



Environmental Effects Monitoring (EEM): Cycle 1 Interpretative Report

Meadowbank Division, Nunavut



Prepared for:

Agnico-Eagle Mines Ltd.
Meadowbank Division

P.O. Box 540
Baker Lake,
Nunavut X0C 0A0

Prepared by:



Azimuth Consulting Group
218-2902 West Broadway Vancouver, BC V6K 2G8
Tel: 604-730-1220 • Fax: 604-739-8511

FINAL

**Environmental Effects Monitoring (EEM): Cycle 1
Interpretative Report
Meadowbank Division, Nunavut**

Prepared for:

**Environment Canada
Prairie and Northern Region**
Room 200
4999 – 98th Ave. NW
Edmonton, AB T6B 2X3

Prepared on behalf of:

Agnico-Eagle Mines Ltd.
Meadowbank Division
Baker Lake, NU
X0A 0A0

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Azimuth Consulting Group Partnership
218-2902 West Broadway
Vancouver, BC
V6K 2G8

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Key personnel conducting this project were as follows:

- Gary Mann (Azimuth) – Gary was responsible for overall management of this project. He also provided oversight and logistical support for the field crew, collaborated on statistical approach development, conducted the statistical analyses, and authored the report.
- Maggie McConnell (Azimuth) – Maggie teamed with Morgan to collect water and sediment quality samples as well as benthic invertebrates. Maggie also coordinated and collaborated in data collation, analysis, interpretation and reporting.
- Morgan Finley (Azimuth) – Morgan teamed with Maggie to collect a number of the field parameters and was responsible for on-site logistics for the 2011 field monitoring program.
- Sue Hertam, Mike Johnson, Laura Henderson, Claire Hrenchuk (North/South Consultants Inc.) – this group of individuals was responsible for on-site logistics with regards to fish sampling. They also collected all of the fish for this study, processed them in the field laboratory, and entered all of the relevant fish data.
- Randy Baker (Azimuth) – Randy provided oversight and logistical support for the field crew, and reviewed the report (except for the section on benthic invertebrates).
- Ryan Hill (Azimuth) – Ryan reviewed the report (particularly the section on benthic invertebrates).
- Brian Pyper (Azimuth Associate) – Brian provided guidance with statistical part of the data assessment.



PROFESSIONAL LIABILITY STATEMENT

This report has been prepared by Azimuth Consulting Group (Azimuth), for the use of Agnico-Eagle Mines Ltd. (AEM), who has been party to the development of the scope of work for this project and understands its limitations. The extent to which previous investigations were relied on is detailed in the report.

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ACRONYMS

AEMP – Aquatic Effects Monitoring Program
AIC – Akaike’s Information Criterion
ANOVA – Analysis of Variance
AWPAR – All Weather Private Access Road
BACI – Before/After Control/Impact
CES – Critical effect size
CREMP – Core Receiving Environment Monitoring Program
DFO – Department of Fisheries and Oceans
DQO – Data Quality Objective
EAS – Effects Assessment Strategy
ECDF – Empirical Cumulative Distribution Function
EEM – Environmental Effects Monitoring
EIA – Environmental Impact Assessment
GPS – Global Positioning System
HEPH – Heavy Extractable Petroleum Hydrocarbons
INUG – Inuggugayualik Lake
ISQG – Interim Sediment Quality Guidelines
LEPH – Light Extractable Petroleum Hydrocarbons
LSM – Least Squared Means
MDL – Method detection limit
MMER – Metal Mining Effluent Regulations
NDEP – No deposit (referring to effluent)
NMR – No measurement required
PAHs – Polycyclic Aromatic Hydrocarbons
PDL – Pipedream Lake
PEL – Probable Effect Level
QA/QC – Quality Assurance / Quality Control



RCA – Reference Condition Approach
RPD – Relative Percent Difference
SD – Standard Deviation
SE – Standard Error
SOP – Standard Operating Procedure
SQG – Sediment Quality Guidelines
SP – Second Portage Lake
TAP – Technical Advisory Panel
TE – Tehek Lake
TEFF – Tehek Lake Farfield
TGD – Technical Guidance Document
TPE, TPN, TPS – Third Portage Lake – East, North, and South Basins
UTM – Universal Transverse Mercator
WAL – Wally Lake



EXECUTIVE SUMMARY

This report, *Environmental Effects Monitoring (EEM) Cycle 1 Interpretative Report for the Meadowbank Mine*, is submitted to meet AEM's obligations under MMER.

Study Design Overview

The August 2011 EEM biological monitoring study targeted the Third Portage Lake North (TPN) exposure area and two reference areas (Innugugayualik Lake [INUG] and Pipedream Lake [PDL]). This type of study design is classified as a multiple control-impact (MC-I) design.

Effluent Water Quality and Receiving Environment Monitoring

The “effluent” discharged at the Meadowbank Mine consists of natural lake water from isolated impoundments that are dewatered (during which exposed lake sediments become re-suspended and increase TSS) as part of mine development. There were a few minor exceedences of effluent discharge limits and some presence of sublethal toxic effects. However, EEM receiving environment monitoring generally found little trace of effluent (e.g., TSS concentrations were generally non-detectable or very low). These results were corroborated by AEM's core receiving environment monitoring program (CREMP), which targets water and sediment quality (as well as benthic invertebrates).

Fish

Due to insufficient numbers of round whitefish (and Arctic char), the originally-designed fish survey was modified in consultation with Environment Canada to focus only on a sublethal study of lake trout. No significant differences were identified for any endpoint and confidence in the results for each endpoint was high.

Recommendations for consideration in the Cycle 2 Study Design include:

- Focus on non-lethal sampling of lake trout only
- Use power analysis results to reduce lake trout sample size to minimize incidental lake trout mortality

Benthic Invertebrate Community

Differences were observed between reference and exposure areas for both benthic invertebrate community endpoints and for supporting environmental variables (i.e., physical variables that often influence benthic community structure) in the EEM data set. However, supplemental analyses were conducted using the CREMP data set (collected since 2006 and with broader spatial coverage) to verify the initial results and to evaluate causality (i.e., due to natural sediment characteristics or to effluent exposure). A weight-of-evidence framework was used to integrate the results of all analyses (i.e., EEM and CREMP). The conclusion for all benthic invertebrate community endpoints was that there were no effluent-related effects at TPN.

Recommendations for consideration in the Cycle 2 Study Design include:

- Change the number of subsample grabs per station from three to two for benthic invertebrates to be consistent with the long-term CREMP data set.

1. INTRODUCTION

1.1. Background

The Meadowbank Mine (65°N, 96°W), owned by Agnico-Eagle Mines Ltd. (AEM), is one of Canada's most northerly operating mines, located approximately 75-km north of the Hamlet of Baker Lake, Kivalliq District, Nunavut. Mine construction began in 2008 under Nunavut Water Board *A License* 2AM-Mead0815 (hereafter referred to as the "NWB A License") and Fisheries and Oceans Canada *Authorization for Works or Undertaking Affecting Fish Habitat* NU-03-0191 (hereafter referred to as the "Authorization"). Meadowbank is an open pit mine and produced its first gold bar in February 2010. Mine status formally changed to "operational" March 1, 2010.

Several lakes are located directly within and adjacent to the boundaries of the target mineral zones on the Meadowbank property. Baseline studies describing the physical, chemical, and biological characteristics of the aquatic environment in the vicinity of the project area were initiated in 1995 and continued through 2007 (BAER 2005; Azimuth 2008a,b). In addition, a comprehensive environmental impact assessment of the aquatic ecosystem (Azimuth, 2005a) and an aquatic effects management program (AEMP, 2005) were prepared to meet regulatory requirements pertaining to mine construction, operation, and closure. A range of monitoring activities have continued through the mine's construction phase and into the operational phase.

The AEMP (2005a) specifically recognized future monitoring obligations under the *Metal Mining Effluent Regulations* (MMER), which were detailed further in a companion management plan (Azimuth, 2005b). Consequently, in addition to data collected expressly to meet the requirements of MMER, there is a substantial amount of complementary data relevant to addressing MMER objectives.

Mine construction activities to date have included the isolation of portions of two lakes using dikes. Dewatering of these impoundments and future pits into adjacent lakes started in 2009. Environment Canada (EC) notified AEM that the Meadowbank Mine is subject to MMER as of December 31, 2009.

It is important to note that, at present, effluent discharge from the Meadowbank Mine consists of natural lake water being dewatered from the isolated impoundments into the north basin of Third Portage Lake. As dewatering progressed, wave action, rain events and other disturbances of the now exposed lake sediments have increased total suspended solids content of the water. Mining activities have formally been underway since March 2010; however, "mining-related" water has not been discharged into the receiving environment (e.g., water used in the mining process and contact water, which is sent to the tailings storage facility and used as reclaim water). While discharge of mining-related water will likely occur in the future (i.e., it is a component of mine development plans), this report addresses present discharges only (i.e., effluent from the dewatering process).

The focus of this report is meeting AEM's obligation to submit an Environmental Effects Monitoring (EEM) Cycle 1 Interpretative Report for the Meadowbank Mine by July 1, 2012. This document reports on the results and interpretation of the biological monitoring study

conducted in August 2011. The final study design (not including any modifications discussed in **Section 2**) was a combination of three information sources:

- 1) Environmental Effects Monitoring (EEM): Cycle 1 Study Design, Meadowbank Division, Nunavut. Report prepared by Azimuth Consulting Group Inc., Vancouver, BC for Environment Canada, Edmonton, AB on behalf of Agnico-Eagle Mines Ltd., Baker Lake, NU. December, 2010.
- 2) Environmental Effects Monitoring (EEM): Cycle 1 Study Design ADDENDUM, Meadowbank Division, Nunavut. Report prepared by Azimuth Consulting Group Inc., Vancouver, BC for Environment Canada, Edmonton, AB on behalf of Agnico-Eagle Mines Ltd., Baker Lake, NU. July, 2011.
- 3) TAP comments on the Meadowbank Gold Mine July 2011 EEM Study Design Addendum and subsequent response email from Azimuth Consulting Group. August, 2011.

The final study design (i.e., combination of all the above documents) is hereafter referred to as the Study Design and cited as Azimuth (2011).

1.2. Regulatory Context

The MMER were promulgated under the *Fisheries Act* in 2002 and have undergone several amendments, most notably in 2006 with the *Regulations Amending the Metal Mining Effluent Regulations* (hereafter referred to as the MMER, 2012). The MMER stipulate the conditions under which deleterious substances may be discharged to the aquatic environment by metal mines in Canada.

Environmental Effects Monitoring (EEM) is one of the three main components of MMER and consists of:

- **Part 1** – Effluent and water quality monitoring studies
- **Part 2** – Biological monitoring studies, including a site characterization, a fish survey, and a benthic invertebrate community survey.

The EEM Program is a cyclical receiving-environment monitoring program conducted to evaluate the effects of metal mining effluents on fish, fish habitat and the use of fisheries resources. Studies are carried out in cycles ranging from 2 to 6 years. This report has been prepared to meet the Cycle 1 interpretative report submission requirements of Part 2 of the EEM.

1.3. Objectives

The *Metal Mining Guidance Document for Aquatic Environmental Effects Monitoring* (hereafter referred to as TGD, Environment Canada, 2002) stipulates that the Cycle 1 interpretative report outline the following information:

- A description of any deviation from the study design that occurred while the biological monitoring studies were being conducted and any impact that the deviation had on the studies

- The latitude and longitude of sampling areas of sampling areas in degrees, minutes and seconds and a description of the sampling areas sufficient to identify the location of the sampling areas
- The dates and times when samples were collected
- The sample sizes
- The results of the data assessment and any supporting raw data
- The identification of any effect on the fish population, fish tissue, and the benthic invertebrate community based on the above data assessment
- A summary of the results of effluent characterization, sublethal toxicity testing and water quality monitoring reported since the day on which the mine became subject to the MMER
- The conclusions of the biological monitoring studies, taking into account the presence of other factors, the above data assessment results and QA/QC
- A description of how the results will impact the study design for subsequent biological monitoring studies
- The date when the next biological monitoring study will be conducted

The MMER (Amended to 2012) and TGD (Environment Canada, 2002; amended to 2012) cover these requirements in great detail; these information sources were relied on heavily to ensure that this Cycle 1 interpretative report meets the expectations of Environment Canada.

1.4. Organization

The remainder of this interpretative report is organized into the following sections:

- Study design update (**Section 2**)
- Effluent water quality and receiving environment monitoring (**Section 3**)
- Fish survey (**Section 4**)
- Benthic invertebrate community survey (**Section 5**)
- Conclusions from biological monitoring (**Section 6**)
- Subsequent biological monitoring (**Section 7**)
- References (**Section 8**)

2. STUDY DESIGN UPDATE

This section describes changes to the biological monitoring studies, and their consequences, relative to the Study Design document (Azimuth, 2011).

2.1. Clarification Regarding Reference Areas

As discussed in the Study Design (Azimuth, 2011), two lakes were selected as reference area lakes for biological monitoring, Inuggugayualik Lake (INUG) and Pipedream Lake (PDL). The south basin of Third Portage Lake (TPS) has also been used as a reference area for EEM-related receiving environment water quality monitoring since the program started in 2010. In an effort to retain continuity and provide compatible supporting water quality data, EEM-related receiving environment sampling was conducted at all three reference areas (i.e., INUG, PDL and TPS) in August 2011. Given the large distance to INUG and PDL and related safety concerns (i.e., requires helicopter access, which is not available throughout the year, or good snow/ice conditions for safe access by tundra buggy), future EEM-related receiving environment monitoring will continue to use TPS as a reference area.

2.2. Study Design and Data Analysis

As planned in the Study Design, both the fish and benthic invertebrate studies were conducted according to a multiple control-impact (MC-I) design. The MC-I design was selected over the more common C-I design; however, some challenges became apparent when analyzing the data from the MC-I design that are described below.

The description of an MC-I design according the technical guidance document [Chapter 8 of TGD (Environment Canada, 2012)] is as follows:

“The MC-I design is similar to the C-I design, except that it employs additional reference areas that are located in adjacent watersheds or bays where the sampled habitat is comparable to that found within the exposure area. This type of design helps to reduce problems with confounding factors (e.g., when a single reference area differs from an exposure area with respect to several environmental variables in addition to the point source effluent). Analogous to a C-I design, a significant difference between an exposure area and the mean of the reference areas, as determined by ANOVA, would represent an effect [Chapter 8 of TGD (Environment Canada, 2012)].”

The main challenge is related to how an effect should be fairly defined. The task is straightforward when the reference areas are very similar. As stated above in the TGD, the reference areas are pooled and are assumed to represent the complete population¹. Similar to the

¹ The within-pooled-reference area SD is used to derive the actual CES (critical effect size), the key source of replication is the “station”, and degrees of freedom for the C-I effect in an ANOVA would be n-2 (where “n” is the number of stations across areas).

C-I design, a significant difference between an exposure area and the mean of the pooled reference areas would represent an effect.

It is not straightforward when the reference areas differ from one another. Consider an example where the reference area means differ by a large magnitude, yet the exposure area is quite similar to one of the reference areas. In this case, assuming that exposure area mean should be equal to the pooled reference area mean does not appear appropriate (i.e., as it is ignoring natural variability among areas)². More appropriate statistical models (see footnote) work well with many areas making up the reference “population” (where it is conceptually similar to the reference condition approach [RCA]), but have severe limitations when the number of reference areas is small. Given the very low degrees of freedom, the power to detect differences is therefore substantially reduced.

To address the challenges laid out above, we propose the following hierarchical approach (see diagram in **Figure 2.2-1**) for all data assessment endpoints:

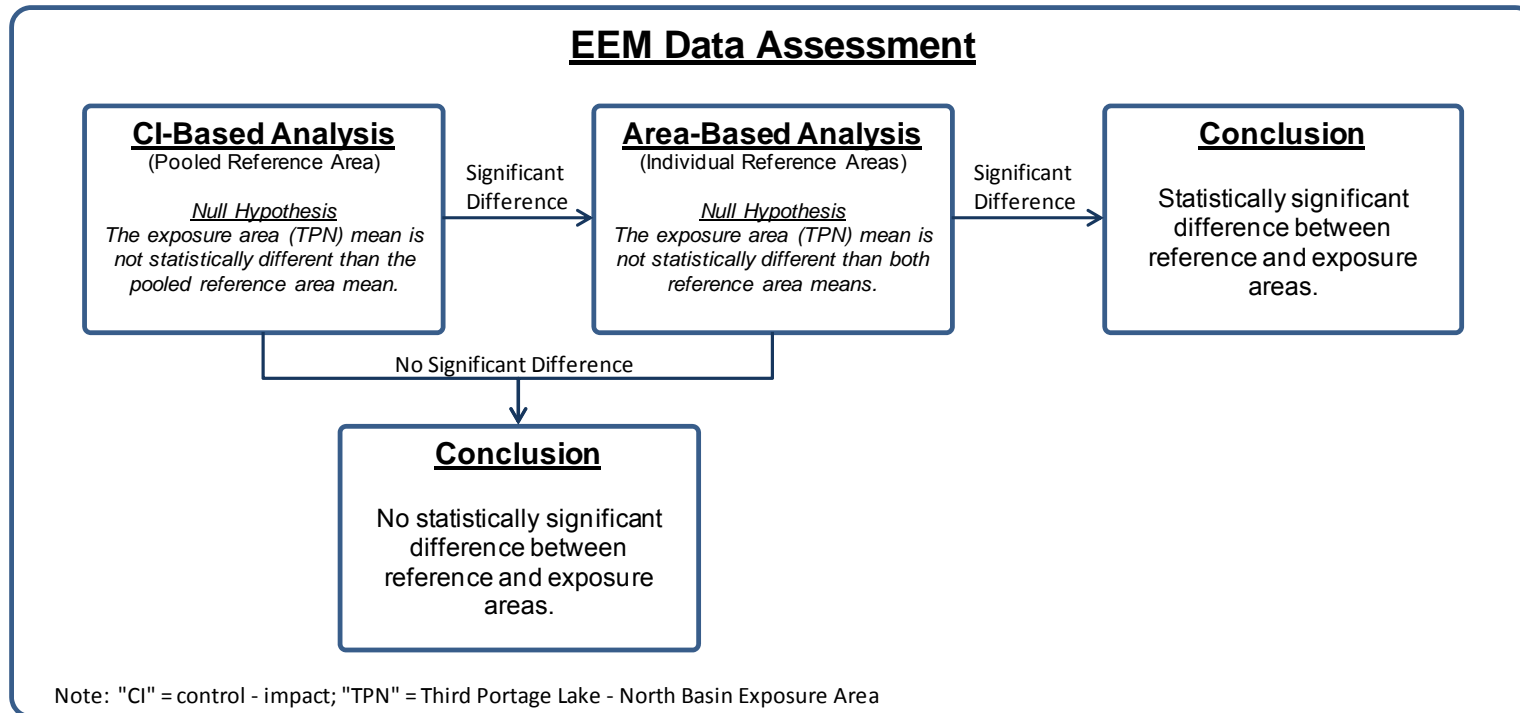
- 1) **CI-BASED**: Initial statistical analyses will be conducted using pooled reference areas (i.e., as per EEM TGD). This approach is conservative in that it assumes that there is no among-reference area variation and it sets degrees of freedom based on “stations.” The implication of this conservatism is that the Type I (i.e., false positive) error rate may be higher than the actual α of 0.1 selected for this study when the two reference areas actually differ. Where statistically significant differences between C-I are identified, those cases would also be analyzed using the second approach.
- 2) **AREA-BASED**: Secondary analyses will test for differences among individual areas. The premise here is that the exposure area must be statistically different from both reference areas in order to be classified as an effect. This approach is less conservative than the first tier in that it assumes that the exposure area mean should not be different than both reference areas individually. Given the present situation (i.e., only two reference areas), this is a reasonable assumption to identify adverse effects. The implication of this lowered conservatism is that the Type II error rate (i.e., false negative) may be higher than the actual β of 0.1 selected for this study.

The contention that Type I and Type II error rates will be “higher” in the above discussion is not due to actual differences in the α or β values used in those statistical procedures, but rather due to the differences in the questions being asked. That said, the practical hierarchical approach described above should serve to support making conclusions regarding C-I differences in this study.

² When reference areas differ, natural among-area variation should be explicitly taken into consideration and the appropriate level of replication for testing a C-I effect become “area” rather than “station”. Ideally, random-effects models (with “area” as a random effect) would be used to test whether an exposure area is different from the population of reference areas. The among-reference-areas SD (estimated for the random “area” effect) would then serve as the basis to determine the actual CES and the degrees of freedom for the C-I fixed effect would be $A-2$ (where “A” is the number of areas).

Where appropriate, and as planned, baseline data (CREMP) will be used to further explore potential C-I differences.

Figure 2.2-1. Schematic of data analysis for EEM data assessment.



2.3. Fish Survey

As described in the Study Design (Azimuth, 2011), the fish survey was planned as a non-lethal study for lake trout (*Salvelinus namaycush*) and a lethal study for round whitefish (*Prosopium cylindraceum*) (see Study Design for rationale). Baseline data suggested that obtaining the target numbers for round whitefish would be the limiting factor of our program.

Gill netting started at the exposure area TPN on August 11 and we had no trouble catching lake trout. However, far fewer round whitefish were caught than anticipated and after several days it was apparent that we would not reach target sample numbers without fishing for several weeks. In addition, lake trout mortality rates were higher than was anticipated. After consultation with Environment Canada, efforts were then also directed to Arctic char (*Salvelinus alpinus*) and trying to avoid lake trout (e.g., by focusing more on smaller mesh sizes shown to preferentially catch round whitefish and Arctic char). These changes did not alter our catch nor help reduce lake trout mortalities, which were climbing towards the *DFO License to Fish for Scientific Purposes* (**Appendix A**) permit level.

After six days, fishing was halted at TPN on August 16 (with the hope of possibly continuing focused efforts in TPN after fishing the reference areas). Although target numbers of lake trout had been exceeded for the non-lethal study, we did not achieve target numbers for round whitefish and Arctic char. Although we processed incidental mortalities of lake trout, round whitefish and Arctic char according to lethal sampling procedures, an insufficient number of sexually mature fish were captured to make any meaningful conclusions (discussed more in **Section 4**).

Fishing started at INUG and PDL on August 16 and continued for three days with large numbers of lake trout captured with higher than desirable mortality. Again, very few sexually mature round whitefish and Arctic char were caught. Given this situation and after consultation with Environment Canada, the decision was made to halt fishing pending preliminary analyses to determine if the lake trout catch at INUG and PDL was sufficient to reach targeted statistical power (i.e., $1 - \beta = 0.9$) for the non-lethal condition endpoint. Results of that analysis confirmed adequate power to detect the CES and the fish survey was terminated.

Given the paucity of sexually mature fish of any species, conclusions regarding potential effluent-related effects to fish are limited to the non-lethal endpoints of lake trout.

2.4. Benthic Invertebrate Community Survey

No changes were made from the planned study design.

3. EFFLUENT WATER QUALITY AND RECEIVING ENVIRONMENT MONITORING

As discussed in detail in the Study Design document (Azimuth, 2011), the nature of the “effluent” discharged at the Meadowbank Mine is unlike most mines subject to MMER. Meadowbank Mine effluent consisted of lake water that was pumped from an impoundment and future pit areas (with “non-contact” site drainage water that has not contacted PAG material) into the nearshore environment of TPN. Resuspension of sediments as impoundment water level dropped resulted in elevated total suspended solids (TSS) and turbidity. To minimize water column TSS, a water treatment facility was installed in to ensure that concentrations for TSS and turbidity met permit levels. “Contact” water (i.e., from plant site, pits, or any active mine areas containing potentially acid generating [PAG] material) was segregated and stored on site for use as process make-up water as required; contact water was not discharged to any receiving environment.

In addition to EEM-related receiving environment monitoring, AEM conducts routine water quality sampling as part of the Core Receiving Environment Monitoring Program (CREMP). CREMP data are used herein to complement the EEM data as needed. Apart from EEM and CREMP, AEM also conducts other dewatering-related sampling in accordance with License A; these are not reported herein.

3.1. Effluent Characterization Summary

3.1.1. Effluent Discharge Location

Effluent is monitored by AEM Meadowbank- Environment Department from a discharge point called DD-FD1, located at latitude N65 01 49.0 and longitude W96 05 25.0.

3.1.2. Effluent Monthly Loadings

MMER requires effluent quality monitoring on a weekly basis to ensure that MMER discharge limits are met. Monthly mean concentrations and associated loadings (based on total effluent volume; kg/month) are shown for 2010 and 2011 in **Table 3.1-1**. TSS exceeded the monthly limits on two occasions in 2010: in April due to a single elevated measure, and in June due to consistently elevated (~ 20 mg/L) readings throughout the month. One pH value was also below 6.0 in July 2010. All other parameters were well below monthly discharge limits for 2010 and therefore required only quarterly monitoring in 2011. TSS and pH were monitored monthly in 2011.

