steel spoon, and four 5-gallon buckets) by scrubbing with the dish cleaning brush and then thoroughly rinsing the equipment with water from a nearby, unimpacted lake.
- 4. Using the clean shovel, collect 1L of sediment from the top 5 cm of the tailings at each of three subsampling areas, <10 m apart. Composite the 1L of sediment from each subsampling location in one bucket to get a total of 3L sediment for a single area.
- 5. Thoroughly mix the composited 3L of sediment and then transfer using a stainless-steel spoon into two 120 mL glass jars.
- 6. Ensure all sediment samples are sealed securely. Pack sediment sampling containers upright in a cooler with ice packs, and packing material, to ensure containers do not break during transport. The ideal storage and transport temperature is 4°C.

Laboratory analysis request is for total metals and CN (WAD and total). A low detection limit is required for the WAD CN (e.g. 0.05 mg/kg in 2022).

IMPORTANT – The hold time for CN is short (14 d from collection to lab prep), so plan accordingly and contact the lab in advance to advise on incoming shipment as well as the low detection limit requirement.

1.22 TAILINGS WATER CHEMISTRY

Collected through regular mine programs (North and South Cell tailings ponded water). No supplemental field collection required.

Appendix B

Toxicity Reference Values, Bio-Transfer Factors, and Tolerable Daily Intake Values

Table B-1: WSLRA Toxicity Reference Values (mg/kg d)

Parameter	Receptor	TRV Basis	Antimony ^{2,3,4}	Arsenic ¹	Barium ¹	Beryllium ^{1,2}	Cadmium ¹	Chromium ^{1,5}	Cobalt ⁷	Copper ¹	Lead ¹	Manganese ¹	Total Hg	Inorg-Hg ¹
Mammals	Northern Red-backed Vole	NOAEL	98.0	0.126	5.1	0.66	1	3.3	0.2	11.7	8	88	na	1
		LOAEL	112.9	<u>1.26</u>	19.8	na	10	13.1	2	15.1	80	284	na	na
	0 "	NOAEL	22.2	0.100			.	0.0	0.0	44.7		22		
	Caribou	NOAEL	98.0	0.126	5.1	0.66	1	3.3	0.2	11.7	8	88	na	1
		LOAEL	112.9	<u>1.26</u>	19.8	na	10	13.1	2	15.1	80	284	na	na
Birds	Lapland Longspur	NOAEL	9.8	2.5	21	0.066	1.5	1	2.37	47	1.1	977	na	0.45
		LOAEL	11.3	7.4	42	na	20	5	4.74	62	11.3	na	na	0.9
	Canada Goose	NOAEL	9.8	2.5	21	0.066	1.5	1	2.37	47	1.1	977	na	0.45
		LOAEL	11.3	7.4	42	na	20	5	4.74	62	11.3	na	na	0.9
	Semi-palmated Sandpiper	NOAEL	9.8	2.5	21	0.066	1.5	1	2.37	47	1.1	977	na	0.45
		LOAEL	11.3	7.4	42	na	20	5	4.74	62	11.3	na	na	0.9

Parameter	Receptor	TRV Basis	MeHg ¹	Molybdenum ¹	Nickel ¹	Selenium ¹	Strontium ^{1,2}	Thallium ^{1,6}	Tin ¹	Uranium ¹	Vanadium ¹	Zinc ¹	CN ⁸
Mammals	Northern Red-backed Vole	NOAEL	0.015	0.26	40	0.2	263	0.0074	23.4	3.1	0.2	160	68.7
		LOAEL	0.025	2.6	80	0.3	na	0.074	35	6.1	2.1	320	na
	Caribou	NOAEL	0.015	0.26	40	0.2	263	0.0074	23.4	3.1	0.2	160	68.7
		LOAEL	0.025	2.6	80	0.3	na	0.074	35	6.1	2.1	320	na
Birds	Lapland Longspur	NOAEL	0.0064	3.5	77.4	0.4	26.3	0.202	6.8	16	11.4	14.5	0.025
		LOAEL	0.064	35.3	107	0.8	na	0.757	16.9	na	na	131	na
	Canada Goose	NOAEL	0.0064	3.5	77.4	0.4	26.3	0.202	6.8	16	11.4	14.5	0.025
		LOAEL	0.064	35.3	107	0.8	na	0.757	16.9	na	na	131	na
	Semi-palmated Sandpiper	NOAEL	0.0064	3.5	77.4	0.4	26.3	0.202	6.8	16	11.4	14.5	0.025
		LOAEL	0.064	35.3	107	0.8	na	0.757	16.9	na	na	131	na

