

Toxicity Testing of Chitosan Lactate

Final Report

Report date: July 1, 2008

Submitted to:

Cascade EcoSolutions 3500 188th Street SW, Suite 322 Lynnwood, WA 98037

Washington Laboratory 5009 Pacific Hwy East Suite 2 Tacoma, WA 98424

TABLE OF CONTENTS

	Page
TA	BLE OF CONTENTSI
ŞIG	NATURE PAGEIII
1.0	INTRODUCTION
2.0	METHODS
3.0	RESULTS1
4.0	RESULTS
5.0	QA/QC 6
5.0	REFERENCES

LIST OF TABLES

Table 3. Table 4.	Summary of test conditions for the 48-hour <i>Daphnia pulex</i> acute survival test
	Results of reference toxicant tests

LIST OF APPENDICES

APPENDIX A - Acute toxicity test data

APPENDIX B - Survival and growth test data

APPENDIX C - Reference Toxicant Tests

APPENDIX D - Chain-of-Custody Form

SIGNATURE PAGE

Laboratory Manager

Project Manager

The results of this study intended for the use of our client, subject to the terms and conditions of their contract with Nautilus Environmental. Any reliance on the data by a third party is at the sole and exclusive risk of that party.

1.0 INTRODUCTION

A toxicity testing program was conducted using Chitosan Lactate, as manufactured and distributed by Cascade EcoSolutions, Lynnwood, Washington, in order to meet the requirements specified in Washington Department of Ecology's laboratory guidance document Appendix G titled "Risk Evaluation of Proposed Treatment Chemicals for Stormwater Runoff" (WDOE, 2005).

The program involved conducting a suite of toxicity tests using the product to determine which was the most sensitive species, and to determine the "toxic threshold", defined as the lowest 25% point estimate from the various tests. The tests performed were designated to evaluate effects of the compound in a "juvenile salmonid or other fish rearing habitat" receiving water. The results of the study are provided in this report.

2.0 METHODS

An acute toxicity test was conducted using *Daphnia pulex*. This test involved a 48hr exposure of the test organisms during which survival was monitored. Methods for the test are summarized in Table 1.

Chronic toxicity tests were conducted using rainbow trout (*Oncorhynchus mykiss*) and fathead minnows (*Pimephales promelas*). Each of these species was evaluated using a 7-day survival and growth test; methods for these tests are summarized in Tables 2 and 3.

The concentration series for each test was established in discussion with Cascade EcoSolutions and was based on previous testing. For all of the tests, the sample was weighed and placed in a flask of dilution water to achieve the highest concentration. The test solution was mixed on a magnetic stir plate overnight at 25°C until completely dissolved. Serial dilutions were then made from this stock solution. This procedure was repeated daily for all renewal solutions. Both methods used for sample preparation achieved the goal of exposing test organisms to the intended concentrations.

Control water for all of the tests was moderately hard synthetic water, prepared by addition of reagent grade salts into deionized water to achieve a hardness of 80 to 100 mg/L.

Summary of test conditions for the 48-hour Daphnia pulex acute survival test. Table 1.

Test organism	Daphnia pulex Daphnia pulex
Test organism source	In-house cultures
Test organism age at initiation	< 24 hours
Test duration	48 hours
Test solution renewal	None
Feeding	None
Test chamber	30 ml plastic cup
Test solution volume	25 mL
Test temperature	20 ± 1°C
Dilution water	Moderately Hard Synthetic Water
Number of organisms/chamber	5
Number of replicates	4
Photoperiod	16 hours light/8 hours dark
Aeration	None
Test Protocol	EPA-821-R-02-012 (USEPA, 2002a)
Test acceptability criterion for controls	≥ 90% survival
Reference toxicant	Copper sulfate

Summary of test conditions for the 7-day rainbow trout survival and growth test. Table 2. Test organism

Oncorhynchus mykiss

Test organism source Thomas Fish Company, Anderson, California Test organism age at initiation

16 days post hatch, 2 days post swim up Test duration

7 days Test solution renewal Daily

Feeding Artemia nauplii twice daily Test chamber

1-L plastic beaker

Test solution volume 500 mL Test temperature 15 ± 1°C

Dilution water Moderately Hard Synthetic Water

Number of organisms/chamber 5 Number of replicates 4

Photoperiod 16 hours light/8 hours dark

Aeration None **Test Protocol** USEPA 2005 Test acceptability criterion for controls ≥90% survival Reference toxicant Copper chloride

Table 3. Summary of test conditions for the 7-day fathead minnow survival and growth

Test organism Pimephales promelas

Test organism source Aquatic BioSystems; Fort Collins, Colorado

Test organism age at initiation <48 hours post hatch

Test duration 7 days Test solution renewal Daily

Feeding Artemia nauplii twice daily Test chamber

500 mL plastic cup

Test solution volume 250 mL Test temperature 25 ± 1°C

Dilution water Moderately Hard Synthetic Water

Number of organisms/chamber 10 Number of replicates 4

Photoperiod 16 hours light/8 hours dark Aeration

None

Test Protocol EPA-821-R-02-013 (USEPA, 2002b)

Test acceptability criterion for controls \geq 80% survival; \geq 0.25 mg average dry weight Reference toxicant

Sodium chloride

3.0 RESULTS

Results of acute toxicity tests conducted with *D. pulex* are provided in Table 4. Results of definitive tests evaluating survival and growth of rainbow trout and fathead minnows are provided in Table 5.