3.1.3. Effluent Quality Monitoring

Water quality samples have been collected from the effluent discharge four³ times each year since the mine became subject to the MMER on Jan.1, 2010. During each sampling event, the following parameters were measured in the field (temperature) or the laboratory: alkalinity⁴, aluminum, ammonia, cadmium, conductivity⁵, hardness, iron, mercury, molybdenum, nitrate and selenium.

Results of the quarterly events at the effluent discharge point are summarized in **Table 3.1-2**. Total mercury concentrations in the dewatering effluent did not exceed the tissue analysis trigger (0.1 µg/L) in 2010, but did during the August sampling event of 2011 (total mercury: 2.93 µg/L)⁶.

3.1.4. Effluent Toxicity Testing

Water samples for acute toxicity testing are conducted monthly with the following tests:

- 48-hr water flea (*Daphnia magna*) survival
- 96-hr rainbow trout (*Oncorhynchus mykiss*) survival.

Toxicity test results for acute lethality are presented in **Table 3.1-3**. Apart from an uncharacteristically low survival for the water flea, *Daphnia magna*, in the March 2010 sample (where there were no MMER water quality exceedances [see **Table 3.1-1**]), the dewatering effluent has not been acutely toxic.

Sublethal toxicity testing of effluent is conducted twice per year with the following tests:

- 72-hr freshwater alga (*Pseudokirchneriella subcapitata*) growth
- 7-day duckweed (*Lemna minor*) weight and growth
- 7-day water flea (*Ceriodaphnia dubia*) reproduction and survival
- 7-day fathead minnow (*Pimephales promelas*) growth and survival

Toxicity test results for sublethal endpoints are presented in **Table 3.1-4**. There was a general lack of sublethal effluent toxicity in 2010 with minor toxicity in 2011. Minor growth inhibition was observed for duckweed fronds (i.e., IC25 of 81.4% effluent) in the duckweed, *Lemna minor*, test for August 2010 without firsthand review of the test results. In 2011 however, *Lemna minor*

³ In 2011, the planned fourth sampling event (due December 2011) was cancelled because effluent discharge was suspended on November 15, 2011.

⁴ Alkalinity and hardness were regulated but not monitored in 2010.

⁵ Conductivity, temperature and selenium were added to the list of regulated parameters in the 2012 version of the MMER so were not monitored in 2010 and 2011

⁶ It should be noted that the “field blank” QA sample which was collected concurrently with the August, 2011 effluent sample also had an elevated mercury concentration (2.5 µg/L), suggesting these results were due to an analytical error by the laboratory.

showed effects both for weight and growth endpoints, particularly in the August sample. Also in the August 2011 sample, *Ceriodaphnia dubia* showed reproductive effects (**Table 3.1-4**). As discussed in **Section 3.2**, however, the dewatering effluent appears to rapidly mix in the receiving environment (which is confirmed by the receiving environment water quality results discussed in **Section 3.3**), so the spatial extent of those potential sublethal effects to duckweed and water fleas in the receiving environment would be very small.

Table 3.1-1. Concentrations of MMER deleterious substances in effluent discharge and loadings based on total monthly effluent volume for 2010 and 2011 (2 pages; see notes at bottom of table).

Month-Year	Total Arsenic		Total Copper		Total Cyanide		Total Lead		Total Nickel	
	Conc. (mg/L)	Loading (kg)	Conc. (mg/L)	Loading (kg)	Conc. (mg/L)	Loading (kg)	Conc. (mg/L)	Loading (kg)	Conc. (mg/L)	Loading (kg)
[Max.Mean]/Month	0.50		0.30		1.00		0.20		0.50	
January-10	0.0005	0.40	0.0012	0.97	NMR	NMR	0.0006	0.46	0.0015	1.17
February-10	0.0020	1.86	0.0010	0.93	NMR	NMR	0.0058	5.52	0.0017	1.58
March-10	0.0009	0.94	0.0012	1.30	0.0055	6.08	0.0006	0.64	0.0014	1.57
April-10	0.0109	8.87	0.0027	2.20	0.0185	15.09	0.0010	0.80	0.0022	1.81
May-10	0.0005	0.39	0.0028	2.15	0.0130	10.12	0.0003	0.23	0.0027	2.10
June-10	0.0007	0.47	0.0031	2.21	0.0124	8.91	0.0024	1.72	0.0043	3.06
July-10	0.0014	0.56	0.0012	0.50	0.0050	2.06	0.0044	1.79	0.0042	1.73
August-10	0.0011	0.48	0.0024	1.02	0.0053	2.24	0.0055	2.31	0.0037	1.54
September-10	0.0011	0.23	0.0010	0.21	0.0050	1.03	0.0010	0.21	0.0051	1.06
October-10	0.0036	0.58	0.0019	0.30	0.0133	2.16	0.0030	0.48	0.0087	1.41
November-10	0.0029	0.43	0.0011	0.16	0.0150	2.21	0.0004	0.05	0.0090	1.33
December-10	0.0025	0.87	0.0013	0.46	0.0162	5.72	0.0003	0.11	0.0096	3.38
January-11	0.0026	0.63	0.0007	0.17	0.0290	7.07	0.0013	0.30	0.0086	2.10
February-11	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP
March-11	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR
April-11	0.0021	0.87	0.0005	0.21	0.0680	28.20	0.0003	0.12	0.0102	4.23
May-11	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR
June-11	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR
July-11	0.0005	0.14	0.0005	0.14	0.0670	19.14	0.0003	0.09	0.0037	1.06
August-11	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR
September-11	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR
October-11	0.0207	6.40	0.0029	0.90	0.0050	1.55	0.0048	1.48	0.0098	3.03
November-11	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR	NMR
December-11	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP

Month-Year	Radium-226		Total suspended Solids		Total Zinc		pH		Total Effluent Volume (m ³)
	Conc. (Bq/L)	Loading (kg)	Conc. (mg/L)	Loading (kg)	Conc. (mg/L)	Loading (kg)	Highest	Lowest	
[Max.Mean]/Month	0.37		15.00		0.50		9.5	6.0	
January-10	0.004	2.83	9.0	7276	0.002	1.41	7.42	7.06	808439
February-10	0.005	5.02	7.0	6691	0.004	4.06	7.24	6.76	955861
March-10	0.004	4.42	8.5	9390	0.003	3.59	7.08	6.87	1104674
April-10	0.014	11.21	21	17330	0.010	8.36	7.75	6.80	815525
May-10	0.007	5.45	13	10118	0.005	3.63	6.98	6.43	778312
June-10	NMR	NMR	22	15812	0.007	4.74	6.97	6.36	718745
July-10	0.013	5.35	12	4858	0.002	0.82	6.94	5.85	411657
August-10	NMR	NMR	6.7	2800	0.011	4.62	6.53	6.28	420064
September-10	NMR	NMR	8.8	1806	0.003	0.62	6.68	6.37	206366
October-10	0.006	0.97	6.7	1081	0.012	1.89	7.00	6.54	162119
November-10	NMR	NMR	2.7	392	0.001	0.20	6.73	6.37	147029
December-10	NMR	NMR	5.8	2046	0.005	1.69	7.07	6.44	352792
January-11	0.071	17.32	4.0	976	0.004	0.98	6.80	6.45	243891
February-11	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP
March-11	NMR	NMR	7.3	1731	NMR	NMR	6.76	6.43	236101
April-11	0.067	27.79	4.5	1866	0.001	0.41	6.76	6.50	414718
May-11	NMR	NMR	6.0	2651	NMR	NMR	6.92	6.55	441824
June-11	NMR	NMR	**	**	NMR	NMR	**	**	144813
July-11	0.025	7.14	9.0	2571	0.001	0.29	6.75	6.31	285660
August-11	NMR	NMR	4.4	642	NMR	NMR	7.30	6.76	145996
September-11	NMR	NMR	5.3	420	NMR	NMR	7.25	7.08	78668
October-11	0.100	30.90	2.6	803	0.001	0.31	7.29	6.62	309007
November-11	NMR	NMR	6.5	2112	NMR	NMR	7.00	6.78	324850
December-11	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP	NDEP

Notes: Shaded values exceed the maximum monthly mean concentrations; NDEP (No Deposit) = measurement not taken because there was no deposit from the final discharge point; NMR (No Measurement Required) = measurement not taken in accordance with the conditions set out in section 12 or 13 of the MMER; ** June 2011 samples for TSS and pH were erroneously not collected; From January 1 to October 23, 2011 the water originated from Second Portage Lake Impoundment Area; From October 22 to November 14, 2011 the water originated from Bay Goose Impoundment Area.

Table 3.1-2. Concentrations of EEM substances in effluent discharge, monitored four times per year for 2010 and 2011.

Parameter	Effluent Discharge Point (DD-FD1)						
	2010				2011		
	19-Apr-10	24-Aug-10	27-Oct-10	29-Nov-10	10-Jan-11	11-Apr-11	24-Aug-11
Alkalinity (mg CaCO ₃ /L)	**	**	**	**	**	45	82
Aluminium-Total (mg/L)	0.630	0.484	1.50	0.619	0.857	1.10	0.541
Ammonia-Total (mg N/L)	0.17	0.47	2.7	4.8	4.8	6.7	<0.05
Cadmium-Total (mg/L)	< 0.00008	**	0.00013	< 0.00008	<0.00008	<0.00008	0.00025
Hardness (mg CaCO ₃ /L)	**	**	**	**	**	215	200
Iron-Total (mg/L)	0.15	0.33	0.51	<0.01	<0.01	<0.01	0.69
Mercury-Total (mg/L)	< 0.00001	< 0.00001	< 0.00001	0.00001	<0.00001	<0.00001	0.00293 ¹
Molybdenum-Total (mg/L)	0.0014	0.0036	0.0069	0.0112	0.0116	0.0148	0.0329
Nitrate-Total (mg N/L)	0.52	**	1.2	1.7	1.1	1.0	1.6

Notes: ** Parameter not measured due to accidental omission on lab analysis request form; Parameters that have been added to the required monitoring list include: field measured conductivity and temperature and laboratory measured total selenium (MMER, 2012); ¹ It should be noted that the “field blank” QA sample which was collected concurrently with the August, 2011 effluent sample also had an elevated mercury concentration (2.5 µg/L), suggesting these results were due to an analytical error by the laboratory.

Table 3.1-3. MMER acute toxicity results in effluent discharge, monitored monthly for 2010 and 2011.

Sampling Month-Year	Effluent Discharge Point (DD-FD1)	
	<i>Daphnia magna</i> LC50 (48hr) % v/v	<i>Oncorhynchus mykiss</i> LC50 (96hr) % v/v
January-10	>100	>100
February-10	>100	>100
March-10	46.2	>100
April-10	>100	>100
May-10	>100	>100
June-10	>100	>100
July-10	>100	>100
August-10	>100	>100
September-10	>100	>100
October-10	>100	>100
November-10	>100	>100
December-10	>100	>100
January-11	>100	>100
February-11	NDEP	NDEP
March-11	NDEP	NDEP
April-11	>100	>100
May-11	NMR	NMR
June-11	NDEP	NDEP
July-11	>100	>100
August-11	NMR	NMR
September-11	NMR	NMR
October-11	>100	>100
November-11	NMR	NMR
December-11	NDEP	NDEP

Notes: "*Daphnia magna*" is an aquatic invertebrate commonly called 'water flea'; "*Oncorhynchus mykiss*" is a fish commonly called 'rainbow trout'; "LC50" = lethal concentration (of effluent) resulting in 50% decrease in survival relative to controls; NDEP (No Deposit) = measurement not taken because there was no deposit from the final discharge point; NMR (No Measurement Required) = measurement not taken in accordance with the conditions set out in section 12 or 13 of the MMER.

Table 3.1-4. EEM sublethal toxicity results in effluent discharge, monitored two times per year for 2010 and 2011.

Sublethal Toxicity Test -Endpoint (% v/v)	Effluent Discharge Point (DD-FD1)			
	2010		2011	
	19-Apr-10	24-Aug-10	10-Jan-11	25-Aug-11
<i>Pseudokirchneriella subcapitata</i> -IC25 growth	>90.9	>90.9	>90.9	>90.9
<i>Lemna minor</i> -IC25 weight	>97	>97	76.2	1.81
<i>Lemna minor</i> -IC25 growth (frond #)	>97	81.4	46.0	1.77
<i>Ceriodaphnia dubia</i> -IC25 reproduction	>100	>100	62.5	10.8
<i>Ceriodaphnia dubia</i> -LC50 survival	>100	>100	>100	66.0
<i>Pimephales promelas</i> - IC25 growth	>100	>100	>100	91.5
<i>Pimephales promelas</i> -LC50 survival	>100	>100	>100	>100

Notes: "IC25" = inhibitory concentration (of effluent) resulting in 25% decrease in response relative to controls; "LC50" = lethal concentration (of effluent) resulting in 50% decrease in survival relative to controls; "*Pseudokirchneriella subcapitata*" is a freshwater alga; "*Lemna minor*" is a freshwater aquatic plant commonly called 'duckweed'; "*Ceriodaphnia dubia*" is an aquatic invertebrate commonly called 'water flea'; "*Pimephales promelas*" is a fish commonly called 'fathead minnow'

3.2. Plume Delineation

As presented in the in the Study Design document (Azimuth, 2011), effluent mixing was modeled by Golder Associates to estimate the location of the 1% dilution boundary defining the TPN exposure area (**Figure 3.2-1**).

Given that the “effluent” consists largely lake water transferred from one basin to another with some TSS, there were no suitable tracers that could provide real-time information for characterizing percent dilution in the receiving environment. While turbidity could be used to qualitatively track the plume, it does not translate well into percent effluent concentration. Notwithstanding, the strategy for plume delineation discussed in the Study Design was to try to roughly map the plume using turbidity with the intent of providing some site-specific characterization to complement the modeled plume results.

A reconnaissance helicopter flight was taken on August 10, 2011 to obtain a “birds-eye view” of the discharge area. As seen by **Photos 3.2-1 through 3.2-3**, apart from some minor signs of surface water flow (i.e., that discharge was occurring), there were no other visual signs of the plume from the air (e.g., turbid water).

The initial plume delineation survey was conducted on August 11, 2011 the first day of fish sampling at TPN. The plan was to conduct turbidity profiling (using the McVan Analite NEP160-3-05R portable turbidity meter/logger with a high sensitivity NEP260 90° probe) over a grid (**Figure 3.2-2**), taking measurements every two meters from the surface to 1m off the bottom. Sampling started at TPN-1 to characterize ambient conditions well away from the effluent discharge location. Background turbidity ranged from 0.11 to 0.37 NTU⁷, which roughly corresponds to 0.1 to 0.3 mg/L TSS⁸. The survey then moved in to characterize the closest stations to the discharge point, stepping out as needed until ambient turbidity levels were found. TPN-4 and TPN-6 (ranging from 0 to 0.22 NTU or 0 to 0.2 mg/L TSS) were consistent with background turbidity levels. TPN-5, located approximately 75 m south of the discharge point, but in line with the prevailing winds, had slightly elevated turbidity (0.56 to 0.94 NTU or 0.4 to 0.6 mg/L TSS) in the top two meters. No further sampling was conducted given that the “plume” was bounded.

Delineation of the discharge location in a grid pattern during the August 11 turbidity survey confirmed that discharge was occurring (**Photos 3.2-4 and 3.2-5**), but that the effluent was quite clear (**Photo 3.2-6**). As these conditions did not change while conducting the biological sampling at TPN (August 11 – 16th), no further turbidity surveys were needed.

The plume delineation survey represents a snapshot of conditions that cannot be assumed to be representative. As seen in **Table 3.1-1**, monthly discharge volumes have been much higher

⁷ Even after calibration, the turbidity meter was reading slightly negative values in the field. Consequently, the values were corrected by adding the most negative reading (0.4 NTU).

⁸ Conversion based on a relationship developed to support dike construction monitoring at Meadowbank. $TSS = 10^{((\text{Log}(\text{NTU}) - 0.2729)/1.2002)}$ (Azimuth, 2010).

previously than in August 2011. Consequently, for the purposes of defining the exposure zone for this study, we will use the modeled 1% dilution zone from the Golder study (see **Figure 3.2-1**).

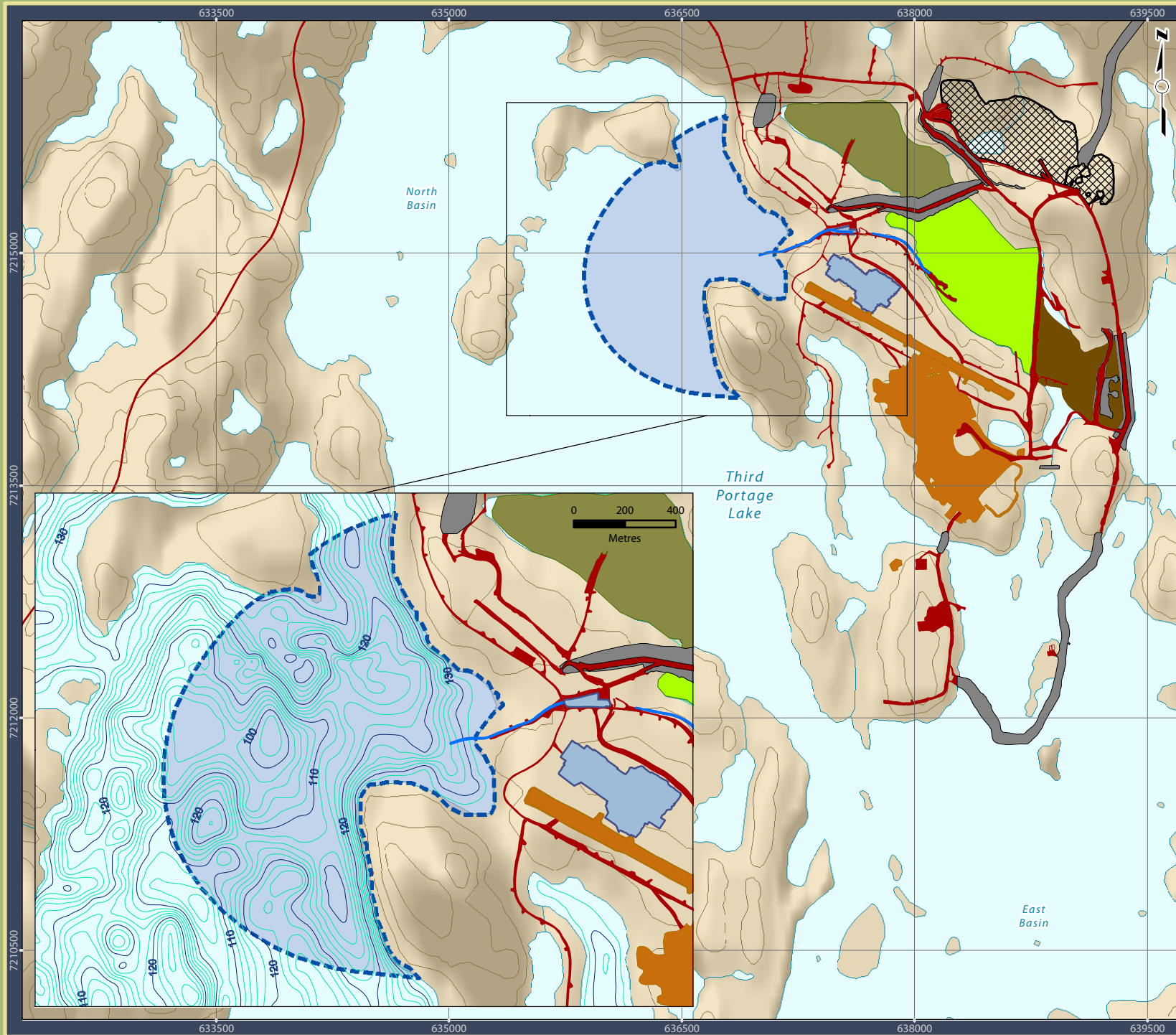


Figure 3.2-1. Dewatering Discharge Outfall and Exposure Area

Legend

- Bathymetry 2m*
- Bathymetry 10m**
- All Weather Private Access Road
- Mine Features**
- Effluent Discharge (Dewatering Pipeline)
- Facilities
- Road
- Dike
- Waste Dump
- Dewatered Lake
- Portage Attenuation Facility
- Tailings Storage Facility
- Water Treatment Facility
- 1% Effluent Exposure Area Delineation

* 2m depth = below lake surface
 ** 10m depth = above sea level

Area of Detail



0 250 500 750 1,000
 Metres

Projection: UTM Zone 14 NAD83

Data Sources:

Natural Resources Canada, GeoBase®
 National Topographic Database
 Agnico-Eagle Mines Limited.
 Azimuth Consulting Group Inc.

Meadowbank Gold Project

Prepared for:



By:



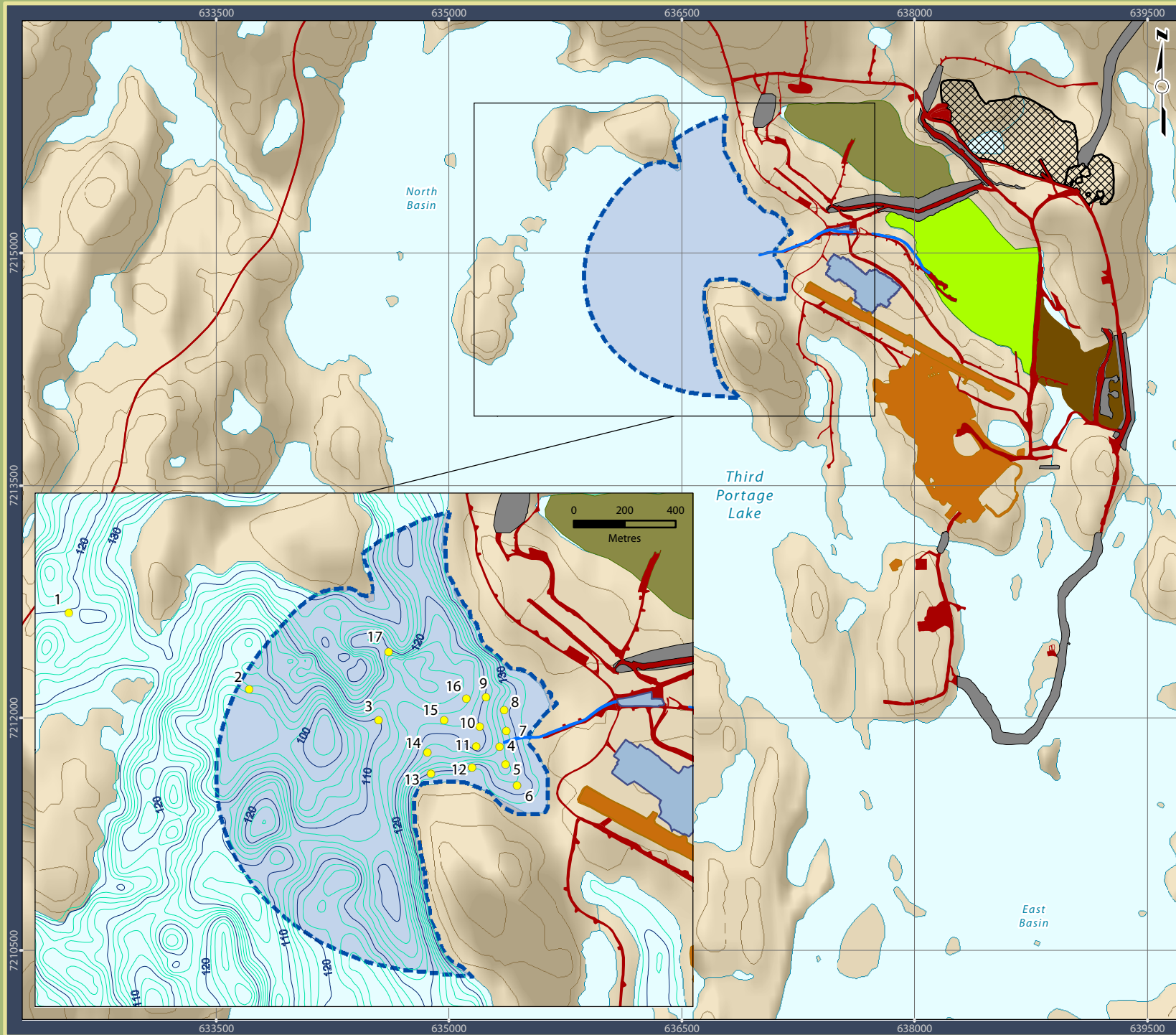


Figure 3.2-2.
Plume Delineation Survey:
Turbidity Profiling Locations

Legend

- Turbidity Profile Location
- Bathymetry 2m*
- Bathymetry 10m**
- All Weather Private Access Road
- Mine Features**
- Effluent Discharge (Dewatering Pipeline)
- Facilities
- Road
- Dike
- Waste Dump
- Dewatered Lake
- Portage Attenuation Facility
- Tailings Storage Facility
- Water Treatment Facility
- 1% Effluent Exposure Area Delineation

* 2m depth = below lake surface
 ** 10m depth = above sea level

Area of Detail



0 250 500 750 1,000
 Metres

Projection: UTM Zone 14 NAD83

Data Sources:

Natural Resources Canada, GeoBase®
 National Topographic Database
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Meadowbank Gold Project

Prepared for:



By:



Photo 3.2-1. “Birds-eye view” of the discharge area in TPN showing no visual signs of a turbidity plume from the air.