Notes:

No allometric scaling for mammals (or birds) per Golder, 2019b

underline corresponds to an unbounded LOAEL (10X safety factor used to derive the NOAEL) (see text for details)

na indicates that there was no TRV (NOAEL or LOAEL) available

References:

Chetty, K.N., D.s.v. Subba Rao, L. Drummond, D. Desaiah. 1979. Cobalt induced changes in immune response and adenosine triphosphate activities in rats. Journal of Environmental Science and Health. Part B. B14(5):525-544.

Golder (Golder Associates Ltd.), 2004. Ecological Risk Assessment for the Lupin Mine Tailings Containment Area. Final Report. Prepared by Golder Associates Ltd. For Kinross Gold Corporation, December, 2004.

Lynch, B.S., C.C. Capen, E.R. Nestmann, G. Veenstra and J.A. Deyo. 1999. Review of subchronic/chronic toxicity of antimony potassium tartrate. Regulatory Toxicology and Pharmacology. 30: 9-17.

Rossi, F., R. Acampora, C. Vacca, S. Maione, M.G. Matera, R. Servodio, and E. Marmo. 1987. Prenatal and postnatal antimony exposure in rats: effects on vasomotor reactivity development of pups. Teratogenesis, Carcinogenesis, and Mutagenesis 7: 491-496 Sample, B., D.M. Opresko and G.W. Suter II. 1996. Toxicological benchmarks for wildlife: 1996 Revision. Prepared for the Oak Ridge National Laboratory (ORNL), Department of Energy. ES/ER/TM-86/R3

Szakmary, E., G. Ungvary, A. Hudak, E. Tatrai, M. Nary. 2001. Effects of cobalt sulfate on prenatal development of mice, rats, and rabbits, an on early postnatal development of rats. Journal of Toxicology and Environmental Health, Part A 62:367-386. Ueberschar, K.-H., S. Matthes, and H. Vogt. 1986. Effect of thallium salts on broiler and leghorn hens (in German). A3.

Van Vleet, J.F. 1982. Amounts of twelve elements required to induce selenium vitamin E deficiency in ducklings. Amercian Journal of Veterinary Research 43(5):851-857.

¹ Sample et al. (1996)

² Bird TRVs calculated by multiplying the mammal TRVs with a safety factor of 0.1 (see text for discussion)

³ NOAEL from Dieter et al. (1991) as quoted in Lynch et al. (1999)

⁴ LOAEL from Rossi et al. (1987)

⁵ Mammals TRV based on chromium VI; bird TRV based on chromium III

⁶ Ueberschar et al. (1986)

⁷Chetty et al. (1979) for mammal NOAEL TRV, Szakmary et al. (2001) for mammal LOAEL TRV, Van Vleet (1982) for bird TRVs.

⁸Bird TRV from Ma and Pritsos (1997) in Golder (2004); uncertainty factor of 10 was applied to account for chronic exposure.

Table B-2: HHRA_{country foods} biotransfer factors.