The chronic test with fathead minnows was the most sensitive test conducted in this evaluation. The highest concentration with no observed effect (NOEC) was 7.5 mg/L Chitosan Lactate for survival and 15 mg/L Chitosan Lactate for growth. The EC25 for fathead minnow survival and growth was 15.7 mg/L and 17.4 mg/L Chitosan Lactate respectively. The toxic threshold, as defined in WDOE (2003) as the lowest 25% point estimate from the suite of tests, was 15.7 mg/L for fathead minnow survival. For application of the product, WDOE requires a three-fold safety margin between the concentration of the product at discharge and the toxic threshold for the product (WDOE, 2003). Thus, the data supports application of Chitosan Lactate at a rate of 5.23 mg/L. Higher applications rates may be appropriate if concentrations of Chitosan Lactate are reduced as a result of interaction with turbidity.

Table 4. Percent survival of test organisms in acute toxicity tests.

Chitosan Lactate mg/L	Daphnia survival	
Control	(% ± S.D.)	
15.0	100.0 ± 0.0	
30.0	90.0 ± 11.5	
60.0	100.0 ± 0.0	
120	100.0 ± 0.0	
240	95.0 ± 10.0	
	40.0 ± 28.3	
Endpoints (mg/L Chitosan Lactate)		
NOEC1	444	
LOEC ²	120	
LC503	240	
℃25⁴	353	
No OL 170	138 Served Effect Concentration	

¹No Observed Effect Concentration ²Lowest Observed Effect Concentration

³ Predicted concentration to cause 50% mortality in test organisms

⁴Predicted concentration to cause 25% mortality in test organisms

Table 5. Results for 7-day survival and growth tests using rainbow trout and fathead minnows.

Chitosan Lactate		Rainbow trout		Fathead minnow	
(mg/L)		Survival (% ± S.D.)	Biomass $(mg \pm S.D.)$	Survival (% ± S.D.)	Biomass (mg ± S.D.)
Control-		100.0 ± 0.0	34.6 ± 1.6	100.0± 0.0	0.44 ± 0.04
0.9375		100.0 ± 0.0	34.1 ± 3.3		
1.875		100.0 ± 0.0	34.9 ± 2.0	14, 14, 14 <u>.</u>	
3.75		100.0 ± 0.0	35.6 ± 1.4	100.0 ± 0.0	0.45 ± 0.05
7.5		100.0 ± 0.0	35.1 ± 2.0	92.5± 9.6	0.38 ± 0.06
15		100.0 ± 0.0	31.2 ± 0.6	82.5 ± 9.6	0.37 ± 0.07
30		95.0 ± 0.10	22.9 ± 3.2	42.5 ± 17.1	0.16 ± 0.08
60			-	0.0 ± 0.0	nc
Endpoints (mg/L (Chitosan	Lactate)			
NOEC		30	15	7.5	15
LOEC		>30	30	15	30
LC50		>30		23.4 (20.1-27.4)	50
LC25		>30		15.7 (12.8-18.4)	
IC501			>30		25.3 (19.9-32.2)
IC25 ²			24.2 (19.6-29.9)		17.4 (1.46-21.5)

¹ Predicted to cause 50% reduction in growth in test organisms

4.0 QA/QC

Control performance criteria specified in the corresponding protocols were met for all of the tests. Results of reference toxicant tests are provided in Table 7. The results of the reference toxicant tests indicated that the test organisms were of suitable sensitivity.

Table 6. Results of reference toxicant tests.

Date	Organism	Test type	Effect concentration	Acceptable range
5/29/08	Daphnia pulex	48-hr survival	7.54 μg/L Cu	2.36 - 19.0
6/3/08	Fathead minnow	7-d survival	4.93 g/L NaCl	4.06 - 5.89
0/0/00	Taulcau Hilliow	7-d growth	3.85 g/L NaCl	2.90 - 5.58
6/11/08	Rainbow trout	7-d survival	67.6 μg/L Cu	34.4 - 84.7
0/11/00	Namibow trout	7-d growth	63.1 μg/L Cu	36.0 - 86.1

²Predicted to cause 25% reduction in growth in test organisms

REFERENCES

- USEPA. 2005. Rainbow Trout Oncorhynchus mykiss and Brook Trout Salvenius continalis 7-day survival and growth method, version 6/16/2005. Office of Research and Development, NERL.
- USEPA. 2002a. Methods for Measuring the Acute Toxicity of Effluents and Receiving Waters to Freshwater and Marine Organisms, Fifth Edition. EPA-821-R-02-012.
- USEPA. 2002b. Short-term methods for estimating the chronic toxicity of effluents and receiving waters to freshwater organisms. Fourth Edition. October 2002. EPA-821-R-02-013.
- WDOE. 2005. Laboratory Guidance and Whole Effluent Toxicity Test Review Criteria. WDOE Publication No. WQ-R-95-80. June 2005.