Photo 3.2-2. “Birds-eye view” of the discharge area in TPN showing no visual signs of a turbidity plume from the air.



Photo 3.2-3. “Birds-eye view” of the discharge area in TPN showing no visual signs of a turbidity plume from the air.



Photo 3.2-4. A closer examination of the discharge location from the August 11, 2011 turbidity survey.



Photo 3.2-5. A closer examination of the discharge location from the August 11, 2011 turbidity survey.



Photo 3.2-6. A view of the clarity of the effluent being discharged during the August 11, 2011 turbidity survey.



3.3. Receiving Environment Monitoring

3.3.1. Water Chemistry – EEM

Water quality samples have been collected for receiving environment monitoring from the exposure area (TPN) and the reference area (TPS) four times in each of the two years since the mine became subject to the MMER (i.e., since Jan.1, 2010), in addition to ‘baseline’ data at this location since 2002. Water quality samples were also collected from the two biological monitoring study reference areas (INUG and PDL) in August 2011. To put water chemistry results in perspective (particularly for August when biological parameters were also collected) **Table 3.3-1** provides a daily summary of effluent volume discharge levels and climate conditions for the month of August 2011. Coordinates for water quality sampling locations are shown in **Table 3.3-3**. All samples were collected according to the guidelines of the existing CREMP water sampling SOP (**Appendix B**) and according to the QA/QC procedures presented in the Study Design document (Azimuth, 2011). Field data sheets showing profiles from August 2011 are presented in **Appendix C**.

During each sampling event, the following parameters were measured in the field (pH, temperature, oxygen, conductivity) and the laboratory: alkalinity, aluminum, ammonia, arsenic, cadmium, conductivity⁹, copper, cyanide, hardness, iron, lead, mercury, molybdenum, nickel, nitrate, pH, radium 226, selenium⁴, TSS, and zinc.

Results of the quarterly events at TPN and TPS are summarized in **Table 3.3-2** (values taken directly from AEM’s annual MMER report; EEM data quality objectives assumed to have been met). Results from the August 2011 event (TPN, TPS, INUG, and PDL) are compiled in **Table 3.3-3** (see **Section 3.3.2** for information regarding data quality). Sample locations for TPN and TPS are the same for all sampling periods. All values are screened against CCME water quality guidelines for the protection of aquatic life (CCME, 2007). Noteworthy results include:

- TSS was typically < 1 mg/L (the laboratory MDL) and never exceeded 2 mg/L.
- Total cyanide exceeded the CCME guideline at TPN (Apr 2010; also detected [but <CCME] in Dec 2011), TPS (Apr 2010; also detected [but <CCME] in Jan 2011 and Aug 2011), INUG (Aug 2011), and PDL (Aug 2011). Given the pattern (e.g., higher at the distant reference area) and nature (occurring when TSS < 1mg/L) of results, they are likely anomalous and unrelated to effluent discharge.
- Total lead exceeded the CCME guideline at TPN (May 2010 and Dec 2011) and TPS (May 2010, Aug 2010, Jan 2011). Again, since the reference area TPS generally had higher concentrations and TSS was generally less than 1 mg/L, these results do not appear related to effluent discharge.

⁹ Conductivity and total selenium were added to the list of regulated parameters in the 2012 version of the MMER so were not monitored in 2010 and 2011.

- Total copper was elevated at TPN and TPS in August 2011. Given that TSS was detected in both samples (2 mg/L at TPN and 1 mg/L at TPS), this result is likely due to the detection of particulate-associated copper. Given that turbidity was largely at background levels during plume delineation surveys conducted in August 2011 (see **Section 3.2**), and that the CREMP results (see **Section 3.3.2**) show non-detectable TSS concentrations at both areas, these results are unlikely related to effluent discharge.
- Total aluminum, copper, lead and zinc exceeded CCME guidelines at TPN in December 2011, despite TSS < 1 mg/L. The aluminum result was due to quite low pH (i.e., the lower 0.005 mg/L guideline is triggered at pH < 6.5) in that sample. These results are not consistent with the CREMP results (see **Section 3.3.2**), so suggest either a highly localized (in space and time) influence of effluent discharge at TPN or some other anomaly. Regardless of the reason, this event occurred after the August 2011 EEM biological monitoring study, so would not be a factor.

3.3.2. Water Chemistry – CREMP

Details regarding CREMP water quality results are provided in Azimuth (2012a). In short, the CREMP sampling methods are the same as those used for EEM receiving environment monitoring (see SOP in **Appendix B**). The TPN area boundaries, however, are broader in the CREMP, extending across much of the north basin of Third Portage Lake (**Figure 3.3-1**).

Water samples were analyzed for a comprehensive list of parameters, as presented in the Study Design document (Azimuth, 2011). Data quality was assessed and met the stipulated objectives of the CREMP (see Azimuth, 2012a for details). Results for INUG, PDL, TPN, and TPS (focusing on the four areas related to EEM) are shown for the August 2011 CREMP sampling event and screened against CCME guidelines in **Table 3.3-4**. Key results related to EEM are presented below, based on all historical CREMP data:

- *TSS* – concentration at all four areas were typically below the MDL (**Figure 3.3-2**). Detectable concentrations (1 to 2 mg/L) found occasionally at all areas except PDL. These results corroborate the EEM findings that TSS concentrations are typically less than 1 mg/L. See TSS EAS report (Azimuth, 2012b) for further detail.
- *Aluminum* – concentrations were consistently low across all four areas, with no apparent pattern related to effluent discharge at TPN (**Figure 3.3-3**). Note that the Dec 2011 results for TPN did not exceed the CCME and that pH was near 7 (**Figure 3.3-4**).
- *Copper* – concentrations were consistently low across all four areas, suggesting that effluent discharge is not affecting water quality (**Figure 3.3-5**). The August 2011 sampling event showed no indication of elevated copper at either TPN or TPS.
- *Lead* – concentrations were typically low (<MDL), with no patterns related to effluent discharge (**Figure 3.3-6**).

3.3.3. Sediment Chemistry

The comprehensive CREMP sediment monitoring data were used to support the EEM study (i.e., no EEM-specific sediment quality monitoring was conducted). Samples were collected according to the existing CREMP sediment (and benthos) sampling SOP (**Appendix D**) and according to the QA/QC procedures presented in the Study Design document (Azimuth, 2011). In short, five replicate samples were collected from each area (synoptically with benthic invertebrate sampling) in August, 2011 using a Petite Ponar grab.

Sampling area locations are shown in **Figure 3.3-7** and results for INUG, PDL, and TPN are presented in **Table 3.3-5** for August 2011, screened against CCME guidelines. Data quality was assessed and met the stipulated objectives of the CREMP (see Azimuth, 2012a for details). Field data sheets for August 2011 sampling are presented in **Appendix E**.

CREMP sediment chemistry monitoring has historically included both grab and core samples. Grab samples, while providing sediment chemistry context for historical deposition patterns, are not ideal compared to core samples for assessing possible subtle changes over time. Grab samples are useful for the collection of sediments to characterize substrate conditions in support benthic invertebrate community surveys.

A comprehensive review of temporal and spatial patterns in the existing sediment chemistry data set for Meadowbank conducted as part of last year's CREMP did not identify any apparent changes in metals concentrations related to effluent discharge to TPN. Further detail regarding CREMP sediment quality sampling is provided in Azimuth (2012a).

Based on the preceding information, the grab sample data for physical parameters (i.e., total organic carbon and particle size) were used as supporting data to help interpret the benthic invertebrate community assessment (see **Section 5**).

Table 3.3- 1. Daily effluent discharge volume and climate conditions for August, 2011.

Days during August 2011	Effluent Volume (m ³)	Climate Conditions ¹				
		Temperature Range (°C)	Temperature Mean (°C)	Precipitation Total (mm)	Wind Speed Mean (km/h)	Wind Speed Max gust (km/h)
1	13706	5.8 - 17.1	11.5	0	17.0	39
2	0	9.4 - 11.3	10.4	8.8	41.5	70
3	0	9.9 - 12.5	11.2	3.5	27.7	46
4	0	8.7 - 14.6	11.7	0	26.0	46
5	4090	6.2 - 19.1	12.7	0	21.4	37
6	18910	10.8 - 21.8	16.3	0	18.3	33
7	24380	8.3 - 19.1	13.7	0	19.8	35
8	13810	8.5 - 19.8	14.2	0	19.1	37
9	1700	8.2 - 17.1	12.7	0	16.9	39
10	5620	9.8 - 14.6	12.2	0.9	13.7	32
11	5050	8.4 - 15.0	11.7	1.8	16.8	<31
12	6600	9.0 - 15.6	12.3	0	14.9	35
13	5015	8.9 - 17.1	13.0	0	18.5	37
14	0	7.5 - 15.6	11.6	0	26.5	43
15	5015	2.9 - 15.7	9.3	0	9.8	<31
16	0	8.6 - 14.2	11.4	0	23.8	48
17	3010	7.9 - 10.7	9.3	2	34.8	54
18	0	8.0 - 10.8	9.4	0	33.5	56
19	4050	8.4 - 11.9	10.2	2.9	20.7	35
20	6620	8.2 - 14.9	11.6	0	12.0	32
21	0	4.8 - 17.8	11.3	0	18.0	37
22	1780	3.0 - 12.9	8.0	0	20.9	44
23	2130	8.7 - 13.8	11.3	0.9	25.4	43
24	5340	8.5 - 12.4	10.5	0	10.0	<31
25	2280	8.7 - 10.9	9.8	0	18.8	<31
26	1130	8.9 - 12.0	10.5	0	19.8	37
27	2040	7.1 - 14.6	10.9	0.9	22.8	41
28	5990	3.7 - 12.8	8.3	1.3	16.6	39
29	2170	3.1 - 10.6	6.9	0.3	16.5	39
30	1400	4.7 - 13.0	8.9	0	11.7	<31
31	4160	5.5 - 10.0	7.8	1.2	19.8	39

Notes: ¹ Climate conditions were taken from Environment Canada's website and are for Baker Lake, Nunavut (75 km from the mine).

Table 3.3- 2. EEM water quality results at TPN and TPS, monitored four times per year for 2010 and 2011 (2 pages; see notes at bottom of table).

Parameter	CCME (2007)	2010				2011			
	Guideline ¹	19-Apr-10	22-May-10	28-Jul-10	31-Aug-10	17-Jan-11	12-Apr-11	24-Aug-11	20-Dec-11
TPN (Exposure Area)									
Alkalinity (mg CaCO ₃ /L)	NG	6	6	40	6	25	15	10	<2
Aluminium-Total (mg/L) ²	0.005 - 0.100	0.026	0.027	0.019	0.010	0.016	0.149	0.009	0.060
Ammonia-Total (mg N/L) ^{2,3}	0.832 - 25.9	0.16	0.31	0.07	0.06	<0.05	0.25	<0.05	<0.05
Arsenic-Total (mg/L)	0.0050	0.0023	<0.0005	<0.0005	<0.0005	<0.0005	0.0008	<0.0005	<0.0005
Cadmium-Total (mg/L) ⁴	0.0000025 - 0.0000046	<0.00008	<0.00008	<0.00008	<0.00008	<0.00008	<0.00008	<0.00002	<0.00002
Copper-Total (mg/L) ⁴	0.002 - 0.002	<0.0005	<0.0005	<0.0005	0.0006	<0.0005	<0.0005	0.0271	0.0021
Cyanide-Total (mg/L)	0.005	0.008	**	<0.005	<0.005	<0.005	<0.005	<0.005	0.005
Dissolved oxygen-Field (mg/L)	NG	14.0	11.3	**	10.6	12.4	19.5	9.2	25.3
Hardness (mg CaCO ₃ /L)	NG	5	6	5	5	6	10	7	10
Iron-Total (mg/L)	0.3	<0.01	<0.01	<0.01	<0.01	0.03	<0.01	<0.01	<0.01
Lead-Total (mg/L) ⁴	0.001 - 0.001	<0.0003	0.0068	<0.0003	<0.0003	<0.0003	<0.0003	<0.0003	0.0109
Mercury-Total (mg/L)	0.000026	<0.00001	<0.00001	<0.00001	<0.00001	<0.00001	<0.00001	<0.00001	<0.00001
Molybdenum-Total (mg/L)	0.073	<0.0005	<0.0005	<0.0005	<0.001	<0.0005	<0.0005	0.0016	<0.0005
Nickel-Total (mg/L) ⁴	0.025 - 0.025	<0.0005	0.0006	0.0012	0.0013	<0.0005	<0.0005	0.0014	0.0006
Nitrate-Total (mg N/L)	2.9	0.01	0.08	0.03	0.03	0.01	0.03	0.05	0.03
pH-Field	6.5 - 9.0	6.88	**	6.72	6.88	7.12	7.17	7.59	6.22
Radium-226 (Bq/L)	NG	<0.002	<0.002	<0.003	<0.003	<0.002	<0.002	<0.002	0.003
Temperature-Field (°C)	NG	0.47	0.74	7.75	9.16	0.31	0.17	10.4	0.38
Total suspended solid (mg/L)	NG	<1	<1	<1	<1	1	<1	2	<1
Zinc-Total (mg/L)	0.030	<0.001	0.006	<0.001	<0.001	<0.001	0.009	<0.001	0.035

	CCME (2007)	2010				2011			
Parameter	Guideline ¹	19-Apr-10	22-May-10	28-Jul-10	31-Aug-10	17-Jan-11	12-Apr-11	24-Aug-11	20-Dec-11
TPS (Reference Area)									
Alkalinity (mg CaCO ₃ /L)	NG	7	4	27	5	18	12	9	<2
Aluminium-Total (mg/L) ²	0.100 - 0.100	0.019	0.024	<0.002	0.018	0.046	0.033	<0.006	0.030
Ammonia-Total (mg N/L) ^{2,3}	0.832 - 8.24	0.17	0.29	0.07	0.06	<0.05	<0.05	<0.05	<0.05
Arsenic-Total (mg/L)	0.0050	0.0033	<0.0005	<0.0005	<0.0005	<0.0005	<0.0005	<0.0005	<0.0005
Cadmium-Total (mg/L) ⁴	0.0000021 - 0.0000050	<0.00008	<0.00008	<0.00008	<0.00008	<0.00008	<0.00008	<0.00002	<0.00002
Copper-Total (mg/L) ⁴	0.002 - 0.002	<0.0005	<0.0005	<0.0005	<0.0005	0.0009	<0.0005	0.0131	0.0010
Cyanide-Total (mg/L)	0.005	0.020	**	<0.005	<0.005	0.005	<0.005	0.005	<0.005
Dissolved oxygen-Field (mg/L)	NG	12.5	11.2	**	10.4	11.9	16.8	9.2	16.8
Hardness (mg CaCO ₃ /L)	NG	4	5	5	5	6	5	6	11
Iron-Total (mg/L)	0.3	<0.01	<0.01	<0.01	<0.01	0.25	<0.01	<0.01	<0.01
Lead-Total (mg/L) ⁴	0.001 - 0.001	<0.0003	0.0108	<0.0003	0.0069	0.0124	<0.0003	0.0005	<0.0003
Mercury-Total (mg/L)	0.000026	<0.00001	<0.00001	<0.00001	<0.00001	<0.00001	<0.00001	<0.00001	<0.00001
Molybdenum-Total (mg/L)	0.073	<0.0005	<0.0005	<0.0005	<0.0005	<0.0005	<0.0005	0.0005	<0.0005
Nickel-Total (mg/L) ⁴	0.025 - 0.025	<0.0005	0.0005	<0.0005	0.0013	<0.0005	<0.0005	0.0008	<0.0005
Nitrate-Total (mg N/L)	2.9	<0.01	<0.01	0.03	0.05	0.01	0.08	0.01	0.03
pH-Field	6.5 - 9.0	7.08	**	7.06	6.91	6.81	7.12	7.92	7.91
Radium-226 (Bq/L)	NG	<0.002	0.003	<0.003	0.003	<0.002	0.002	<0.002	0.003
Temperature-Field (°C)	NG	0.52	0.22	9.22	9.37	0.37	0.36	10.7	0.42
Total suspended solid (mg/L)	NG	<1	<1	1	<1	<1	<1	1	<1
Zinc-Total (mg/L)	0.030	<0.001	0.006	0.003	<0.001	0.004	0.003	<0.001	0.006

Notes: NG = no guideline; ¹ CCME (Canadian Council of Ministers of the Environment) Canadian Water Quality Guidelines for the Protection of Aquatic Life, 1999, updated December 2007; ² Guideline is pH dependent; ³ Guideline is temperature dependent; ⁴ Guideline is hardness dependent; **Parameter not measured due to malfunctioning field equipment or accidental omission on lab analysis request form; Shaded values exceed the CCME guideline; Parameters that have been added to the required monitoring list include: field measured conductivity and laboratory measured total selenium (MMER, 2012).

Table 3.3- 3. EEM water quality results at INUG, PDL, TPN and TPS, monitored at the same time that the biological monitoring studies were conducted (August, 2011).

Area		INUG	PDL	TPN	TPS
Date		18-Aug-11	18-Aug-11	24-Aug-11	24-Aug-11
Latitude (dd mm ss.s)	CCME (2007)	N65 03 08.1	N65 06 17.8	N65 01 51.8	N64 58 10.0
Longitude (dd mm ss.s)	Guideline ¹	W96 23 22.5	W96 13 05.2	W96 05 33.3	W96 09 55.8
Alkalinity (mg CaCO ₃ /L)	NG	10	24	10	9
Aluminium-Total (mg/L) ²	0.100 - 0.100	0.010	0.007	0.009	<0.006
Ammonia-Total (mg N/L) ^{2,3}	0.274 - 0.832	<0.05	<0.05	<0.05	<0.05
Arsenic-Total (mg/L)	0.0050	<0.0005	<0.0005	<0.0005	<0.0005
Cadmium-Total (mg/L) ⁴	0.0000011 - 0.0000034	<0.00002	<0.00002	<0.00002	<0.00002
Copper-Total (mg/L) ⁴	0.002 - 0.002	<0.0005	<0.0005	0.0271	0.0131
Cyanide-Total (mg/L)	0.005	0.016	0.007	<0.005	0.005
Dissolved oxygen-Field (mg/L)	NG	8.5	8.8	9.2	9.2
Hardness (mg CaCO ₃ /L)	NG	2	4	7	6
Iron-Total (mg/L)	0.3	<0.01	<0.01	<0.01	<0.01
Lead-Total (mg/L) ⁴	0.001 - 0.001	0.0006	<0.0003	<0.0003	0.0005
Mercury-Total (mg/L)	0.000026	**	**	<0.00001	<0.00001
Molybdenum-Total (mg/L)	0.073	<0.0005	<0.0005	0.0016	0.0005
Nickel-Total (mg/L) ⁴	0.025 - 0.025	<0.0005	<0.0005	0.0014	0.0008
Nitrate-Total (mg N/L)	2.9	<0.01	<0.01	0.05	0.01
pH-Field	6.5 - 9.0	7.82	8.32	7.59	7.92
Radium-226 (Bq/L)	NG	<0.002	<0.002	<0.002	<0.002
Temperature-Field (°C)	NG	12.0	11.4	10.4	10.7
Total suspended solid (mg/L)	NG	<1	<1	2	1
Zinc-Total (mg/L)	0.030	<0.001	<0.001	<0.001	<0.001

Notes: NG = no guideline; ¹ CCME (Canadian Council of Ministers of the Environment) Canadian Water Quality Guidelines for the Protection of Aquatic Life, 1999, updated December 2007; ² Guideline is pH dependent; ³ Guideline is temperature dependent; ⁴ Guideline is hardness dependent; **Parameter not measured due to malfunctioning field equipment or accidental omission on lab analysis request form; Shaded values exceed the CCME guideline; Parameters that have been added to the required monitoring list include: field measured conductivity and laboratory measured total selenium (MMER, 2012).

Table 3.3- 4. CREMP water quality results at INUG, PDL, TPN and TPS, monitored during August, 2011 (2 pages; see notes at bottom of table).

	CCME (2007)	INUG		PDL		TPN		TPS	
Parameter	Guideline ¹	13-Aug-11	13-Aug-11	11-Aug-11	11-Aug-11	23-Aug-11	23-Aug-11	23-Aug-11	23-Aug-11
Field Parameters (@ 3m)									
DO (mg/L)	NG	8.47	NA	8.80	NA	9.16	8.91	9.18	9.19
pH	NG	7.44	7.42	7.48	7.39	8.03	8.01	7.76	8.09
Conductivity (µS/cm)	NG	13.8	13.9	19.0	19.0	21.4	21.1	20.6	20.6
Temperature (°C)	NG	14.1	14.3	13.0	12.6	10.5	10.5	10.7	10.6
Physical Tests (mg/L)									
Conductivity (µS/cm)	NG	12.5	12.5	20.2	20.2	20.3	16.9	17.5	17.8
Hardness	NG	5.08	5.11	8.10	8.15	7.47	6.57	6.37	6.55
pH	6.5 - 9.0	6.94	7.05	7.23	7.23	7.00	7.08	7.13	7.12
Total Suspended Solids	NG	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0
Total Dissolved Solids	NG	16.0	18.0	12.0	16.0	10.0	6.0	<5.0	6.0
Turbidity (NTU)	NG	0.28	0.30	0.24	0.24	0.88	0.28	0.28	0.26
Anions & Nutrients (mg/L)									
Alkalinity - Bicarbonate	NG	7.1	7.0	10.0	10.1	7.5	7.6	7.7	7.7
Alkalinity - Carbonate	NG	<0.60	<0.60	<0.60	<0.60	<0.60	<0.60	<0.60	<0.60
Alkalinity - Hydroxide	NG	<0.40	<0.40	<0.40	<0.40	<0.40	<0.40	<0.40	<0.40
Alkalinity - Total	NG	5.8	5.8	8.2	8.3	6.2	6.2	6.3	6.3
Ammonia (as N) ^{2,3}	2.61 - 8.24	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010
Bromide	NG	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10
Chloride	NG	0.61	0.60	0.54	0.54	0.52	0.53	0.51	0.51
Fluoride	NG	0.053	0.053	0.034	0.034	0.059	0.059	0.059	0.059
Nitrate (as N)	2.9	<0.0050	<0.0050	<0.0050	<0.0050	0.0326	0.0300	0.0255	0.0251
Nitrite (as N)	0.060	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010
Total Kjeldahl Nitrogen	NG	0.157	0.116	0.118	0.123	0.163	0.100	0.134	0.112
Total Phosphate (as P)	0.004	0.0060	0.0087	0.0041	0.0068	0.0036	0.0027	0.0034	0.0051
Silicate (as SiO ₂)	NG	0.293	0.289	0.293	0.284	0.203	0.198	0.199	0.200
Sulphate (SO ₄)	NG	0.77	0.75	1.43	1.43	2.87	2.83	2.72	2.72
Carbon (mg/L)									
Dissolved Organic Carbon	NG	2.6	2.7	1.7	2.3	1.9	1.9	1.7	1.9
Total Organic Carbon	NG	2.6	2.7	2.1	2.3	1.9	2.0	2.0	2.0
Plant Pigments (µg/L)									
Chlorophyll-a	NG	-	-	-	-	0.76	0.81	0.68	0.80
Phaeophytin-a	NG	-	-	-	-	0.20	0.20	0.23	0.19
Total Metals (mg/L)									
Aluminum ²	0.100 - 0.100	0.0134	0.0097	0.0057	0.0064	0.0089	0.0093	0.0093	0.0089
Antimony	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Arsenic	0.0050	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Barium	NG	0.00203	0.00197	0.00201	0.00220	0.00383	0.00265	0.00266	0.00253
Beryllium	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Bismuth	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Boron	NG	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010
Cadmium ⁴	0.0000026 - 0.0000038	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010
Calcium	NG	1.06	1.03	2.07	2.13	1.89	1.70	1.66	1.66
Cesium	NG	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010
Chromium	0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010
Cobalt	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Copper ⁴	0.002 - 0.002	0.00051	0.00046	0.00059	0.00054	0.00050	0.00042	0.00039	0.00048
Iron	0.3	0.015	0.019	<0.010	<0.010	0.047	<0.010	0.015	<0.010
Lead ⁴	0.001 - 0.001	<0.000090	<0.000090	<0.000090	<0.000090	<0.000090	<0.000090	<0.000090	<0.000090
Lithium	NG	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020
Magnesium	NG	0.638	0.617	0.727	0.730	0.703	0.707	0.680	0.671
Manganese	NG	0.00266	0.00258	0.00133	0.00138	0.00606	0.00498	0.00447	0.00459
Mercury	0.000026	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010
Molybdenum	0.073	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Nickel ⁴	0.025 - 0.025	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020
Phosphorus	NG	<0.20	<0.20	<0.20	<0.20	<0.20	<0.20	<0.20	<0.20