		Caribou	Canada Goose				
СОРС	Feed	to-Muscle	Muscle-to-	Muscle-to-	Feed-to-Muscle		
	d/kg	Source ²	Kidney ¹	Liver ¹	d/kg	Source ³	
Antimony	0.0012	IAEA 2010	1.17	0.82	0.006	Staven et al. 2003	
Arsenic	0.002	USEPA 2005	6.45	0.64	0.83	Staven et al. 2003	
Barium	0.00014	IAEA 2010	40.9	2.66	0.019	IAEA 2010	
Beryllium	0.001	USEPA 2005	2.33	0.87	0.4	Staven et al. 2003	
Cadmium	0.0058	IAEA 2010	2049	287	1.75	IAEA 2010	
Chromium	0.0055	USEPA 2005	0.52	0.78	0.8	Staven et al. 2003	
Cobalt	0.00043	IAEA 2010	10.5	18.6	0.97	IAEA 2010	
Copper	0.01	RAIS 2012	2.06	13.7	0.5	Staven et al. 2003	
Lead	0.0007	IAEA 2010	33.2	250	0.8	IAEA 2010	
Manganese	0.0005	IAEA 2010	0.85	0.68	0.05	IAEA 2010	
Inorg-Hg	0.00609	NCRP 1989	105	15	0.03	IAEA 2010	
MeHg	0.00078	USEPA 2005	105	15	0.03	Staven et al. 2003	
Molybdenum	0.001	IAEA 2010	44.2	153	0.18	IAEA 2010	
Nickel	0.006	USEPA 2005	1.68	1.21	0.001	Staven et al. 2003	
Selenium	0.32	IAEA 2010	19.3	2.28	9.7	IAEA 2010	
Strontium	0.008	IAEA 2010	5.27	1.28	0.08	Staven et al. 2003	
Thallium	0.04	USEPA 2005	14.9	2.51	0.8	Staven et al. 2003	
Tin	0.001	RAIS 2012	3.88	9.44	0.8	IAEA 2010	
Uranium	0.00039	IAEA 2010	3.23	2.61	0.75	IAEA 2010	
Vanadium	0.0025	RAIS 2012	4.33	5.85	2	Staven et al. 2003	
Zinc	0.16	IAEA 2010	0.95	1.11	0.47	IAEA 2010	

Notes:

- 1 All muscle-to-organ factors were obtained from Gamberg (2012)
- 2 For Caribou feed-to-muscle factors, all values are for beef, with the exception of selenium, which is for pig.
- 3 All values for Canada Goose are for chicken

References

IAEA, 2010. Handbook of parameter values for the prediction of radionuclide transfer in terrestrial and freshwater environments. International Atomic Energy Agency. Technical reports series, ISSN 0074–1914; No. 47. Vienna, Austria.

Gamberg, M., 2012. Caribou muscle-to-organ transfer factors for metals. Unpublished. Personal communication (mary.gamberg@gmail.com). February 28, 2012.

NCRP. 1989. Screening Techniques for Determining Compliance with Environmental Standards. Releases of Radionuclides to the Atmosphere. National Council on Radiation Protection and Measurements, Bethesda, Maryland.

RAIS, 2012. Chemical Specific Parameters. The Risk Assessment Information System. University of Tennesee. Available at: http://rais.ornl.gov/cgi-bin/tools/TOX search?select=chem spef. Accessed March, 2012.

Staven, L.H., Rhoads, K., Napier, B.A. and D.L. Strenge. 2003. A Compendium of Transfer Factors for Agricultural and Animal Products. Pacific Northwest National Laboratory. PNNL-13421. Prepared for the U.S. Department of Energy. June, 2003.

USEPA, 2005. The Hazardous Waste Companion Database. Human Health Risk Assessment Protocol (HHRAP) for Hazardous Waste Combustion Facilities, Final. United States Environmental Protection Agency. Available at: http://www.epa.gov/wastes/hazard/tsd/td/combust/risk.htm. Accessed March, 2012.

Table B-3: HHRA_{country foods} tolerable daily intake (TDI) values used in the baseline (2005) assessment and subsequent updates (2011, 2014, 2017).