Parameter	CCME (2007)	INUG		PDL		TPN		TPS	
	Guideline ¹	13-Aug-11	13-Aug-11	11-Aug-11	11-Aug-11	23-Aug-11	23-Aug-11	23-Aug-11	23-Aug-11
Potassium	NG	0.360	0.349	0.327	0.329	0.448	0.433	0.428	0.424
Rubidium	NG	0.00049	0.00047	0.00039	0.00040	0.00063	0.00059	0.00061	0.00058
Selenium	0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010
Silicon	NG	0.167	0.166	0.151	0.159	0.140	0.115	0.111	0.102
Silver	0.0001	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010
Sodium	NG	0.508	0.498	0.465	0.471	0.843	0.857	0.815	0.802
Strontium	NG	0.00597	0.00579	0.00835	0.00859	0.00813	0.00783	0.00769	0.00733
Tellurium	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Thallium	0.0008	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010
Thorium	NG	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010
Tin	NG	<0.00020	<0.00020	<0.00020	<0.00020	0.00035	<0.00020	<0.00020	<0.00020
Titanium	NG	0.00027	0.00022	<0.00020	<0.00020	<0.00020	<0.00020	0.00023	0.00024
Tungsten	NG	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010
Uranium	NG	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010
Vanadium	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Zinc	0.030	<0.0050	<0.0050	<0.0050	<0.0050	<0.0050	<0.0050	<0.0050	<0.0050
Zirconium	NG	<0.00040	<0.00040	<0.00040	<0.00040	<0.00040	<0.00040	<0.00040	<0.00040
Dissolved Metals (mg/L) ⁵									
Aluminum ²	0.100 - 0.100	0.0033	0.0034	0.0026	0.0023	0.0098	0.0039	0.0033	0.0031
Antimony	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Arsenic	0.0050	<0.00020	<0.00020	<0.00020	<0.00020	0.00032	<0.00020	<0.00020	<0.00020
Barium	NG	0.00185	0.00185	0.00196	0.00197	0.00226	0.00284	0.00267	0.00270
Beryllium	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Bismuth	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Boron	NG	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010
Cadmium ⁴	0.0000026 - 0.0000038	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010
Calcium	NG	1.00	0.986	2.01	2.05	1.65	1.58	1.56	1.59
Cesium	NG	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010
Chromium	0.0010	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020
Cobalt	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Copper ⁴	0.0020 - 0.0020	0.00040	0.00036	0.00048	0.00042	0.00064	0.00036	0.00034	0.00035
Iron	0.300	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010	<0.010
Lead ⁴	0.0010 - 0.0010	<0.000090	<0.000090	<0.000090	0.000173	<0.000090	<0.000090	<0.000090	<0.000090
Lithium	NG	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020	<0.0020
Magnesium	NG	0.626	0.644	0.747	0.735	0.812	0.639	0.605	0.625
Manganese	NG	0.00042	0.00033	0.00055	0.00060	0.00041	0.00449	0.00356	0.00297
Mercury	NG	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010	<0.000010
Molybdenum	0.073	<0.00010	<0.00010	<0.00010	<0.00010	0.00016	0.00018	0.00016	0.00016
Nickel ⁴	0.025 - 0.025	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010
Phosphorus	NG	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10
Potassium	NG	0.337	0.331	0.318	0.317	0.415	0.404	0.396	0.393
Rubidium	NG	0.00045	0.00044	0.00039	0.00038	0.00065	0.00054	0.00055	0.00056
Selenium	0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010	<0.0010
Silicon	NG	0.139	0.139	0.137	0.132	0.097	0.096	0.100	0.097
Silver	0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010
Sodium	NG	0.495	0.497	0.473	0.456	0.510	0.783	0.727	0.733
Strontium	NG	0.00527	0.00534	0.00781	0.00800	0.00795	0.00717	0.00727	0.00726
Tellurium	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Thallium	0.00080	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010
Thorium	NG	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010
Tin	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Titanium	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Tungsten	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Uranium	NG	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010	<0.00010
Vanadium	NG	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020	<0.00020
Zinc	0.030	<0.0020	<0.0020	<0.0020	0.0021	<0.0020	<0.0020	<0.0020	<0.0020
Zirconium	NG	<0.00040	<0.00040	<0.00040	<0.00040	<0.00040	<0.00040	<0.00040	<0.00040

Notes: NG = no guideline; ¹ CCME (Canadian Council of Ministers of the Environment) Canadian Water Quality Guidelines for the Protection of Aquatic Life, 1999, updated December 2007; ² Guideline is pH dependent; ³ Guideline is temperature dependent; ⁴ Guideline is hardness dependent; ⁵ Guidelines have not yet been made for "Dissolved Metals" thus were screened against CCME guidelines for "Total Metals"; **Shaded** values exceed the CCME guideline.

Table 3.3- 5. CREMP sediment quality results at INUG, PDL, and TPN during August, 2011 (2 pages; see notes at bottom of table).

Area ID	Sediment Quality		INUG					PDL					TPN				
Replicate ID	Guidelines (CCME 2002) ¹		1	2	3	4	5	1	2	3	4	5	1	2	3	4	5
Date	ISQG	PEL	14-Aug-11	14-Aug-11	14-Aug-11	14-Aug-11	14-Aug-11	13-Aug-11	13-Aug-11	13-Aug-11	13-Aug-11	13-Aug-11	19-Aug-11	19-Aug-11	19-Aug-11	19-Aug-11	19-Aug-11
Physical & Organic Parameters																	
Moisture (%)	NG	NG	79.0	79.7	84.6	83.1	82.4	78.8	79.3	72.1	79.4	80.8	76.1	85.8	71.9	57.8	39.2
pH	NG	NG	5.96	6.08	5.69	5.73	6.01	7.87	5.87	5.97	5.87	6.12	6.06	6.03	5.63	5.95	5.25
Total Organic Carbon (% dw)	NG	NG	2.93	2.99	4.84	4.01	4.22	2.44	2.64	1.77	2.76	3.09	1.80	3.71	1.25	0.66	0.51
Particle Size																	
% Gravel (>2mm)	NG	NG	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10	<0.10	0.64	<0.10	<0.10	3.62	4.49
% Sand (2.00mm - 0.063mm)	NG	NG	11.5	13.6	5.08	2.69	2.71	13.7	14.9	15.8	5.71	12.6	46.0	14.5	46.8	38.7	25.3
% Silt (0.063mm - 4µm)	NG	NG	75.1	78.9	90.1	87.3	88.1	74.0	75.0	76.0	77.3	81.4	42.8	70.8	46.5	46.8	42.2
% Clay (<4µm)	NG	NG	13.4	7.49	4.86	9.98	9.20	12.3	10.1	8.18	16.9	6.05	10.6	14.7	6.74	10.9	28.1
Aggregate Organics (mg/kg)																	
Mineral Oil and Grease	NG	NG	<900	<950	<1300	<1125	<1050	<950	<950	<675	<900	<1025	<800	<1400	<750	<500	<500
Hydrocarbons (mg/kg)																	
EPH10-19	NG	NG	<370	<400	<510	<520	<480	<350	<430	<290	<400	<400	<340	<640	<300	<200	<200
EPH19-32	NG	NG	<370	<400	<510	<520	<480	<350	<430	<290	<400	<400	<340	<640	<300	<200	<200
LEPH	NG	NG	<370	<400	<510	<520	<480	<350	<430	<290	<400	<400	<340	<640	<300	<200	<200
HEPH	NG	NG	<370	<400	<510	<520	<480	<350	<430	<290	<400	<400	<340	<640	<300	<200	<200
Polycyclic Aromatic Hydrocarbons (mg/kg)																	
Acenaphthene	0.00671	0.0889	<0.0050	<0.0050	<0.0065	<0.0065	<0.0065	<0.0050	<0.0055	<0.0050	<0.0050	<0.010	<0.0050	<0.0080	<0.0050	<0.0050	<0.0050
Acenaphthylene	0.00587	0.128	<0.0050	<0.0050	<0.0065	<0.0065	<0.0065	<0.0050	<0.0055	<0.0050	<0.0050	<0.0050	<0.0050	<0.0080	<0.0050	<0.0050	<0.0050
Anthracene	0.0469	0.245	<0.0040	<0.0040	<0.0052	<0.0052	<0.0052	<0.0040	<0.0044	<0.0040	<0.0040	<0.0040	<0.0040	<0.0064	<0.0040	<0.0040	<0.0040
Benz(a)anthracene	0.0317	0.385	<0.010	<0.010	<0.013	<0.013	<0.013	<0.010	<0.011	<0.010	<0.010	<0.010	<0.010	<0.016	<0.010	<0.010	<0.010
Benzo(a)pyrene	0.0319	0.782	<0.010	<0.010	<0.013	<0.013	<0.013	<0.010	<0.011	<0.010	<0.010	<0.010	<0.010	<0.016	<0.010	<0.010	<0.010
Benzo(b)fluoranthene	NG	NG	<0.010	<0.010	<0.013	<0.013	<0.013	<0.010	<0.011	<0.010	<0.020	<0.020	<0.010	<0.016	<0.020	<0.020	<0.010
Benzo(b+j+k)fluoranthene	NG	NG	<0.015	<0.015	<0.018	<0.018	<0.018	<0.015	<0.016	<0.015	<0.022	<0.022	<0.015	<0.023	<0.022	<0.022	<0.015
Benzo(g,h,i)perylene	NG	NG	<0.010	<0.010	<0.013	<0.013	<0.013	<0.010	<0.011	<0.010	<0.010	<0.010	<0.010	<0.016	<0.010	<0.010	<0.010
Benzo(k)fluoranthene	NG	NG	<0.010	<0.010	<0.013	<0.013	<0.013	<0.010	<0.011	<0.010	<0.010	<0.010	<0.010	<0.016	<0.010	<0.010	<0.010
Chrysene	0.0571	0.862	<0.010	<0.010	<0.013	<0.013	<0.013	<0.010	<0.011	<0.010	<0.010	<0.010	<0.010	<0.016	<0.010	<0.010	<0.010
Dibenz(a,h)anthracene	0.00622	0.135	<0.0050	<0.0050	<0.0065	<0.0065	<0.0065	<0.0050	<0.0055	<0.0050	<0.0050	<0.0050	<0.0050	<0.0080	<0.0050	<0.0050	<0.0050
Fluoranthene	0.111	2.355	<0.010	<0.010	<0.013	<0.013	<0.013	<0.010	<0.011	<0.010	<0.010	<0.010	<0.010	<0.016	<0.010	<0.010	<0.010
Fluorene	0.0212	0.144	<0.010	<0.010	<0.013	<0.013	<0.013	<0.010	<0.011	<0.010	<0.010	<0.010	<0.010	<0.016	<0.010	<0.010	<0.010
Indeno(1,2,3-c,d)pyrene	NG	NG	<0.010	<0.010	<0.013	<0.013	<0.013	<0.010	<0.011	<0.010	<0.010	<0.010	<0.010	<0.016	<0.010	<0.010	<0.010
2-Methylnaphthalene	0.0202	0.201	<0.010	<0.010	<0.013	<0.013	<0.013	<0.010	<0.011	<0.010	<0.010	<0.010	<0.010	<0.016	<0.010	<0.010	<0.010
Naphthalene	0.0346	0.391	<0.020	<0.020	<0.020	<0.020	<0.020	<0.020	<0.020	<0.010	<0.020	<0.020	<0.020	<0.030	<0.020	<0.010	<0.010
Phenanthrene	0.0419	0.515	<0.010	<0.010	<0.013	<0.013	<0.013	<0.010	<0.011	<0.010	<0.010	<0.010	<0.010	<0.016	0.011	<0.010	<0.010
Pyrene	0.053	0.875	<0.010	<0.010	<0.013	<0.013	<0.013	<0.010	<0.011	<0.010	<0.010	<0.010	<0.010	<0.016	<0.010	<0.010	<0.010
B(a)P Total Potency	NG	NG	<0.020	<0.020	<0.020	<0.020	<0.020	<0.020	<0.020	<0.020	<0.020	<0.020	<0.020	<0.020	<0.020	<0.020	<0.020
IACR (CCME)	NG	NG	<0.15	<0.15	<0.15	<0.15	<0.15	<0.15	<0.15	<0.15	<0.15	<0.15	<0.15	<0.17	<0.15	<0.15	<0.15

Area ID Replicate ID Date	Sediment Quality		INUG					PDL					TPN				
	Guidelines (CCME 2002) ¹		1	2	3	4	5	1	2	3	4	5	1	2	3	4	5
	ISQG	PEL	14-Aug-11	14-Aug-11	14-Aug-11	14-Aug-11	14-Aug-11	13-Aug-11	13-Aug-11	13-Aug-11	13-Aug-11	13-Aug-11	19-Aug-11	19-Aug-11	19-Aug-11	19-Aug-11	19-Aug-11
Total Metals (mg/kg dw)																	
Aluminum	NG	NG	21900	21000	21600	19800	15500	19000	15100	17300	23900	17200	13300	17600	11900	14700	21100
Antimony	NG	NG	0.16	0.14	0.21	0.18	0.16	0.24	0.18	0.23	0.30	0.20	0.15	0.20	0.10	0.10	0.11
Arsenic	5.9	17	23.4	6.63	34.1	155	78.3	20.2	7.45	91.9	41.4	8.45	8.89	13.7	5.93	15.6	10.7
Barium	NG	NG	103	107	133	116	90.8	81.9	77.1	72.3	104	90.2	48.6	73.4	41.1	56.2	102
Beryllium	NG	NG	1.21	1.19	1.35	1.20	0.89	0.94	0.70	0.77	1.12	0.76	0.70	1.07	0.64	0.83	0.90
Bismuth	NG	NG	1.14	1.05	1.58	1.07	0.86	0.86	0.60	0.72	1.10	0.70	0.77	1.13	0.66	0.85	1.05
Cadmium	0.6	3.5	0.11	0.10	0.25	0.18	0.15	0.12	0.19	0.12	0.14	0.22	0.07	0.24	0.06	<0.050	0.10
Calcium	NG	NG	1630	1800	1790	1600	1330	1790	1660	1690	2140	1850	1220	1590	867	829	1610
Chromium	37.3	90	110	103	108	96.9	76.0	124	97.2	114	157	111	89.5	115	73.5	88.8	97.4
Cobalt	NG	NG	15.3	9.26	14.0	13.0	14.1	11.6	8.25	17.5	20.4	9.61	7.56	10.5	6.88	9.72	13.6
Copper	35.7	197	43.8	39.5	48.2	42.2	33.1	46.3	33.5	35.5	57.9	38.8	33.4	54.1	27.9	34.3	26.7
Iron	NG	NG	57700	27800	64700	121000	94900	33400	19700	58900	46200	22600	20800	27200	16700	24400	31100
Lead	35	91.3	12.5	12.1	14.9	12.2	10.5	13.7	10.4	11.8	16.6	12.0	11.3	15.2	8.67	10.2	13.8
Lithium	NG	NG	34.2	33.6	33.4	28.8	17.6	32.0	27.0	28.6	37.9	30.2	19.9	32.1	19.2	31.7	55.9
Magnesium	NG	NG	9870	9620	9520	8560	6900	10400	8150	9830	12400	9320	6830	8430	6070	7480	10500
Manganese	NG	NG	1530	406	1340	1540	2680	359	216	873	1410	255	279	407	190	456	700
Mercury	0.170	0.486	0.016	0.016	0.028	0.023	0.022	0.010	0.008	0.011	0.012	0.010	0.011	0.016	0.007	0.006	<0.0050
Molybdenum	NG	NG	4.20	2.02	4.59	7.61	10.2	2.80	1.26	4.64	3.46	1.42	1.56	2.75	1.72	3.63	2.68
Nickel	NG	NG	72.0	65.9	85.9	67.6	61.4	74.7	64.1	71.4	99.8	71.6	44.6	64.4	39.9	45.6	63.1
Phosphorus	NG	NG	560	637	1230	3190	841	531	318	964	547	387	374	569	213	228	305
Potassium	NG	NG	3520	3330	3450	3170	2290	2360	1890	2140	3220	2110	1650	2290	1550	1960	3720
Selenium	NG	NG	0.46	0.39	0.68	0.66	0.57	0.41	0.29	0.39	0.34	0.31	0.29	0.70	0.23	<0.20	<0.20
Silver	NG	NG	<0.10	0.12	0.19	0.17	0.12	0.15	0.17	0.13	<0.10	0.19	<0.10	0.18	<0.10	<0.10	<0.10
Sodium	NG	NG	120	120	140	120	<100	110	<100	<100	100	<100	<100	120	<100	<100	130
Strontium	NG	NG	23.0	23.9	24.4	22.5	17.9	20.2	17.4	19.3	24.0	18.0	10.8	13.9	8.65	8.38	15.7
Thallium	NG	NG	0.203	0.185	0.217	0.188	0.160	0.169	0.133	0.159	0.212	0.154	0.144	0.217	0.130	0.174	0.285
Tin	NG	NG	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0
Titanium	NG	NG	556	541	525	472	384	486	394	485	501	402	384	454	380	438	968
Uranium	NG	NG	16.1	13.9	16.8	14.1	10.8	8.13	5.82	6.31	10.7	6.70	9.16	15.2	8.09	10.0	12.2
Vanadium	NG	NG	35.1	34.8	34.9	31.5	24.8	33.7	25.3	31.1	41.0	28.9	24.8	31.9	21.0	25.4	38.4
Zinc	123	315	80.0	76.2	89.6	76.3	61.9	76.4	60.5	67.0	87.3	67.8	56.0	84.3	54.0	66.4	86.9

Notes: NG = no guideline; ¹ CCME (Canadian Council of Ministers of the Environment) Canadian Sediment Quality Guidelines for the Protection of Aquatic Life, 1999, updated in 2002; ISQG = Interim freshwater Sediment Quality Guideline; PEL = Probable Effect Level; **Shaded** concentrations = or > ISQG; **Boxed** concentrations also > PEL.

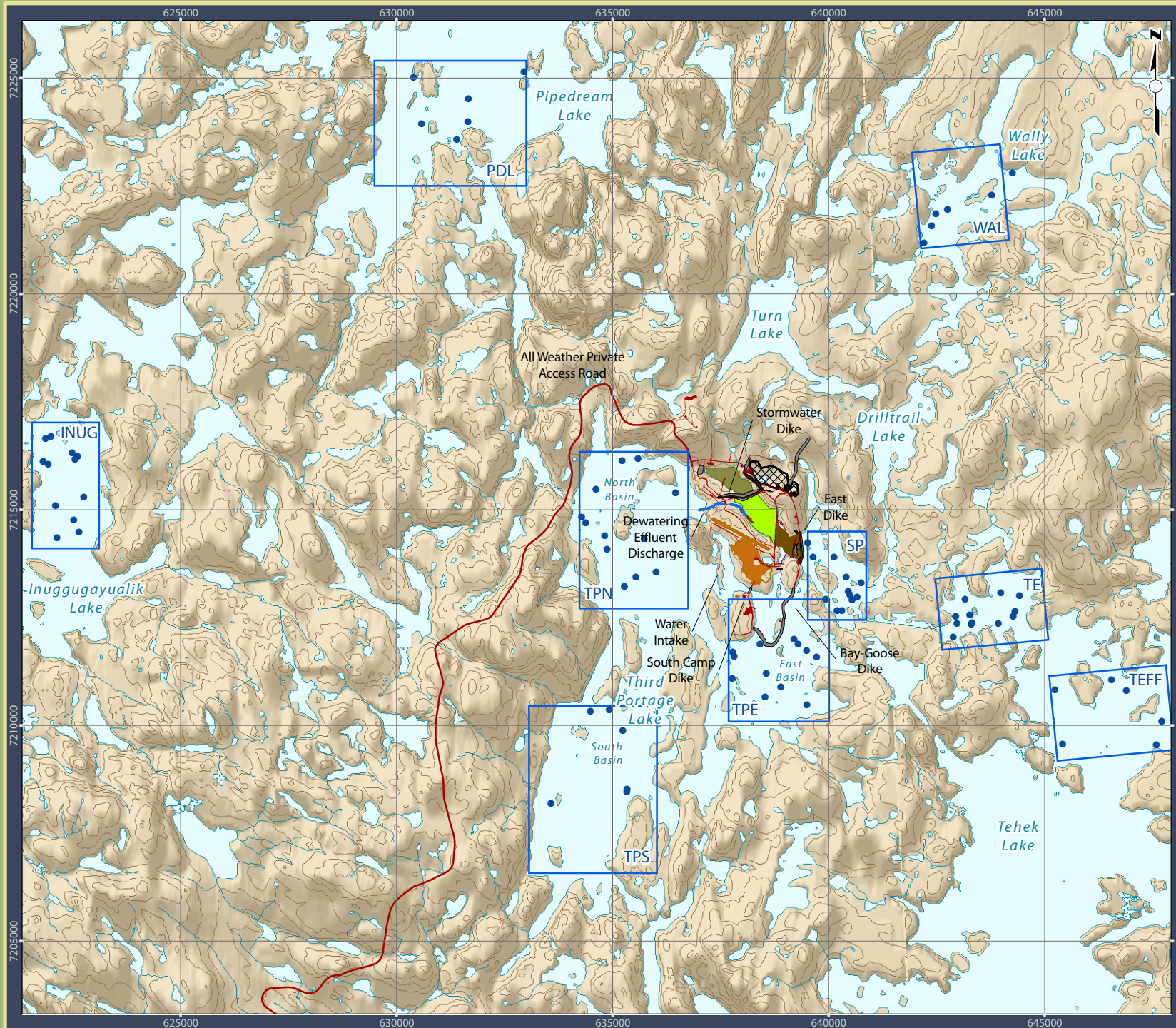
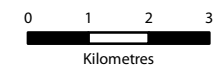
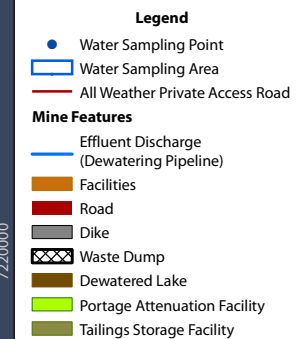


Figure 3.3-1. CREMP Water Quality Monitoring and Sampling Areas 2011



Projection: UTM Zone 14 NAD83

Data Sources:
 Natural Resources Canada, GeoBase®
 National Topographic Database
 Agnico-Eagle Mines Limited.
 Azimuth Consulting Group Inc.

Meadowbank Gold Project

Prepared for:



By: **CASLYS CONSULTING**

Figure 3.3- 2. Total Suspended Solids (TSS; mg/L) in water samples from CREMP monitoring at Meadowbank study lakes since 2006 (relevant sampling areas for EEM are TPN, INUG, PDL, and TPS).

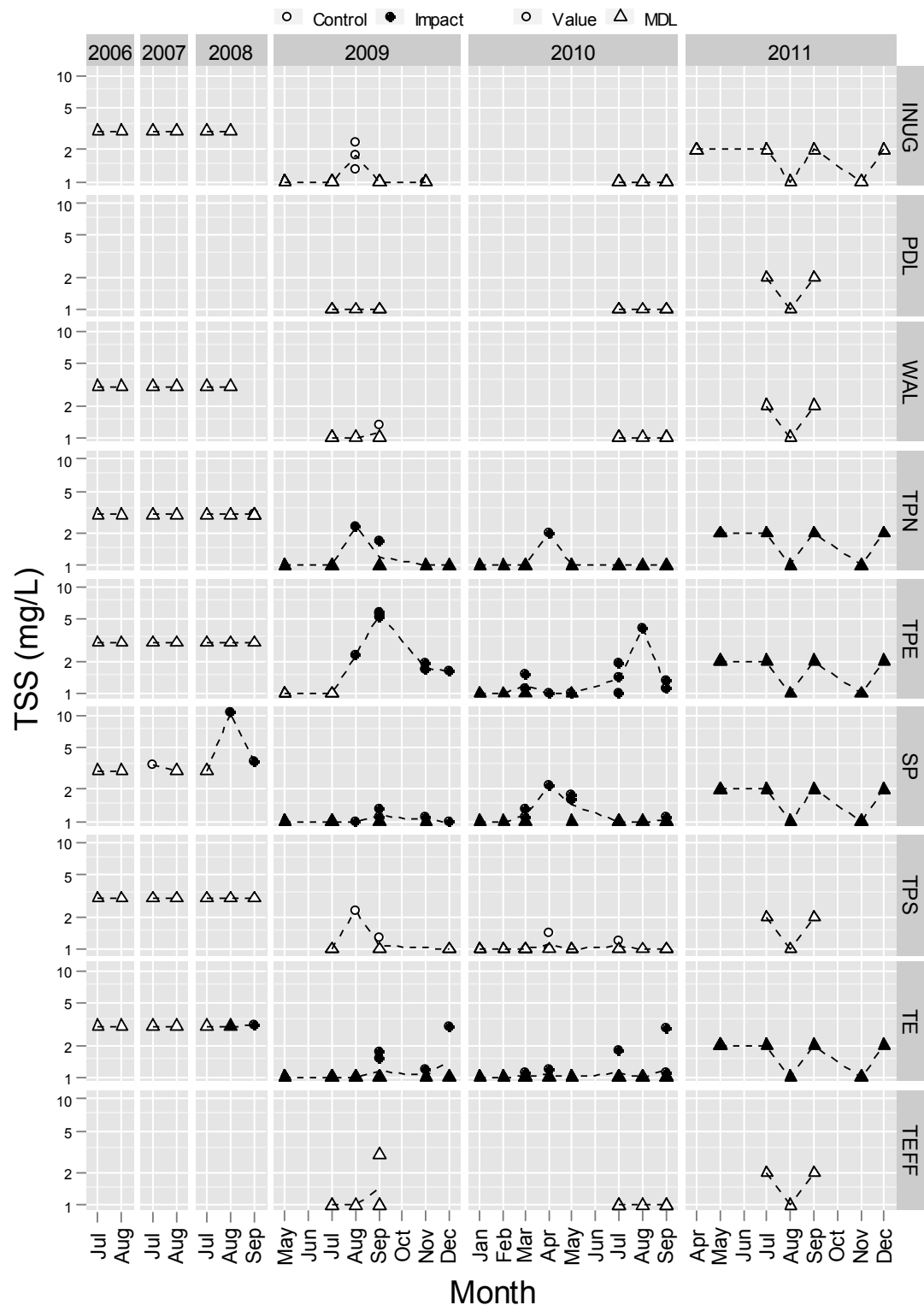


Figure 3.3- 3. Total Aluminum (mg/L) in water samples from CREMP monitoring at Meadowbank study lakes since 2006 (relevant sampling areas for EEM are TPN, INUG, PDL, and TPS).

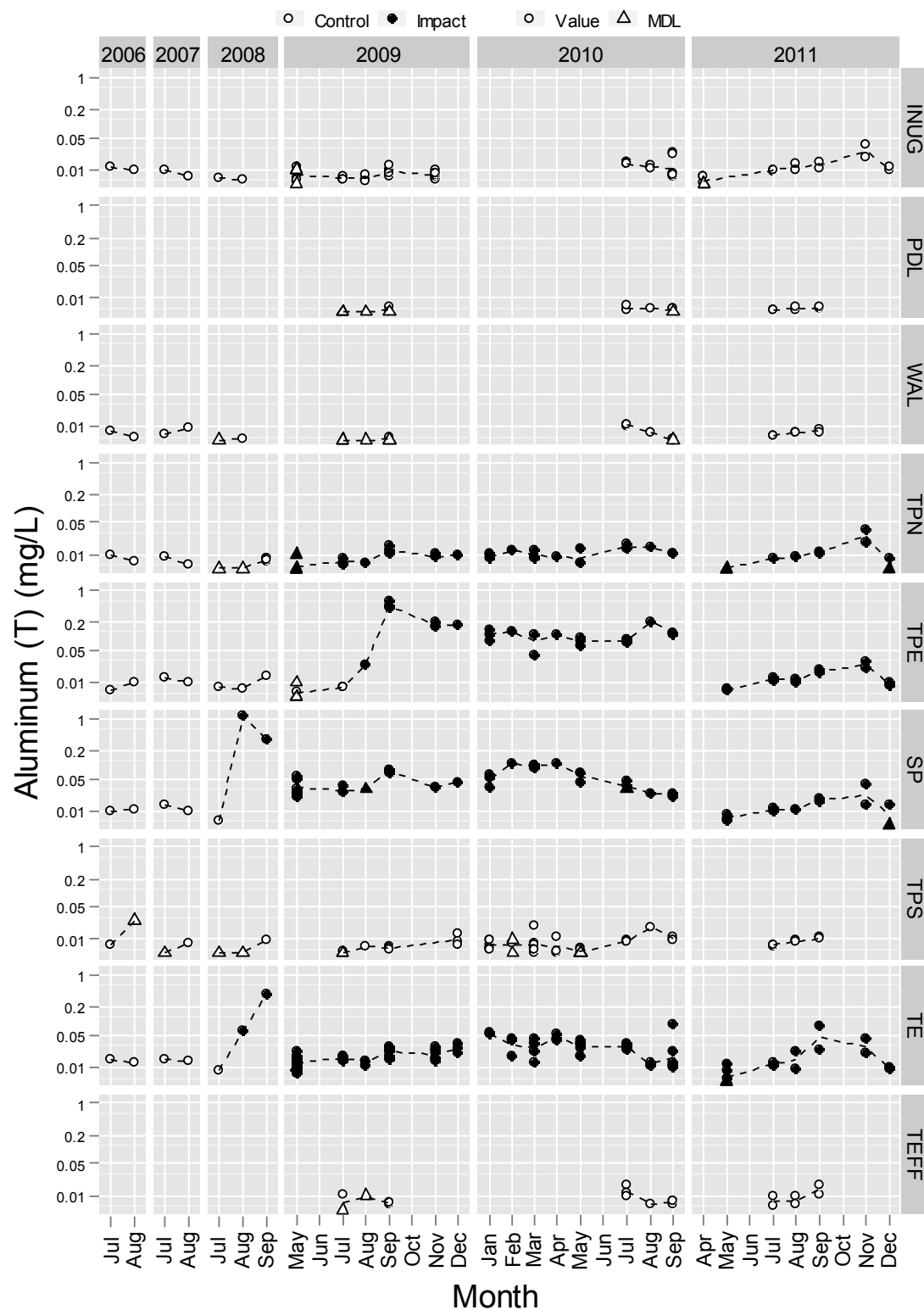


Figure 3.3- 4. pH in water samples from CREMP monitoring at Meadowbank study lakes since 2006 (relevant sampling areas for EEM are TPN, INUG, PDL, and TPS).

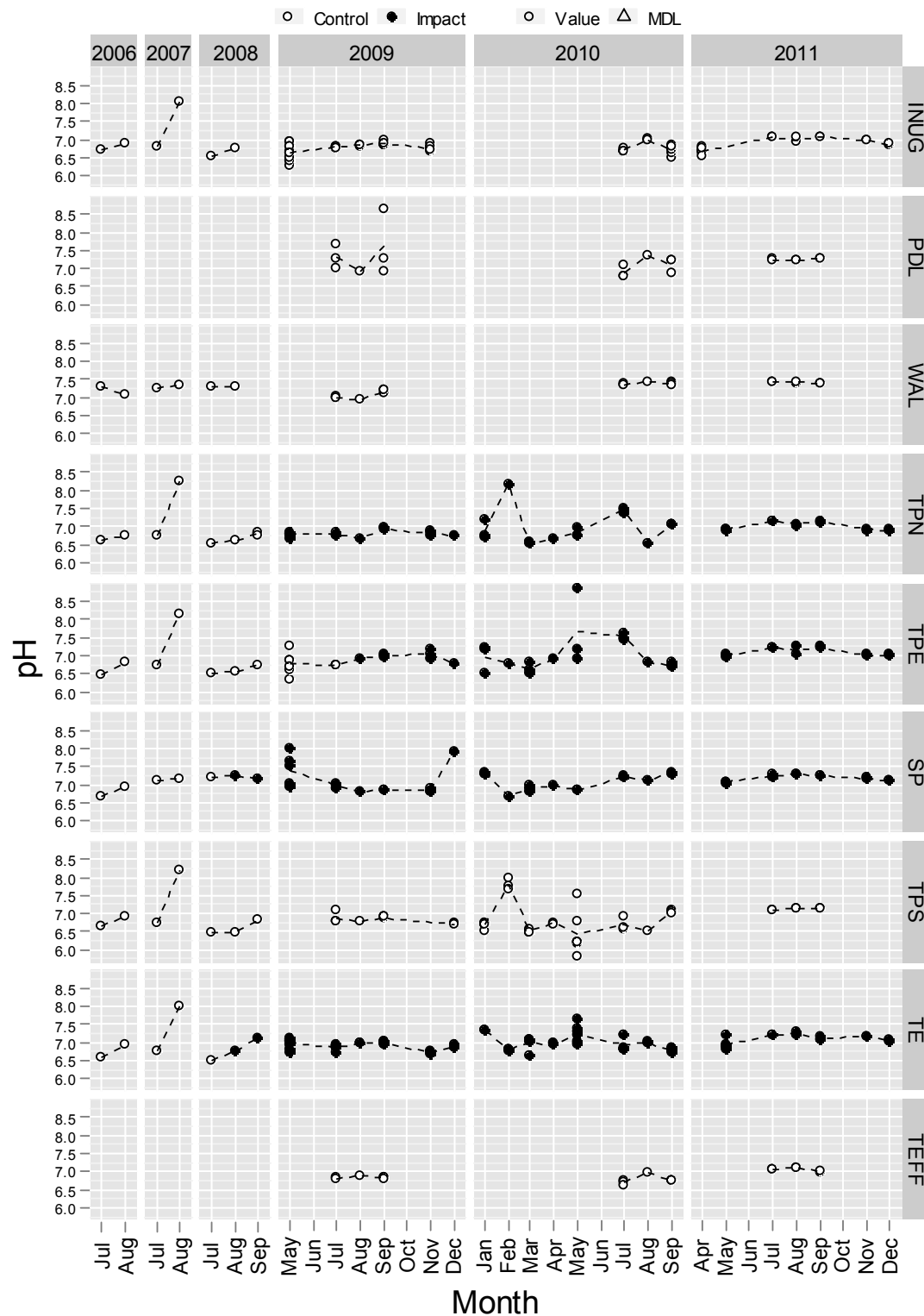


Figure 3.3- 5. Total Copper (mg/L) in water samples from CREMP monitoring at Meadowbank study lakes since 2006 (relevant sampling areas for EEM are TPN, INUG, PDL, and TPS).

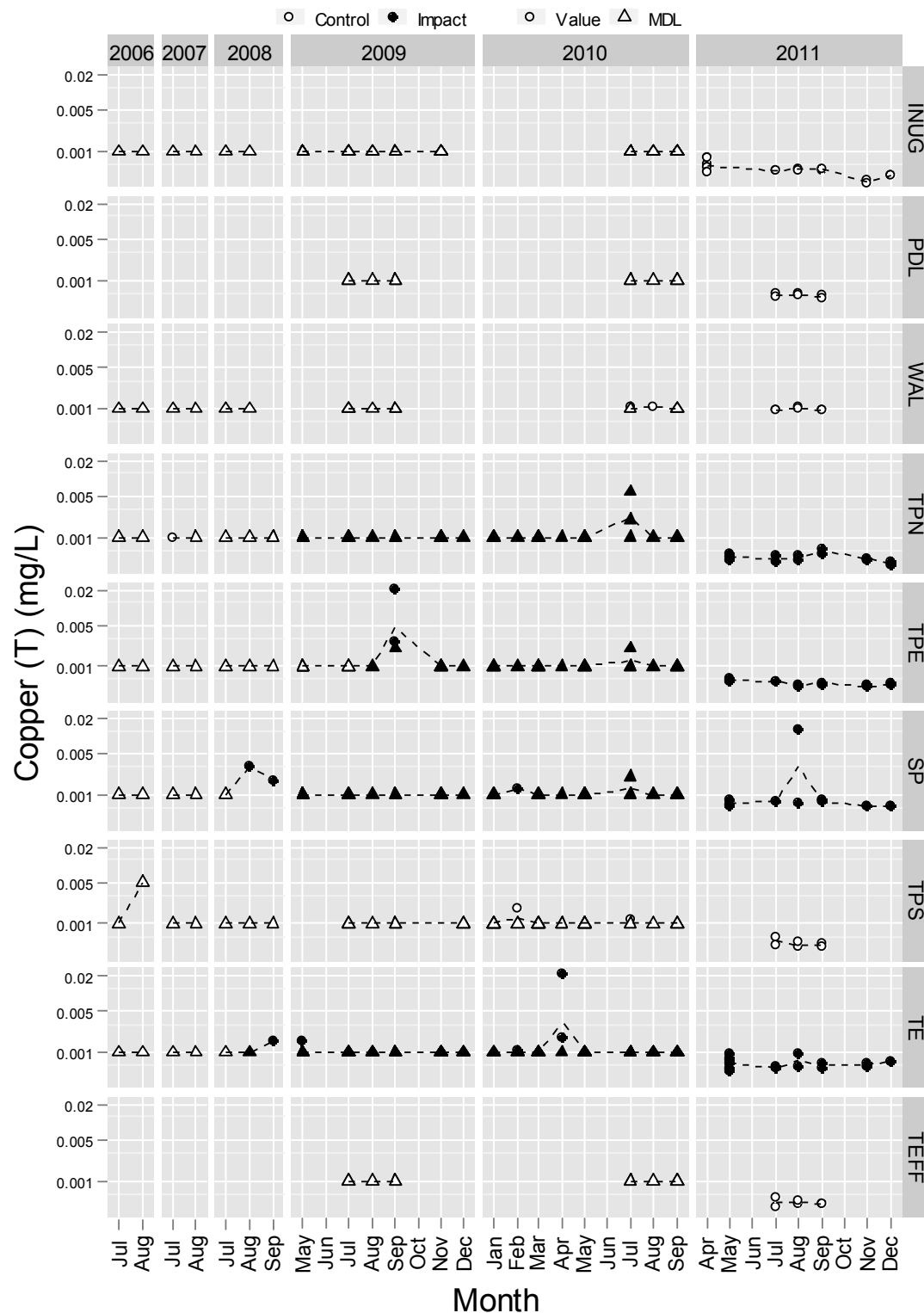
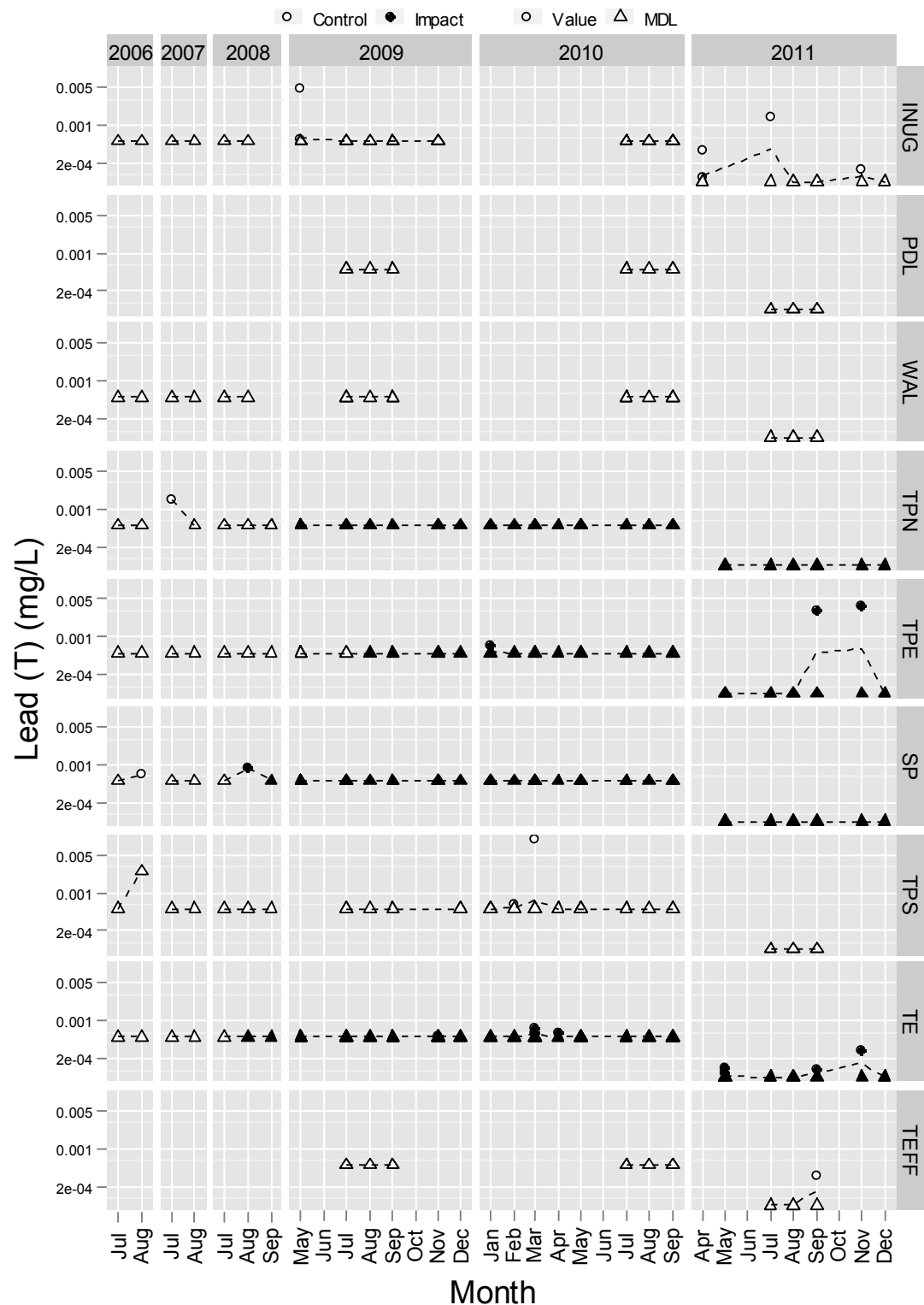
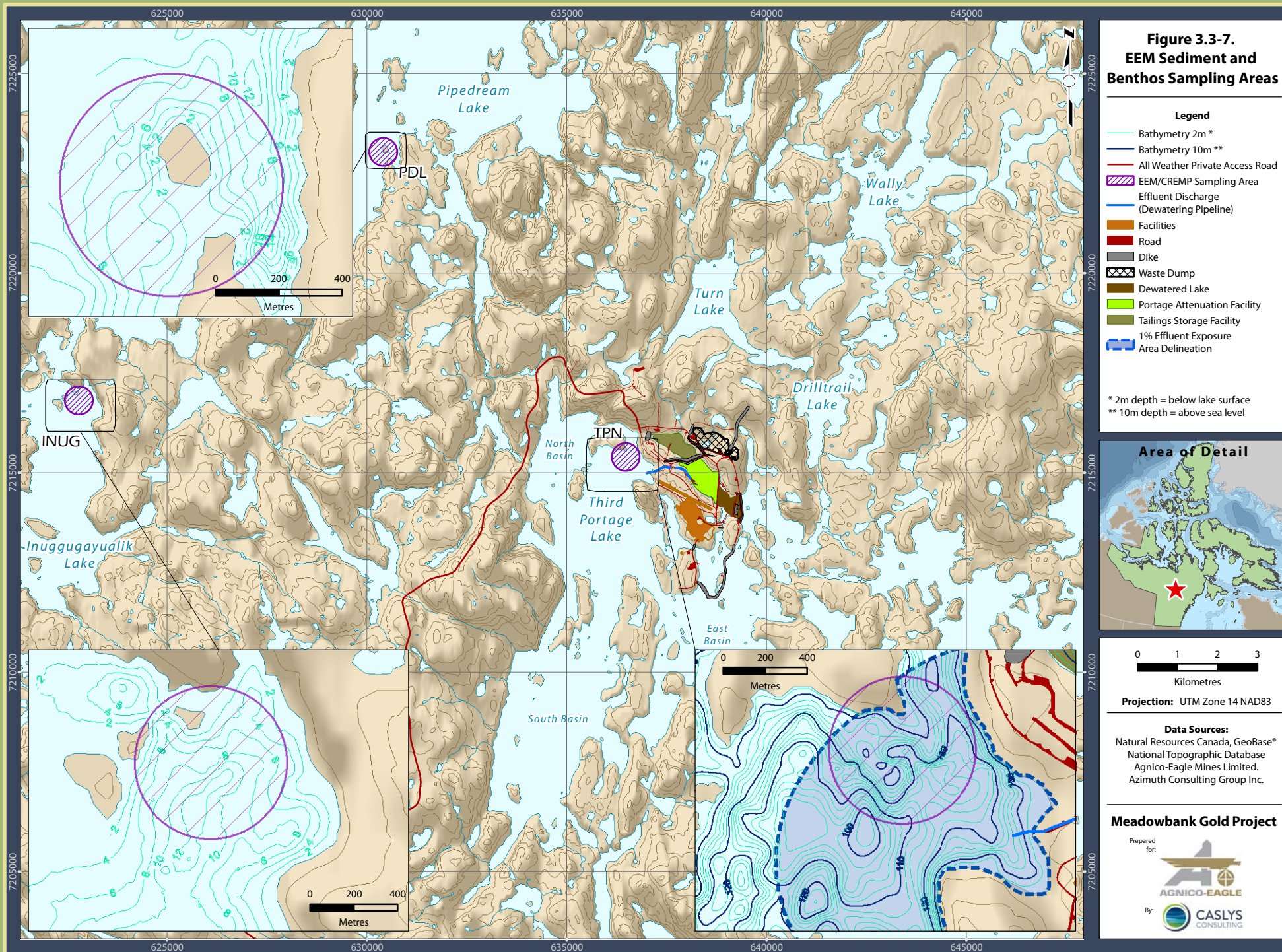


Figure 3.3- 6. Total Lead (mg/L) in water samples from CREMP monitoring at Meadowbank study lakes since 2006 (relevant sampling areas for EEM are TPN, INUG, PDL, and TPS).



**Figure 3.3-7.
EEM Sediment and
Benthos Sampling Areas**



4. FISH SURVEY

4.1. Sample Collection

Fish were collected from each of the three sampling areas (INUG, PDL, and TPN) from August 11–18th, 2011. Sampling locations (UTM coordinates) and dates are presented in **Table 4.1-1** and gillnet sets locations are illustrated in **Figure 4.1-1**.

All fish were collected and analyzed according to the guidelines of the fish sampling SOP (**Appendix F**) and according to the QA/QC procedures presented in the Study Design document (Azimuth, 2011). Field and field laboratory data sheets from August 2011 are presented in **Appendix G**. Fish catch summary statistics are discussed and presented in **Section 4.3**.

Table 4.1- 1. Fish sampling areas: gillnet locations (UTM coordinates, NAD83) (2 pages; see notes at bottom of table).

Area ¹	Net ID	Sampling Date	Point A		Point B	
			Latitude (dd.mm.ss.s)	Longitude (dd.mm.ss.s)	Latitude (dd.mm.ss.s)	Longitude (dd.mm.ss.s)
INUG	1	16-Aug-11	65 03 12.4	96 23 32.8	65 03 10.9	96 23 35.5
	2	16-Aug-11	65 02 50.7	96 23 06.6	65 02 52.5	96 23 17.0
	3	17-Aug-11	65 03 15.5	96 23 04.2	65 03 16.5	96 23 14.7
	4	17-Aug-11	65 03 03.8	96 22 47.2	65 03 03.3	96 22 57.5
	5	17-Aug-11	65 02 53.1	96 22 55.2	65 02 54.0	96 22 58.8
	6	17-Aug-11	65 02 57.0	96 22 42.5	65 02 58.0	96 22 51.5
	7	17-Aug-11	65 02 30.6	96 23 02.4	65 02 33.2	96 23 02.4
	8	17-Aug-11	65 02 36.9	96 22 57.8	65 02 37.1	96 23 01.1
	9	18-Aug-11	65 03 14.9	96 23 51.0	65 03 18.0	96 24 00.8
	10	18-Aug-11	65 03 16.6	96 23 39.3	65 03 17.7	96 23 42.1
	11	18-Aug-11	65 02 34.7	96 22 54.2	65 02 34.7	96 23 05.8
	12	18-Aug-11	65 02 51.0	96 23 06.9	65 02 51.2	96 23 10.8
	13	18-Aug-11	65 03 15.5	96 23 57.1	65 03 19.7	96 23 58.2
	14	18-Aug-11	65 02 45.8	96 23 03.7	65 02 45.6	96 23 08.5
PDL	1	16-Aug-11	65 06 14.4	96 13 18.2	65 06 17.7	96 13 27.2
	2	16-Aug-11	65 06 22.3	96 13 29.6	65 06 23.6	96 13 34.2
	3	17-Aug-11	65 06 30.9	96 12 53.9	65 06 39.2	96 12 57.4
	4	17-Aug-11	65 06 27.0	96 13 07.0	65 06 28.9	96 13 17.7
	5	18-Aug-11	65 06 19.1	96 13 21.5	65 06 13.9	96 13 19.8
	6	18-Aug-11	65 06 13.9	96 13 19.8	65 06 13.1	96 13 31.1
	7	18-Aug-11	65 06 12.1	96 13 23.7	65 06 13.6	96 13 26.5
	8	18-Aug-11	65 06 07.1	96 13 23.5	65 06 08.1	96 13 34.9

Area ¹	Net ID	Sampling Date	Point A		Point B	
			Latitude (dd.mm.ss.s)	Longitude (dd.mm.ss.s)	Latitude (dd.mm.ss.s)	Longitude (dd.mm.ss.s)
TPN	1	11-Aug-11	65 02 06.0	96 06 07.6	65 02 01.7	96 06 09.5
	2	11-Aug-11	65 02 07.7	96 06 00.6	65 02 05.4	96 05 52.7
	3	11-Aug-11	65 01 45.0	96 05 51.5	65 01 50.2	96 05 51.5
	4	11-Aug-11	65 02 00.4	96 05 31.4	65 01 58.4	96 05 40.9
	5	12-Aug-11	65 01 44.1	96 05 32.2	65 01 47.7	96 05 27.2
	6	12-Aug-11	65 02 06.1	96 06 04.0	65 02 02.3	96 05 58.3
	7	13-Aug-11	65 02 04.5	96 07 09.5	65 01 59.9	96 07 10.7
	8	13-Aug-11	65 01 42.2	96 05 55.3	65 01 44.4	96 06 07.3
	9	13-Aug-11	65 01 50.5	96 05 21.0	65 01 51.3	96 05 31.9
	10	13-Aug-11	65 01 44.9	96 05 44.6	65 01 50.3	96 05 42.9
	11	13-Aug-11	65 01 56.6	96 05 25.7	65 01 58.2	96 05 24.0
	12	14-Aug-11	65 01 46.5	96 05 20.6	65 01 47.9	96 05 30.9
	13	14-Aug-11	65 01 42.9	96 05 18.7	65 01 45.0	96 05 28.5
	14	14-Aug-11	65 01 47.5	96 05 21.2	65 01 46.0	96 05 20.2
	15	14-Aug-11	65 01 45.0	96 05 19.0	65 01 43.1	96 05 17.6
	16	14-Aug-11	65 01 45.3	96 05 46.5	65 01 50.8	96 05 47.5
	17	14-Aug-11	65 02 00.2	96 05 29.8	65 01 56.1	96 05 37.7
	18	14-Aug-11	65 02 03.2	96 05 35.2	65 02 01.7	96 05 37.8
	19	14-Aug-11	65 01 42.5	96 05 55.1	65 01 46.4	96 06 02.7
	20	15-Aug-11	65 02 10.8	96 05 42.2	65 02 09.9	96 05 45.8
	21	15-Aug-11	65 02 06.7	96 06 02.8	65 02 04.9	96 06 01.8
	22	15-Aug-11	65 01 45.9	96 06 06.7	65 01 49.9	96 06 11.5
	23	15-Aug-11	65 02 04.8	96 05 37.4	65 02 02.5	96 05 47.8
	24	15-Aug-11	65 01 49.4	96 05 25.0	65 01 47.7	96 05 25.0
	25	15-Aug-11	65 01 52.5	96 05 19.5	65 01 52.5	96 05 24.1

Notes: ¹ Areas are as follows: TPN=Third Portage Lake - North; INUG=Inuggugayualik Lake; PDL=Pipedream Lake.