	-	ΓDI (μg/kg·	day)		Source	
COPC						
	2005	2011	2014 & 2017	2005	2011	2014 & 2017
Antimony	0.4	0.4	0.4		IRIS ¹	
Arsenic	2	0.3	0.3	HC 2002	IF	RIS ¹
Barium	200	200	200	I	RIS ¹	HC 2010
Beryllium	2	2	2		IRIS ¹	
Cadmium	1	0.8	1*	HC	2004	HC 2010
Chromium	5.4	1	1	HC 1996	HC 2004	HC 2010
Cobalt	1.4	1.4	1.4		RIVM 200	1
Copper	250	30	91#	HC 2002	HC 2004	HC 2010
Lead	3.6	3.6	0.1+	HC	2004	HC 2014
Manganese	140	-	136 [#]	I	RIS ¹	HC 2010
Inorg-Hg	0.71	0.3	0.3	HC 2002	HC 2004	HC 2010
MeHg	0.2	0.1	0.2^	HC 2002	IRIS ¹	HC 2010
Molybdenum	5	5	23000#	I	RIS ¹	HC 2010
Nickel	17	17	20	IM	2001	IRIS ¹
Selenium	5	5	6200 [#]	I	RIS ¹	HC 2010
Strontium	600	-	600	IRIS ¹		
Thallium	0.07	0.07	0.01*	IRIS ¹		USEPA 2012
Tin	-	200	300	- ITER 2012		ATSDR 2005
Uranium	-	0.6	0.6	- HC 2004		HC 2010
Vanadium	5	5	5		IRIS ¹	
Zinc	700	300	480 [#]	HC 2002	IRIS ¹	HC 2010

Notes:

Inorganic arsenic was the only COPC identified as a potential carcinogen through the oral ingestion route, and the cancer slope factor was 1.80 (mg/kg·day)⁻¹ (Health Canada, 2010).

*provisional or screening value

^value for women of child-bearing age and children <12 yrs

+median dietary lead exposure for the Canadian population

1 - IRIS database accessed in the assessment year - see References.

References:

ATSDR, 2005. Toxicological profile for tin and tin compounds. US Department of Health and Human Services, Public Health Service, Agency for Toxic Substance and Disease Registry.

HC 1996 - reference not sourced (in Wilson, 2006)

HC (Health Canada), 2002. Toxicological Reference Doses for Organic Contaminants. Last updated 20 August 2002. Developed by Food Directorate of Health Canada, Ottawa, Ontario.

HC (Health Canada) 2004. Federal Contaminated Site Risk Assessment in Canada – Part I: Guidance on Human Health Preliminary Quantitative Risk Assessment (PQRA) and Part II: Health Canada Toxicological Reference Values (TRVs). Environmental Health Assessment Services, Safe Environments Programme, Health Canada, Ottawa, Ontario.

HC (Health Canada), 2010. Federal Contaminated Site Risk Assessment in Canada - Part II: Toxicological Reference Values (TRVs). Environmental Health Assessment Services, Safe Environments Programme, Health Canada, Ottawa, Ontario.

HC (Health Canada), 2014. Letter from Nicole Cote, Safe Environments Directorate, Health Canada. To: info@nirb.ca. Subject: Health Canada's review of the additional information provided regarding the 2011-2012 Annual Monitoring Report for the Meadowbank Gold Project, 2012 (NIRB File No. 03MN107). January 27, 2014.

IM (Institute of Medicine), 2001. Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic, Boron, Chromium, Copper, Iodine, Iron, Manganese, Molybdenum, Nickel, Silicon, Vanadium and Zinc. Food and Nutrition Board, Institute of Medicine. National Academy Press, Washington, DC. Available online: http://www.nap.edu/openbook.php?record_id=10026&page=525

ITER, 2012 International Toxicity Estimates for Risk (ITER) database. Bethesda, MD, National Institutes of Health, National Library of Medicine, Toxicology Data Network (TOXNET) (http://toxnet.nlm.nih.gov. Accessed March, 2012.

IRIS - United States Environmental Protection Agency's Integrated Risk Information System (IRIS). Online: https://www.epa.gov/iris (Accessed in the corresponding assessment year)

RIVM. 2001. Re-evaluation of human-toxicological maximum permissible risk levels. National Institute of Public Health and the Environment, Bilthoven, The Netherlands, March 2001.

USEPA. 2012. Provisional Peer-Reviewed Toxicity Values for Thallium and Compounds - Final. Superfund Health Risk Technical Support Center, National Center for Environmental Assessment, Office of Research and Development. US Environmental Protection Agency. Cincinnati, OH 45268. October 25, 2012.

Wilson Scientific Consulting Inc. (Wilson). 2006. Human Health Risk Assessment of Consumption of Country Foods for the Meadowbank Gold Project. Prepared for Cumberland Resources Ltd.

^{*}essential trace element toxicity value (toddlers)