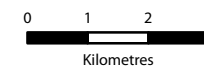
**Figure 4.1-1.
EEM Fish Sampling
Areas**

Legend

- Bathymetry 2m *
- Bathymetry 10m **
- All Weather Private Access Road
- EEM Gillnet location
- Effluent Discharge
(Dewatering Pipeline)
- Facilities
- Road
- Dike
- Waste Dump
- Dewatered Lake
- Portage Attenuation Facility
- Tailings Storage Facility
- 1% Effluent Exposure
Area Delineation

* 2m depth = below lake surface
** 10m depth = above sea level

Area of Detail



Projection: UTM Zone 14 NAD83

Data Sources:

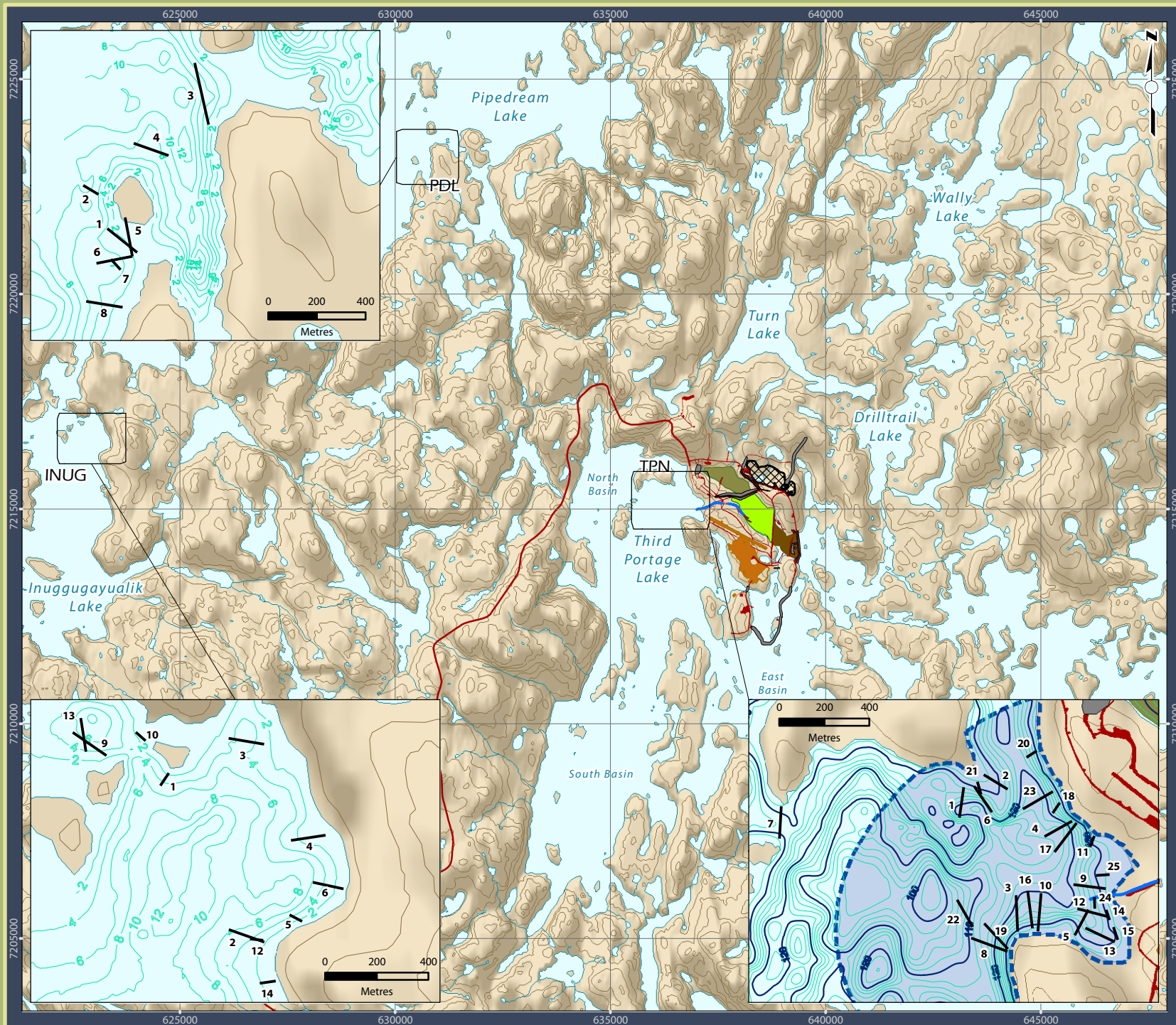
Natural Resources Canada, GeoBase®
National Topographic Database
Agnico-Eagle Mines Limited.
Azimuth Consulting Group Inc.

Meadowbank Gold Project

Prepared
for:



By:



4.2. QA/QC

The fish sampling SOP (**Appendix F**) was developed to ensure that the data quality objectives stipulated in the Study Design (Azimuth, 2011) were met. Two senior scientists were on site for the first week of the program to tailor the SOP (with any last minute modifications needed to adapt to the specific situation encountered in the field) and to ensure that its contents were all well understood by the experienced field crew.

In addition to the detailed SOP, the primary QA method in the field involved the completion of data sheets to provide a record and hard copy of relevant observations and to ensure that all relevant information was collected in the field. Reliable sample tracking, logging, and data recording was practiced and documented to establish continuity between the sample collected and the results reported. Raw fish data as entered into excel (e.g., database), is presented in **Appendix H**.

Although much of the survey information is collected while in the field, variables such as gonad and liver weights and fecundity/egg size were determined at an on-site “laboratory” facility. Again, in addition to the detailed SOP, data sheets were used to ensure that all relevant information was properly collected and recorded for each fish, along with any specific comments (**Appendix G**). Weight scales were selected for appropriate precision and tested (and calibrated if needed) for accuracy prior to their use and periodically through the study.

Finally, all fish otoliths and pectoral fin rays were sent to North South Consultants (Winnipeg, MB) for ageing analysis. Regardless of the preparation method used, all structures were viewed a minimum of two times, and, if consistency was not met between the first two reads, a third was undertaken. If consistency could not be accomplished within three reads the structure was deemed un-ageable and no age was assigned. All readings were conducted as “blind” (independent from each other). A minimum of 10 % QA/QC was then performed on the structures. Over 75% of the repeat age determinations were the same as the final age determinations (**Table 4.2-1**), and of the remaining repeats, RPD values were always less than 11% (i.e., final and repeat age determination did not differ by more than 2 years; in most cases they differed by only 1 year). Raw fish ageing data is presented in **Appendix I**.

Overall, the quality assurance measures stipulated in the Study Design were implemented and the data quality of the resulting fish data set is considered appropriate for this EEM study.

Table 4.2- 1. QA/QC data from fish ageing analysis (2 pages; see notes at bottom of table).

Fish Species	Age Structure	Sample ID	Final Age Determination (yrs)	Repeat Age Determination (yrs)	RPD (%)
ARCH	OT	INUG-07-15	5	5	0
LKTR	OT	INUG-02-05	8	8	0
LKTR	OT	INUG-05-07	23	24	-4.3
LKTR	OT	INUG-07-12	8	8	0
LKTR	OT	INUG-13-17	20	19	5.1
RNWH	OT	INUG-01-15	9	9	0
RNWH	OT	INUG-08-04	16	16	0
RNWH	OT	INUG-13-14	15	14	6.9
ARCH	OT	TPN-25-03	5	5	0
ARCH	OT	TPN-10-03	3	3	0
LKTR	OT	TPN-06-21	6	6	0
LKTR	OT	TPN-10-22	18	19	-5.4
LKTR	OT	TPN-16-12	22	22	0
LKTR	OT	TPN-24-05	29	28	3.5
ARCH	OT	PDL-03-06	6	6	0
LKTR	OT	PDL-02-01	10	10	0
LKTR	OT	PDL-03-15	22	22	0
LKTR	OT	PDL-04-12	6	6	0
RNWH	OT	PDL-01-08	12	12	0
RNWH	OT	PDL-03-18	16	15	6.5
LKTR	FR	INUG-01-09	5	5	0
LKTR	FR	INUG-05-05	15	15	0
LKTR	FR	INUG-08-17	16	17	-6.1
LKTR	FR	INUG-14-01	5	5	0
RNWH	FR	INUG-01-14	8	8	0
RNWH	FR	INUG-01-21	14	14	0
RNWH	FR	INUG-08-11	12	12	0
RNWH	FR	INUG-13-12	14	14	0
RNWH	FR	INUG-14-11	12	12	0
ARCH	FR	TPN-10-07	10	10	0
ARCH	FR	TPN-25-09	10	9	10.5
ARCH	FR	TPN-06-11	8	8	0
ARCH	FR	TPN-10-04	2	2	0

Fish Species	Age Structure	Sample ID	Final Age Determination (yrs)	Repeat Age Determination (yrs)	RPD (%)
LKTR	FR	TPN-02-02	11	11	0
LKTR	FR	TPN-05-14	3	3	0
LKTR	FR	TPN-05-20	21	22	-4.7
LKTR	FR	TPN-06-29	14	14	0
LKTR	FR	TPN-11-04	5	5	0
LKTR	FR	TPN-16-15	18	18	0
LKTR	FR	TPN-24-01	6	6	0
LKTR	FR	TPN-05-01	10	9	10.5
LKTR	FR	TPN-06-02	15	15	0
LKTR	FR	TPN-10-09	4	4	0
LKTR	FR	TPN-11-01	5	5	0
LKTR	FR	TPN-16-14	11	10	9.5
LKTR	FR	TPN-25-01	9	9	0
RNWH	FR	TPN-11-06	14	14	0
LKTR	FR	PDL-04-02	20	21	-4.9
LKTR	FR	PDL-08-04	6	6	0
LKTR	FR	PDL-03-03	5	5	0
LKTR	FR	PDL-04-04	16	16	0
LKTR	FR	PDL-08-05	4	4	0
RNWH	FR	PDL-01-07	8	8	0
RNWH	FR	PDL-03-18	12	11	8.7
ARCH	FR	INUG-02-13	4	4	0
LKTR	FR	INUG-02-09	14	14	0
LKTR	FR	INUG-06-01	5	5	0
LKTR	FR	INUG-08-03	13	13	0
LKTR	FR	INUG-13-11	14	14	0
LKTR	FR	INUG-14-12	20	22	-9.5

Notes: RPD = Relative Percent Difference (%) = ((original - duplicate) / (original + duplicate)/2) x 100; ARCH: Arctic char; LKTR: lake trout; RNWH: round whitefish; OT = otolith; FR = pectoral fin ray.

4.3. Data Assessment

4.3.1. Catch Summary and Target Endpoints

As described in **Section 2.3**, the fish survey had targeted non-lethal sampling of lake trout (*Salvelinus namaycush*) and lethal sampling of round whitefish (*Prosopium cylindraceum*). However, catch (summarized in **Table 4.3-1** and **Figure 4.3-1**) was insufficient for round whitefish, and an alternative, Arctic char (*Salvelinus alpinus*), particularly at TPN.

Figures 4.3-2 and 4.3-3 show maturity status and reproductive status (of mature fish) by species and lake (area). These figures show the paucity of appropriate data for the lethal study due to the low numbers of ripe individuals of any species. While plans were adapted to collect the full suite of lethal sampling endpoints on incidental lake trout mortalities, the final sample sizes for gonad weight (**Table 4.3-2**) or liver weight (**Table 4.3-3**) were too low across all species to make any meaningful insights regarding effluent-related effects. As shown in **Figures 4.3-4 and 4.3-5**, the low numbers for lake trout were primarily due to a large fraction of mature fish being in the resting phase. Consequently, this study focuses on non-lethal endpoints (summarized in **Table 4.3-4**) for lake trout only (note that outlier screening and summary statistics are shown for all three species).

In support of future studies, lake trout incidental mortality status is shown by gillnet panel mesh size (**Figures 4.3-6**), gillnet set type (i.e., day or night; **Figures 4.3-7**) and fork length (**Figure 4.3-8**). Overall, incidental mortality rates were higher than expected across all mesh sizes. There was a general pattern of relatively lower mortality rates with smaller and larger mesh sizes, but the pattern was not consistent across lakes. Not unexpectedly, mortality rates were higher for the longer overnight sets (n.b., catch was also substantially higher during the overnight sets). In general, it also appears that incidental mortality rates decrease with increasing fork length. These results will be discussed further in **Section 7**.

4.3.2. Data Quality and Outlier Identification

Data quality procedures outlined in the Study Design (Azimuth, 2011) were followed to minimize errors in the fish survey results. Notwithstanding the use of rigorous recording and transcription procedures, the data were carefully screened to identify outliers by examining the condition (k) distribution (**Figure 4.3-9**) and body weight-length relationships (**Figure 4.3-10**) by species and lake.

The following outliers were identified:

- 1) Five fish were removed from the data set for incomplete measurements (generally very small fish with no reliable weight or age structure result).
- 2) Eleven fish were removed from the data set for either abnormally low [$k < 0.75$] or high [$k > 1.4$] condition and are presumed due to errors in recording of length or weight. These are clearly shown in **Figure 4.3-10**.

A summary of outliers by species and lake is shown in **Table 4.3-5**. Updated catch results without outliers is shown in **Table 4.3-6**.

4.3.3. Summary Statistics

Key variables used to assess the non-lethal endpoints (see **Table 4.3-4**) include:

- Fork length (mm)
- Body weight (g)
- Age – pectoral fin (yrs)
- Age – otolith (yrs)

Summary statistics by species and area are shown in **Table 4.3-7**. Summary statistics for the opportunistically-collected lethal parameters are shown in **Appendix J**.

4.3.4. Visual Fish Condition

Results for external and internal visual fish condition observations are shown in **Table 4.3-8**. Key results for lake trout were:

- Similar rates of external (DELT¹⁰) abnormalities across lakes.
- No external parasites were found at all
- Minor internal abnormalities in two fish from TPN
- Internal parasite loads were similar at TPN and PDL; INUG were higher

Overall, no major differences in lake trout external or internal condition were observed among areas. No conclusions can be drawn from the round whitefish and Arctic char observations due to the limited sample sizes.

4.3.5. Modification of Age Variables for Statistical Analyses

As described in the Study Design (Azimuth, 2011), lake trout ages derived from pectoral fin rays, particularly for older fish, are less reliable than ages derived from otoliths. Consequently, age estimates from both structures were made on incidental mortalities in an effort to determine the reliability of estimates based on pectoral fin rays.

The breakdown of age structure collection by species and area is shown in **Table 4.3-9**. For lake trout, both ageing structures (i.e., “PEC/OTO”) were collected for 134 fish (126 lake trout had age estimated from pectoral fin rays only). Given the higher than expected incidental lake trout mortalities and the resulting larger data set, sufficient data were available to determine the

¹⁰ DELT is a detailed visual examination focusing on specific deformities, morbidities or abnormalities present externally (i.e., deformities, erosions, lesions and tumors). We use the term herein to also describe the internal fish health examinations.

statistical relationship between the two ageing methods. This relationship could then be used to correct the pectoral fin ray ages.

The raw (i.e., untransformed) relationship between PEC-based and OTO-based age estimates is shown in **Figure 4.3-11** relative to a 1:1 relationship (red dashed line). Overall, the PEC-based age estimates are biased about 5 years lower than the OTO-based ages. There was a pattern of increasing variance with increasing age, so variables were $\sqrt{x+0.5}$ -transformed to run the final linear regression model. The final model used to correct PEC-based age estimates to be consistent with OTO-based age estimates was:

$$\text{Age}_{\text{OTOpred}} = (0.966 + (0.915 * (\text{PEC} + 0.5)^{0.5}))^2 - 0.5$$

$p < 0.001$ Adjusted R-squared: 0.77

The corrected PEC age estimates were used in all subsequent analyses.

4.3.6. Statistical Analyses

As described in **Section 2.2** and shown in **Figure 2.2-1**, a hierarchical approach was used to test for statistical differences among sampling areas. Initial analyses were conducted to test for differences between the pooled reference areas (i.e., INUG and PDL pooled as “REF” for reference) and TPN (as “EXP” for exposure); this is hereafter referred to as “C-I” (or “CI-based”). If statistical differences were identified, then analyses would test for differences among areas (i.e., testing to see if TPN is different from both INUG and PDL) (referred to hereafter as “area-based”).

The statistical procedures recommended by the EEM technical guidance document (Environment Canada, 2012) are shown in **Table 4.3-10**. The fish survey data from the non-lethal lake trout study has two statistical analyses: analysis of covariance (ANCOVA) and the two-sample Kolmogorov-Smirnov (K-S) test. These analyses are described in more detail in the following sections.

4.3.6.1. ANCOVA

As discussed in the EEM technical guidance document (Environment Canada, 2012), analysis of covariance (ANCOVA) has four key assumptions:

- The relationship between the response and covariate is linear;
- The slopes of the regression lines among areas are parallel;
- The covariate is fixed and measured without error; and
- The residuals are normally distributed and independently distributed with zero mean and a common variance.

The steps followed for endpoints analyzed using ANCOVA were:

- 1) Plot the relationships between response variable (e.g., weight for the condition endpoint) and covariate (e.g., length for the condition endpoint), highlighting C – I groups.

- 2) Examine residuals graphically to determine the need for transformations.
- 3) Conduct linear regression analysis of relationship for each group (C, I) to ensure that the underlying relationships are linear for both groups.
- 4) Run ANCOVA model with interaction term to assess whether slopes can be assumed parallel between groups (slopes assumed parallel if $p > 0.05$ for interaction term).
- 5) If slopes are parallel, then run ANCOVA model without interaction to determine whether the intercepts differ significantly by group (C, I).
- 6) Examine final ANCOVA model residuals graphically to check for underlying patterns that would violate model assumptions of normality and homogeneity of variance.
- 7) Covariate data were centered (at fork length = 500 mm or age = 20 years) and the ANCOVA model re-run to compute meaningful least squares mean (LSM) estimates for each group (n.b., centering a covariate does not change the other general model results). These were then back-transformed to estimate effect size in the I group relative to the C group.

CI-Based ANCOVA results are presented in **Table 4.3-11**; an overview by endpoint is provided below.

Growth (body weight against age)

- Response variable and covariate were both log-transformed (**Figure 4.3-12**) based on visual examination of the variables in untransformed, log-linear, and log-log space. While age usually does not require transformation, there was a distinct curvature to the residuals when only body weight was log-transformed.
- The linear regressions models fit for each group (C and I) were highly significant (**Table 4.3-11**), explaining over 85% of the variability in each of the groups.
- The interaction term was not statistically significant ($p=0.48$), so the slopes were assumed parallel.
- There was no significant difference between C and I intercepts ($p=0.21$).
- Visual observations of ANCOVA model residuals showed no major departures from model assumptions.
- The observed effect size based on the modeled means was 1.3%

Growth (fork length against age)

- Response variable and covariate were both log-transformed (**Figure 4.3-13**) based on visual examination of the variables in untransformed, log-linear, and log-log space. As mentioned above, there was a distinct curvature to the residuals when age was not transformed.

- The linear regressions models fit for each group (C and I) were highly significant (**Table 4.3-11**), explaining over 85% of the variability in each of the groups.
- The interaction term was not statistically significant ($p=0.44$), so the slopes were assumed parallel.
- There was no significant difference between C and I intercepts ($p=0.11$).
- Visual observations of ANCOVA model residuals showed no major departures from model assumptions.
- The observed effect size based on the modeled means was 0.8%

Condition (body weight against fork length)

- Response variable and covariate were both log-transformed (**Figure 4.3-14**) based on visual examination of the variables in untransformed and log-log space.
- The linear regressions models fit for each group (C and I) were highly significant (**Table 4.3-11**), explaining over 99% of the variability in each of the groups.
- The interaction term was not statistically significant ($p=0.08$), but it was marginal. The assumption of slope equality in this case is fairly robust given that the actual slopes of the individual regressions were 3.05 and 3.00 (only a 1.7% slope difference in I relative to C).
- There was no significant difference between C and I intercepts ($p=0.45$).
- Visual observations of ANCOVA model residuals showed no major departures from model assumptions.
- The observed effect size based on the modeled means was -0.9%.

Given that no differences were found in the CI-based ANCOVA analyses, no area-based ANCOVA analyses were conducted.

4.3.6.2. Kolmogorov-Smirnov (K-S) Test

The Kolmogorov-Smirnov (K-S) test is a non-parametric test for equality of continuous, one-dimensional probability distributions. While the one-sample version of the test can be used to compare a sample to a reference probability distribution, the two-sample test recommended by the EEM TGD (Environment Canada, 2012) compares two sample distributions. As shown in **Table 4.3-10**, this statistical test was used to determine whether the length (or age) frequency distributions differed significantly between C and I groups.

The steps followed for endpoints analyzed using the K-S test were:

- 1) Estimated the empirical cumulative density function (ECDF) for each group (C-I).

- 2) Plotted the cumulative frequency distribution and density function for each group.
- 3) Conducted the two-sample K-S test (two-sided).

The results are presented below for each distribution type.

Length Frequency Distribution

- Cumulative frequency (**Figure 4.3-15**) and density (**Figure 4.3-16**) plots for fork length show similar distributions. Both the C and I areas show the “classic” bimodal size distribution seen in Arctic lakes¹¹.
- The K-S test showed no significant difference between the two length frequency distributions ($D=0.138$, $p=0.173$).

Age Frequency Distribution

- Cumulative frequency (**Figure 4.3-17**) and density (**Figure 4.3-18**) plots for age show similar distributions. Again, both the C and I areas show the anticipated bimodal size distribution.
- The K-S test showed no significant difference between the two age frequency distributions ($D=0.142$, $p=0.173$).

4.3.7. Post-hoc Assessment of Power and Confidence

This section examines results of statistical analyses with the intent of providing insights into the degree of confidence of their conclusions. This is typically done in different ways for each statistical procedure, but essentially focuses on understanding the degree of uncertainty (e.g., using 95% confidence intervals) for observed effect size estimates in the context of the EEM-recommended critical effect size (CES). *Post-hoc* power analyses can also provide insights for planning subsequent EEM cycles.

For endpoints analyzed using ANCOVA, the following approaches were taken:

- 1) Comparisons of observed effect sizes and their 95% confidence intervals to the EEM-recommended critical effect sizes (CES) to determine the confidence that the observed effects do not exceed the CES.
- 2) Estimation of statistical power of detecting the CES based on the executed study design.
- 3) Estimation of the minimum detectable effect size for the targeted statistical power of 0.9 (i.e., $\beta = 0.1$) based on the executed study design.

¹¹ As discussed in the Study Design (Azimuth, 2011), the magnitude of the bimodal result is likely an artefact of gillnet sampling as the two fish out programs conducted at Meadowbank showed much higher numbers of smaller fish after all fish had been caught from two large impoundment areas. That said, bimodality of some degree is common in size frequency distributions for fish Arctic lakes (Johnson, 2002).

- 4) Estimation of minimum sample size required to detect the CES based on the targeted statistical power.

Key results for the ANCOVA endpoints were as follows:

- *Observed Effect Sizes* – observed effect sizes and their 95% confidence limits are shown in **Figure 4.3-19** relative to the respective EEM CES. Simulations (i.e., random generation of data based on the properties of the existing data) were used to quantitatively estimate the confidence that the observed effect size (ES) was less than the CES; confidence was high for all three ANCOVA variables (**Table 4.3-12**). As an example, **Figure 4.3-20** shows the cumulative response ratio distribution (i.e., exposure mean/reference mean) compiled from 1000 simulations; 0 cases were below the CES of 25% (0.75 in response ratio).
- *Power to Detect CES* – the statistical power equations from the EEM TGD (Environment Canada, 2012) were used to estimate power (see textbox). Power to detect the CES was high for all three endpoints (**Table 4.3-12**).
- *Minimum Effect Size for Target Power* – the statistical power equations from the EEM TGD (Environment Canada, 2012) were used to estimate minimum detectable effect size for power $(1-\beta) = 0.9$ (see textbox). The minimum detectable effect sizes were all lower than the CES (**Table 4.3-12**). The growth endpoint had the highest minimum detectable effect size at 19.1%; both condition and length-at-age had low minimum detectable effect sizes (4.4% and 5.6%, respectively).
- *Minimum Sample Size for CES with Target Power* – the statistical power equations from the EEM TGD (Environment Canada, 2012) were used to estimate minimum sample size required to detect the EEM CES with power $(1-\beta) = 0.9$ (see textbox). The minimum sample sizes ranged from 7 fish to 61 fish (**Table 4.3-12**), well below the target sample size of 100 fish for a non-lethal study.

Post-hoc Power Analyses for ANCOVA

EEM TGD provides the following equation for determining ANCOVA power:

$$t_{\beta} = \sqrt{\frac{n}{2((SD_z)/(CES_z))^2}} - t_{\alpha}$$

Where:

SDz = sqrt(mean squares error)

CESz = effect size

n = sample size

t = t value for $\beta_{(1 \text{ tail})}$ or $\alpha_{(2 \text{ tail})}$

The equation can also be solved for CES (i.e., to determine minimum effect size) or n (i.e., to determine minimum sample size).

EEM TGD provides no guidance on assessing confidence or power for the Kolmogorov-Smirnov (K-S) test used to compare the age and size frequency distributions. Unlike parametric tests like ANOVA or t-test that essentially compare means (as an indicator of magnitude) and assume that the underlying distribution is normal, the K-S test takes both distribution shape and magnitude into consideration. Unfortunately, while the concept of CES (which is based on magnitude alone)

is easily applied to ANOVA, it is not apparent how it can apply to frequency distributions where both shape and magnitude are important. Consequently, we relied only on presentation of statistical confidence bands of the reference area ECDF to help provide some context for the observed results. Given that two ECDFs with minor differences will have statistically significant differences when sample size (n) is sufficiently high, this approach is limited by the lack of a pre-defined ecologically-relevant effect size (i.e., so that effects are defined in terms of ecological significance rather than statistical significance).

The confidence bands were constructed for each reference area ECDF (i.e., age or length) as follows:

- 1) Calculate the *Dvoretzky-Kiefer-Wolfowitz inequality* value (epsilon) using the following equation: $\varepsilon = \sqrt{\frac{1}{2n} \log\left(\frac{2}{\alpha}\right)}$. Epsilon represents the predicted difference based on sample size (n = 100) and $\alpha = 0.025$ for the 95% confidence band (i.e., the band ranging from 2.5% to 97.5%).
- 2) Add (for upper band) or subtract (for lower band) the epsilon value from the reference ECDF to construct upper and lower confidence bands, respectively.

The resulting 95% confidence bands are shown for the age and length ECDFs in **Figure 4.3-21** and **Figure 4.3-22**, respectively. For both age and length, the exposure area ECDFs are quite close to the reference area confidence bands (e.g., ages 13 – 15 years or fork length 375 – 500 mm), highlighting a difference (i.e., the pooled reference area has more younger/smaller fish) that is not statistically significant, but reasonably close.

More insight into this pattern can be provided from looking at the ECDFs and density plots for the individual areas (**Figures 4.3-23 through 4.3-26**), which both show that TPN and INUG are quite similar (K-S test results for no significant difference for both age and length). PDL, however, shows age and length distributions that are quite different from the two other sites (K-S test results were significant [$p < 0.1$] for the age and length distributions for both TPN-PDL and INUG-PDL). This provides more confidence from an ecological perspective that the age and length frequency distributions at TPN are not meaningfully different from both reference areas.

Table 4.3- 1. Catch results across area and species.

	LKTR	RNWH	ARCH
TPN	125	2	33
INUG	88	39	5
PDL	58	7	8

Notes: these catch data were calculated prior to outlier removal.

Table 4.3- 2. Mature fish with gonad weight measurements by sex, area and species.

		LKTR	RNWH	ARCH
TPN	F	0	1	0
	M	0	1	0
INUG	F	6	14	1
	M	2	20	0
PDL	F	4	4	0
	M	3	2	0

Table 4.3- 3. Fish with liver weight measurements by sex, area and species.

		LKTR	RNWH	ARCH
TPN	F	0	1	0
	M	0	0	0
INUG	F	19	17	1
	M	19	20	0
PDL	F	17	4	1
	M	13	3	2

Table 4.3- 4. Primary and supporting endpoints for non-lethal lake trout study.

Endpoint Type	Effect Indicator	EEM Targeted Endpoints for Non-lethal Survey	Non-lethal Endpoints for Meadowbank Fish Survey	Rationale for Difference
Primary	Growth (<i>Energy Use</i>)	Size (length and weight) of YOY (age 0+) at end of growth period	Size-at-age (body weight against age)	As per Study Design, YOY not targeted; endpoint selected is consistent with lethal study
	Reproduction (<i>Energy Use</i>)	Relative abundance of YOY (% composition of YOY)	None	As per Study Design, YOY not targeted
	Condition (<i>Energy Storage</i>)	Body weight relative to length	Body weight relative to length	No difference
	Survival	Length frequency distribution	Length frequency distribution	No difference
Supporting	(<i>Energy Use</i>)	Body weight (whole)	Not used	Redundant to body wt against age used for Growth (see above)
		Length	Not used	Redundant to length frequency used for survival (see above) and length against age (see below)
		Size-at-age (length against age)	Size-at-age (length against age)	No difference

Table 4.3- 5. Outliers across area and species.

	LKTR	RNWH	ARCH
TPN	8	0	2
INUG	2	0	1
PDL	3	0	0

Table 4.3- 6. Catch results across area and species.

	LKTR	RNWH	ARCH
TPN	117	2	31
INUG	86	39	4
PDL	55	7	8

Table 4.3- 7. Summary statistics for key variables by species and area.

Species	Area	Fork Length (mm)							Body Weight (g)						
		n	mean	median	SD	SE	min	max	n	mean	median	SD	SE	min	max
LKTR	TPN	117	494	535	164	15	169	961	117	1652	1538	1504	139	50	10294
LKTR	INUG	86	480	515	169	18	152	903	86	1552	1435	1460	157	36	6750
LKTR	PDL	55	434	403	209	28	132	869	55	1493	640	1831	247	23	7600
RNWH*	TPN	2	372	372	16	12	360	383	2	584	584	48	34	550	618
RNWH*	INUG	39	374	382	40	6	241	430	39	545	570	158	25	126	884
RNWH*	PDL	7	358	377	56	21	247	412	7	571	564	237	89	158	838
ARCH*	TPN	31	351	302	160	29	126	604	31	728	294	819	147	20	2425
ARCH*	INUG	4	385	262	326	163	150	868	4	2009	172	3762	1881	40	7650
ARCH*	PDL	8	218	245	56	20	114	272	8	130	157	68	24	18	213

Species	Area	Age - Pectoral Fin (yrs)							Age - Otolith (yrs)						
		n	mean	median	SD	SE	min	max	n	mean	median	SD	SE	min	max
LKTR	TPN	111	13.5	14.0	5.8	0.5	3	26	59	16.5	17.0	8.6	1.1	3	40
LKTR	INUG	85	12.8	14.0	6.8	0.7	1	30	42	16.1	17.0	7.6	1.2	4	40
LKTR	PDL	52	11.6	10.0	7.3	1.0	2	30	31	14.1	12.0	6.1	1.1	4	29
RNWH*	TPN	2	14.0	14.0	0.0	0.0	14	14	2	14.0	14.0	0.0	0.0	14	14
RNWH*	INUG	39	11.5	12.0	2.9	0.5	4	16	38	14.6	15.0	4.1	0.7	6	24
RNWH*	PDL	7	10.1	12.0	3.8	1.4	5	14	7	14.4	14.0	4.2	1.6	9	22
ARCH*	TPN	22	5.7	5.5	3.0	0.6	1	10	17	5.1	5.0	2.3	0.6	2	11
ARCH*	INUG	4	5.8	3.5	5.6	2.8	2	14	1	5.0	5.0	--	--	5	5
ARCH*	PDL	0	--	--	--	--	--	--	5	7.6	6.0	2.7	1.2	5	11

Notes: n = sample size; SD = standard deviation; SE = standard error; min = minimum; max = maximum; -- = not applicable; * = non-sentinel species.

Table 4.3- 8. External and internal visual condition observations by area and species.

Area	Species	External Condition			Internal Condition		
		n	DELT	Parasites	n	DELT	Parasites
INUG	LKTR	88	3 fish with a dorsal fin deformity 1 fish with a tiny lower caudal fin lobe 1 fish with a bi-lobed adipose fin 1 fish with a missing adipose fin 1 fish with a scar on its stomach	None	43	None	High amount in 9% of fish Moderate amount in 35% of fish Low amount in 51% of fish None in 5% of fish Presence of cestodes in 1 fish
INUG	RNWH	39	None	None	38	None	Low amount in 5% of fish None in 95% of fish
INUG	ARCH	5	1 fish with a damaged caudal fin	None	2	None	High amount in 1 of the 2 fish Low amount in 1 of the 2 fish
PDL	LKTR	58	2 fish with a dorsal fin deformity 2 fish with a clip or nick in adipose fin 1 fish with red spots b/w pectoral fins 1 unusually skinny fish with a flaccid belly 1 fish with a small scratch on its dorsal fin	None	31	None	Moderate amount in 22% of fish Low amount in 71% of fish None in 7% of fish Presence of cestodes in 2 fish
PDL	RNWH	7	None	None	7	None	Low amount in 1 of the 7 fish None in 6 of the 7 fish
PDL	ARCH	8	1 fish with a torn left operculum	None	5	None	Low amount in 4 of the 5 fish None in 1 of the 5 fish
TPN	LKTR	125	2 fish with a clipped or damaged adipose fin 1 fish with a deformed upper caudal fin lobe 1 fish with a missing lower caudal fin lobe 1 fish with a damaged dorsal fin 1 fish with a missing adipose fin 1 unusually skinny fish	None	59	1 fish with a discoloured liver 1 fish with a pinched swim bladder	Moderate amount in 19% of fish Low amount in 64% of fish None in 17% of fish Presence of a parasite cyst in 2 fish Presence of tapeworm in 1 fish
TPN	RNWH	2	1 fish with a deformed left pectoral fin 1 fish with a stunted adipose fin	None	2	None	None
TPN	ARCH	33	1 fish missing part of left operculum 1 fish with red spots around anus and peduncle	None	16	None	High amount in 6% of fish Moderate amount in 25% of fish Low amount in 38% of fish None in 31% of fish Presence of a parasite cyst in 1 fish

Table 4.3- 9. Summary of ageing structures analyzed across areas and species.

		LKTR	RNWH	ARCH
TPN	None	6	0	5
	OTO	0	0	6
	PEC	58	0	11
	PEC, OTO	60	2	11
INUG	None	1	0	0
	OTO	0	0	0
	PEC	44	1	3
	PEC, OTO	43	38	2
PDL	None	3	0	3
	OTO	0	0	0
	PEC	24	0	0
	PEC, OTO	31	7	5

Notes: OTO = otolith; PEC = pectoral fin ray.

Table 4.3- 10. EEM-recommended statistical analyses for primary and supporting endpoints used in this study.

Endpoint Type	Effect Indicator	Non-lethal Endpoints for Meadowbank Fish Survey	Recommended Statistical Procedures
Primary	Growth (<i>Energy Use</i>)	Size-at-age (body weight against age)	ANCOVA
	Condition (<i>Energy Storage</i>)	Body weight against length	ANCOVA
	Survival	Length (or Age) frequency distribution	Kolmogorov-Smirnov
Supporting	(<i>Energy Use</i>)	Size-at-age (length against age)	ANCOVA

Table 4.3- 11. Regression and ANCOVA results for lake trout.

Area	N	Log-Transformed		R ²	Slopes Different?		Log-Transformed		Means Different?		Antilog LSM	Magnitude Difference
		Intercept	Slope		p-value	sig. at p<0.05	LSM [*]	SE	p-value	sig. @ p<0.1		
Model: log10(Weight.g)~log10(Age.yr)												
Exposure	116	-0.418	2.77	0.887	-	-	3.183	0.099	-	-	1524	-
Reference	141	-0.069	2.49	0.863	0.481	no	3.177	0.032	0.213	no	1505	1.3%
Model: log10(Length.mm)~log10(Age.yr)												
Exposure	116	1.54	0.908	0.893	-	-	2.724	0.031	-	-	530	-
Reference	141	1.64	0.827	0.869	0.436	no	2.720	0.010	0.112	no	525	0.8%
Model: log10(Weight.g)~log10(Length.mm)												
Exposure	117	-5.12	3.05	0.994	-	-	3.106	0.038	-	-	1277	-
Reference	141	-4.99	3.00	0.995	0.083	no	3.110	0.005	0.454	no	1289	-0.9%

Notes: "N" = sample size; "LSM" = least squares mean; "SE" = standard error of the mean; * LSM estimated for either a 500-mm (for length as covariate) or 20-yr old (for age as covariate) lake trout; "Age.yr" is the corrected pectoral fin ray age (see **Section 4.3.5**).

Table 4.3- 12. Power analysis information and results for lake trout endpoints tested using ANCOVA.

Endpoint Type	Effect Indicator	Non-lethal Endpoints for Meadowbank Fish Survey	Obs. ES	Mean Square of Residuals	CES	n	α	β	Confidence that ES<CES	Power for Prescribed CES	Minimum ES (%)	n for CES and Target Power
Primary	Growth (<i>Energy Use</i>)	Size-at-age (body weight against age)	1.3%	0.033	0.25	100	0.1	0.1	1.0	0.98	19.1%	61
	Condition (<i>Energy Storage</i>)	Body weight against length	-0.9%	0.002	0.1	100	0.1	0.1	1.0	0.99	4.4%	21
Supporting	(<i>Energy Use</i>)	Size-at-age (length against age)	0.8%	0.0032	0.25	100	0.1	0.1	1.0	1	5.6%	7

Notes: "Obs." = observed; "ES" = effect size (% relative to REF); "CES" = critical effect size; "n" = sample size; " α " = Type I (false positive) error rate; " β " = Type II (false negative) error rate; See text for description of last four columns.

Figure 4.3- 1. Fish survey catch results by species and sampling area.

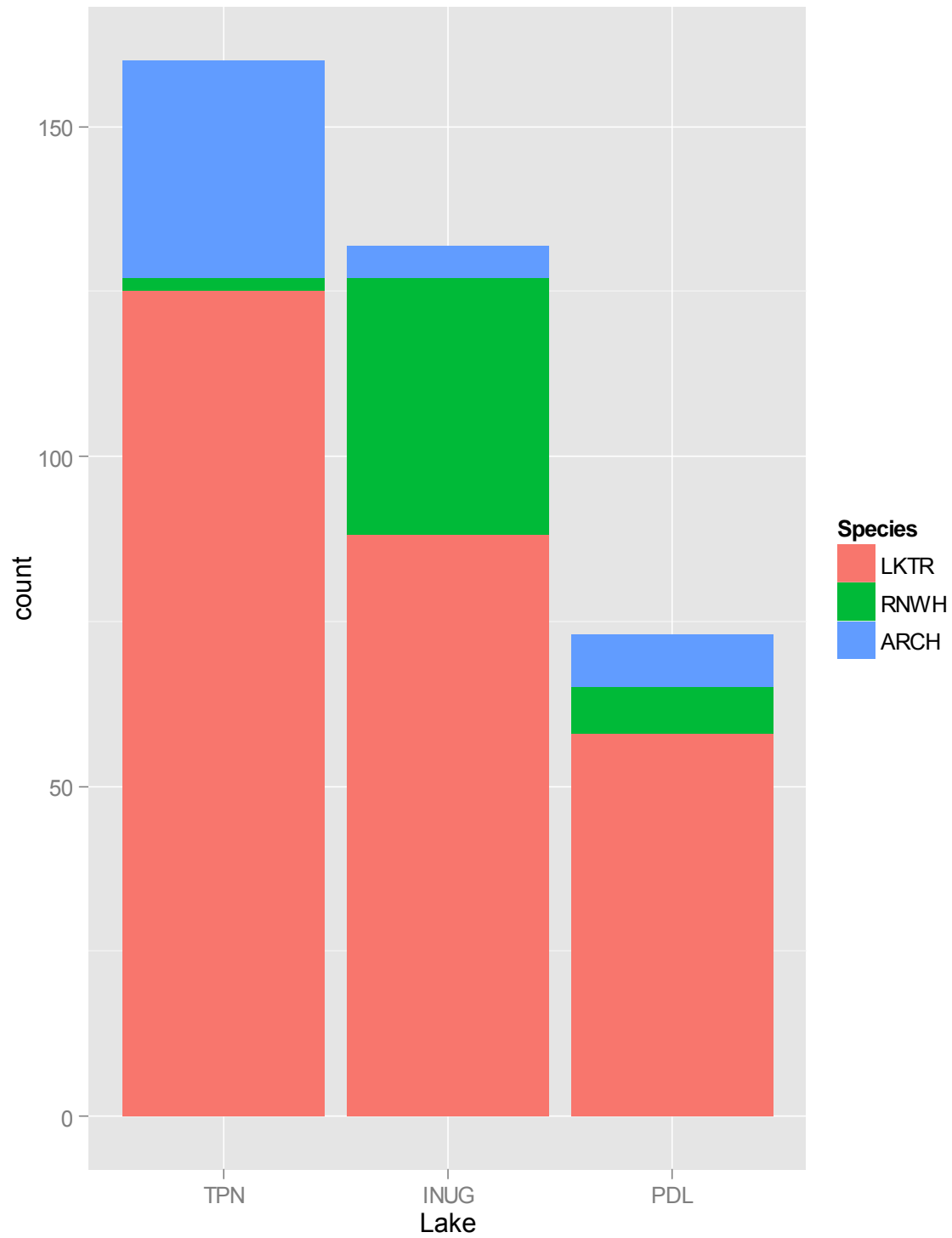


Figure 4.3- 2. Fish maturity status (blank/U=unknown, IM=immature, MA=mature) by sex, species and sampling area.

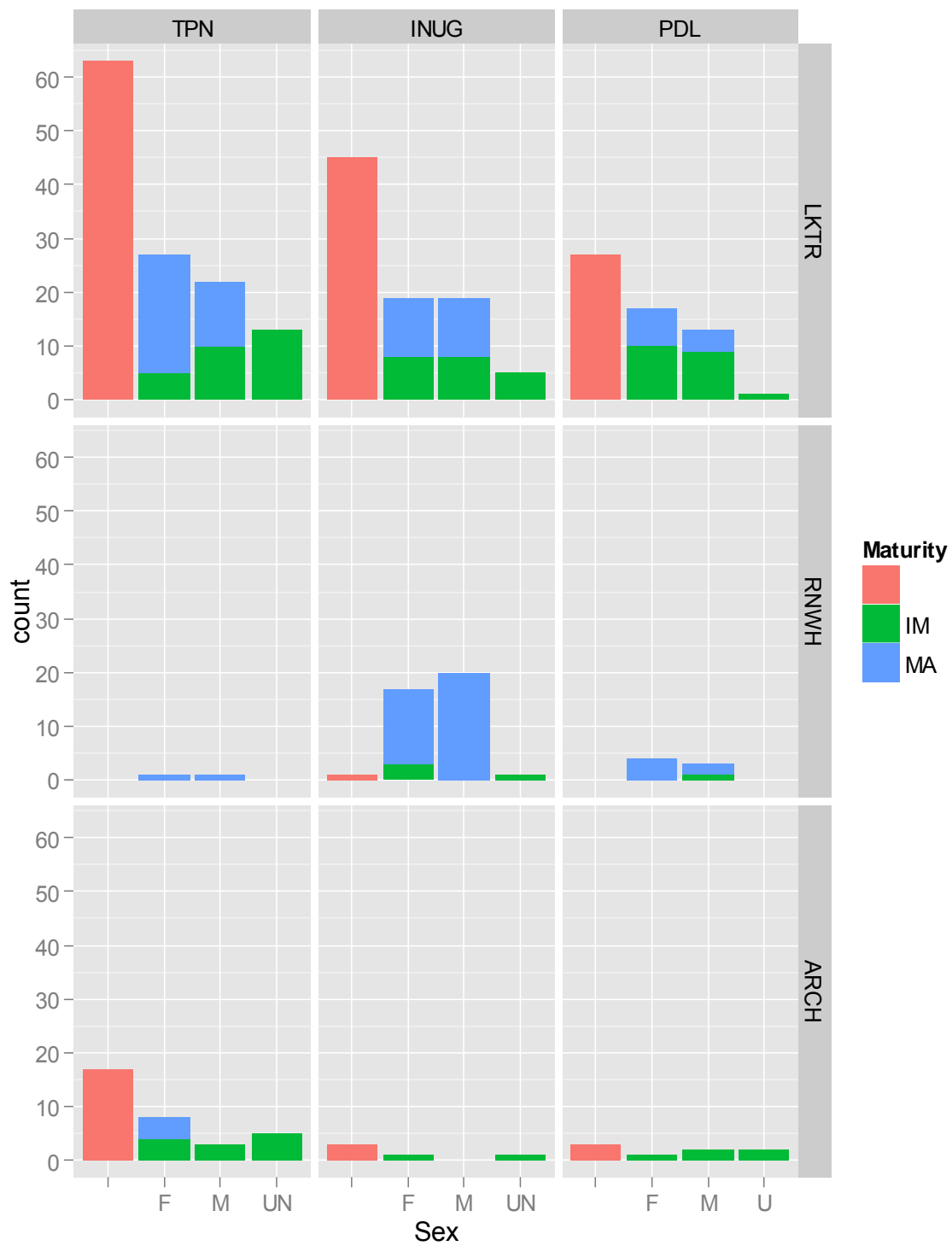


Figure 4.3- 3. Reproductive status (RI=ripe, RS=resting) of mature fish by sex, species and sampling area.

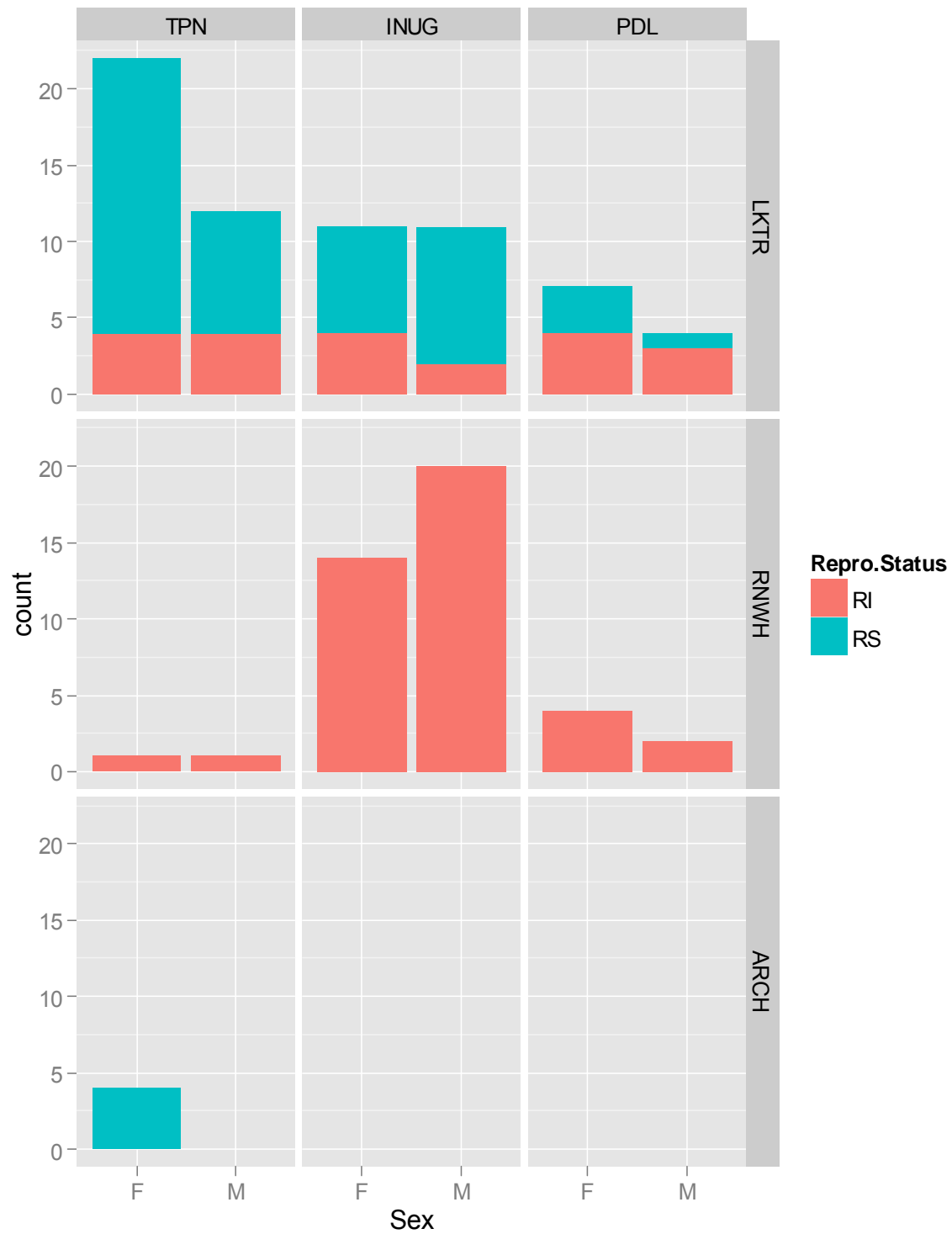


Figure 4.3- 4. Lake trout maturity status (blank/U=unknown, IM=immature, MA=mature) by fork length (mm) and sampling area.

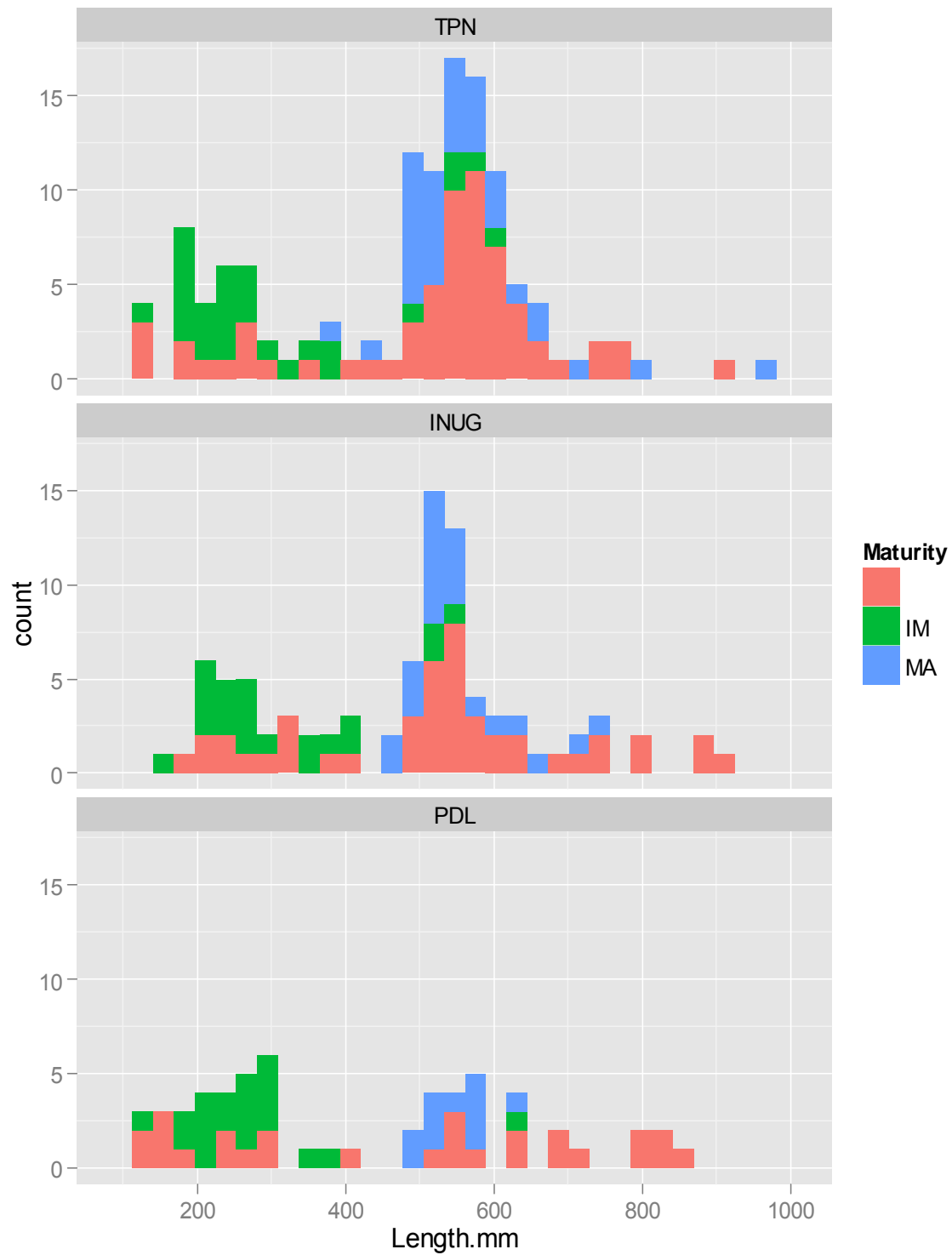


Figure 4.3- 5. Reproductive status (RI=ripe, RS=resting) of mature lake trout by fork length (mm) and sampling area.

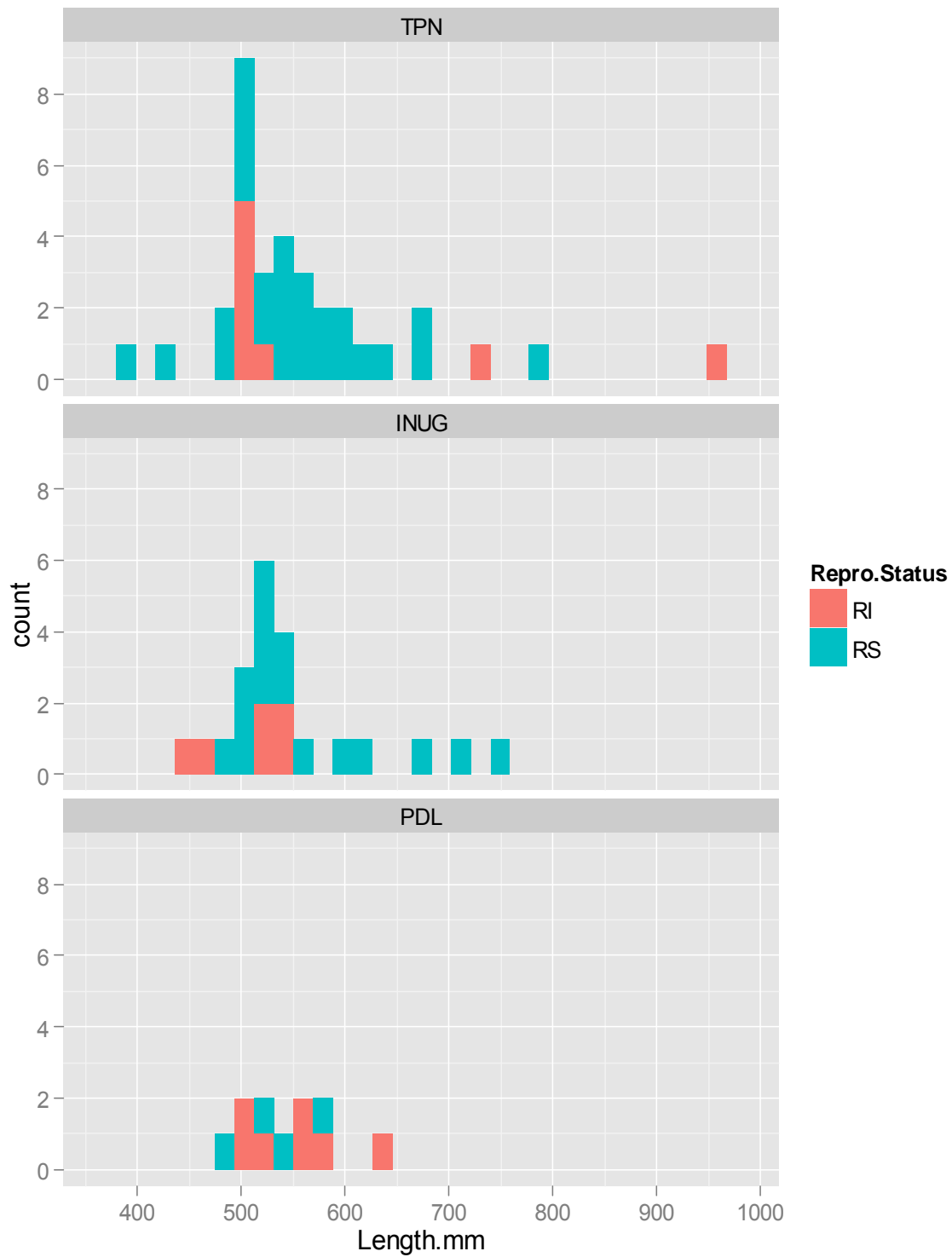


Figure 4.3- 6. Lake trout incidental mortality status (N=alive, Y=dead) by mesh size (mm) and sampling area.

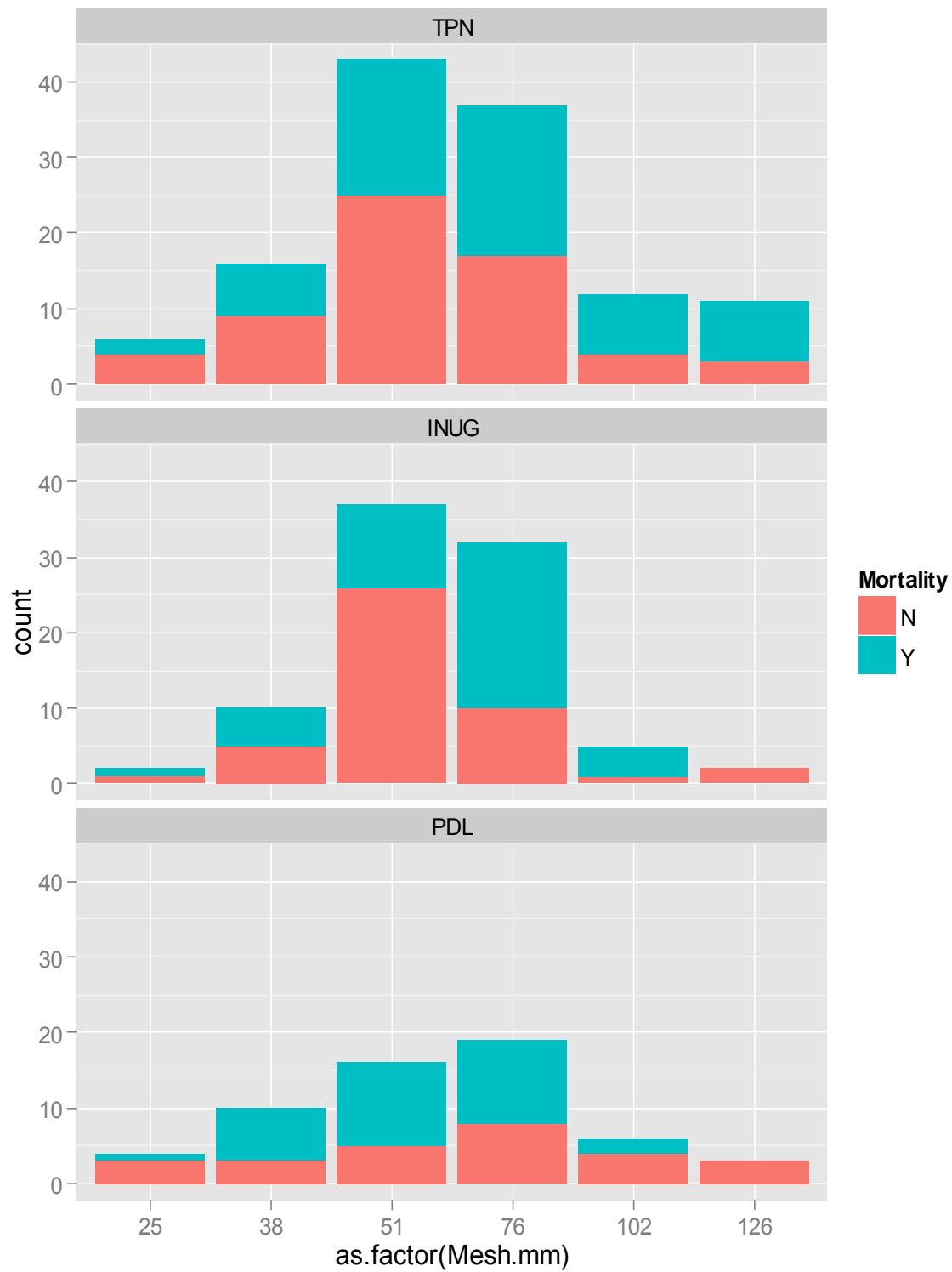


Figure 4.3- 7. Lake trout incidental mortality status (N=alive, Y=dead) by gillnet set type (day or night) and sampling area.

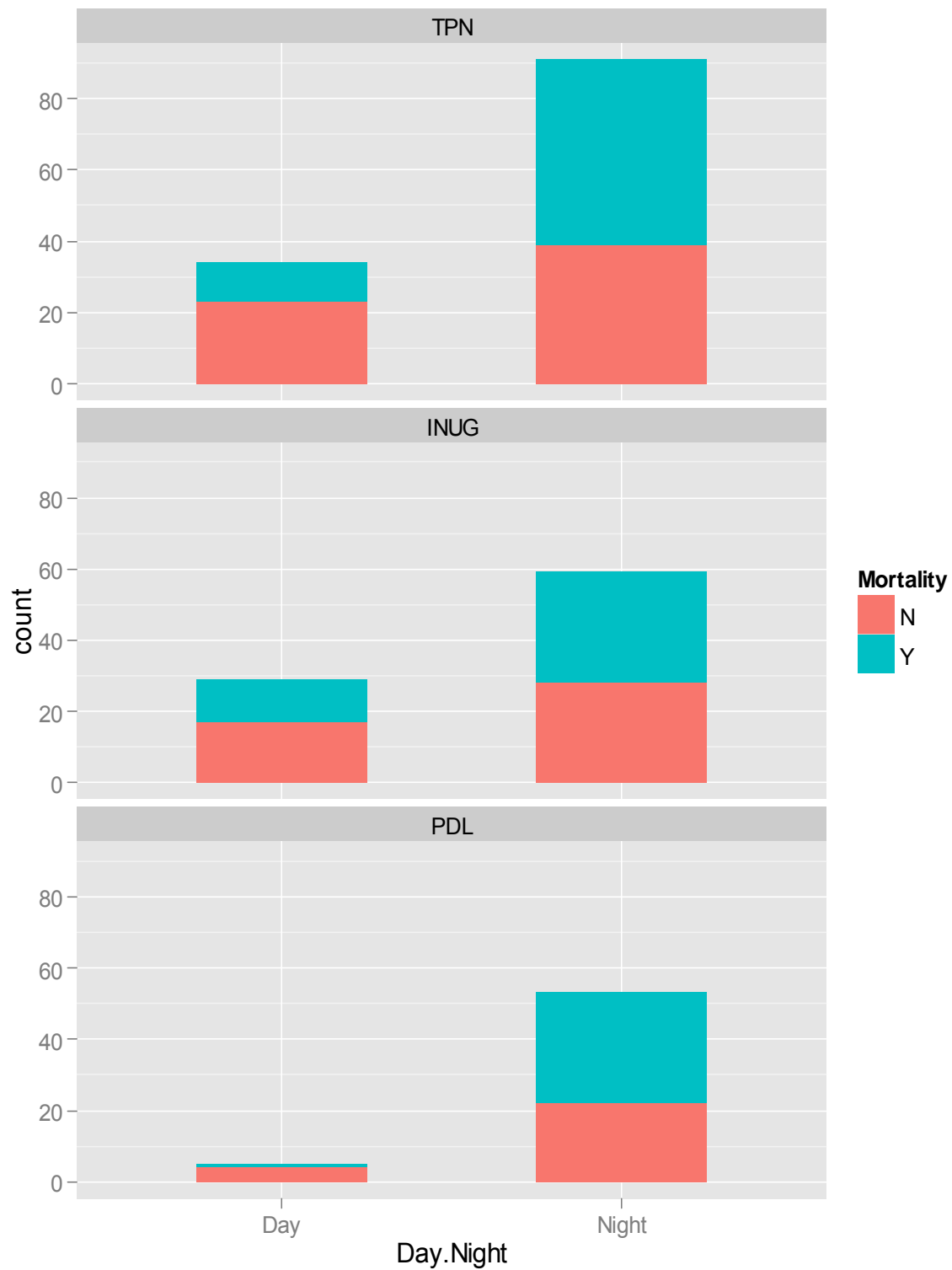


Figure 4.3- 8. Lake trout incidental mortality status (N=alive, Y=dead) by fork length (mm) and sampling area.

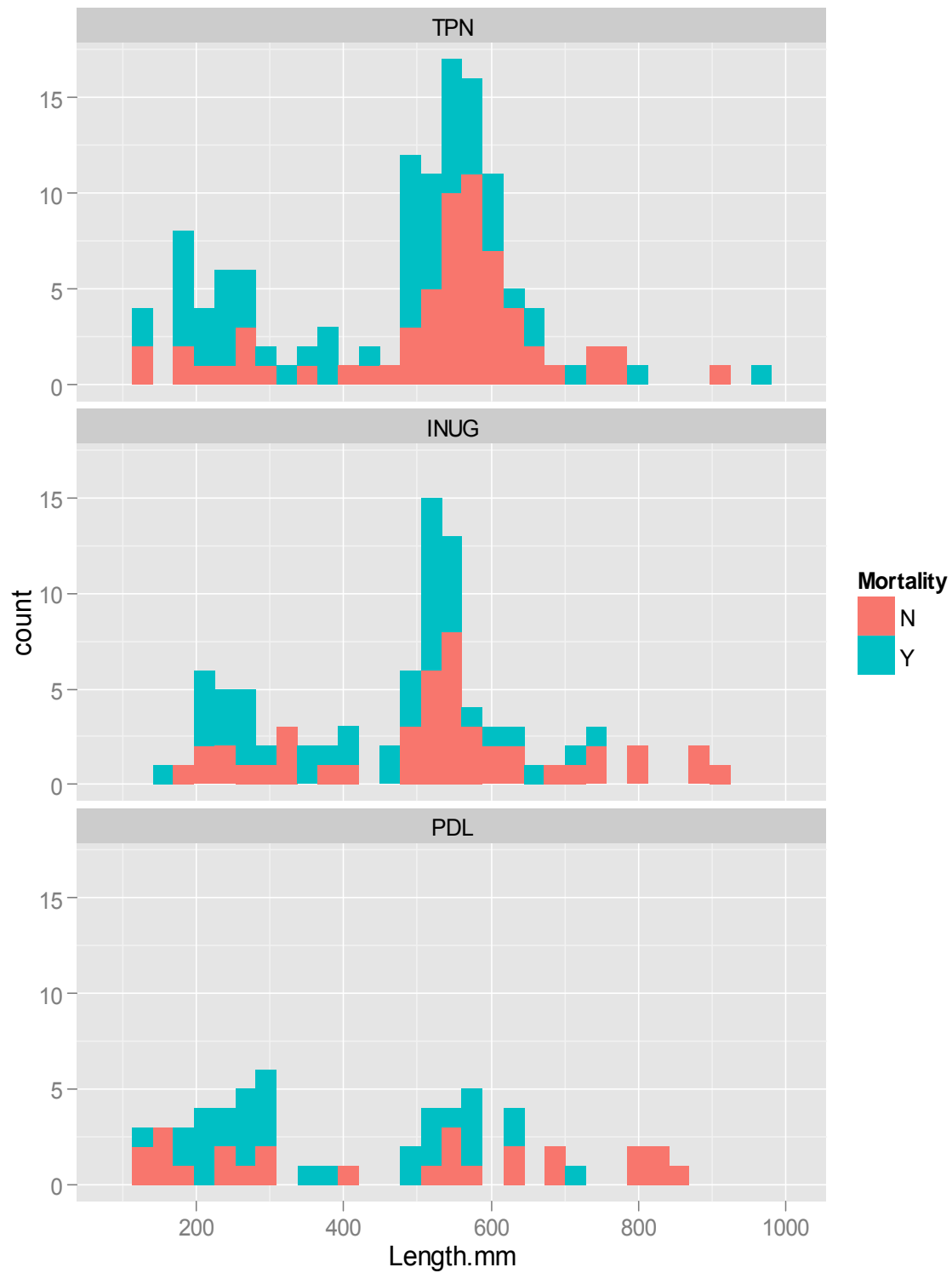


Figure 4.3- 9. Condition (k) distribution by species highlighting sampling areas.



Figure 4.3- 10. Body weight (g) – fork length (mm) plots by species highlighting sampling areas (outliers circled).

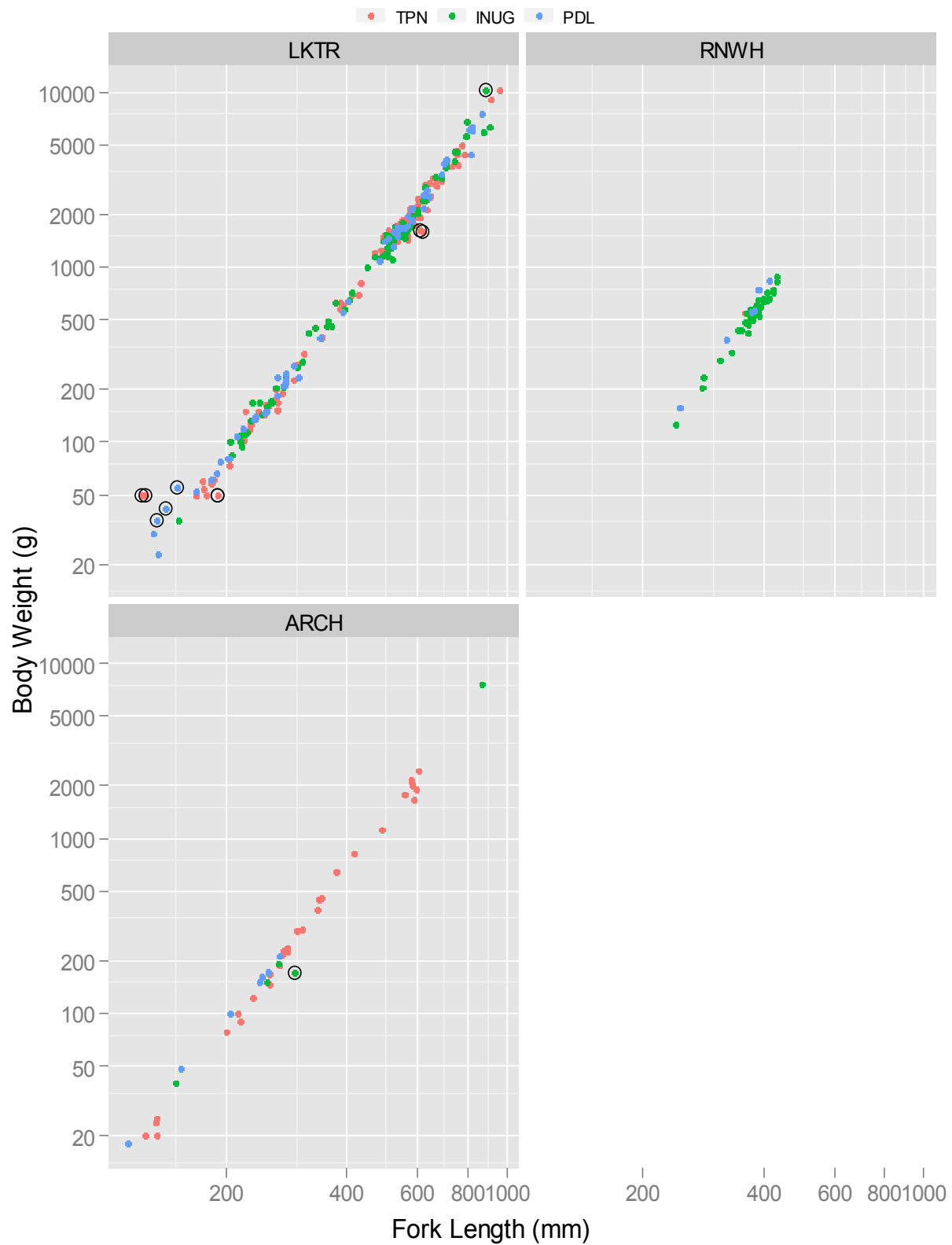


Figure 4.3- 11. Relationship between PEC-based and OTO-based age estimates (un-transformed data) relative to a 1:1 relationship (red dashed line).

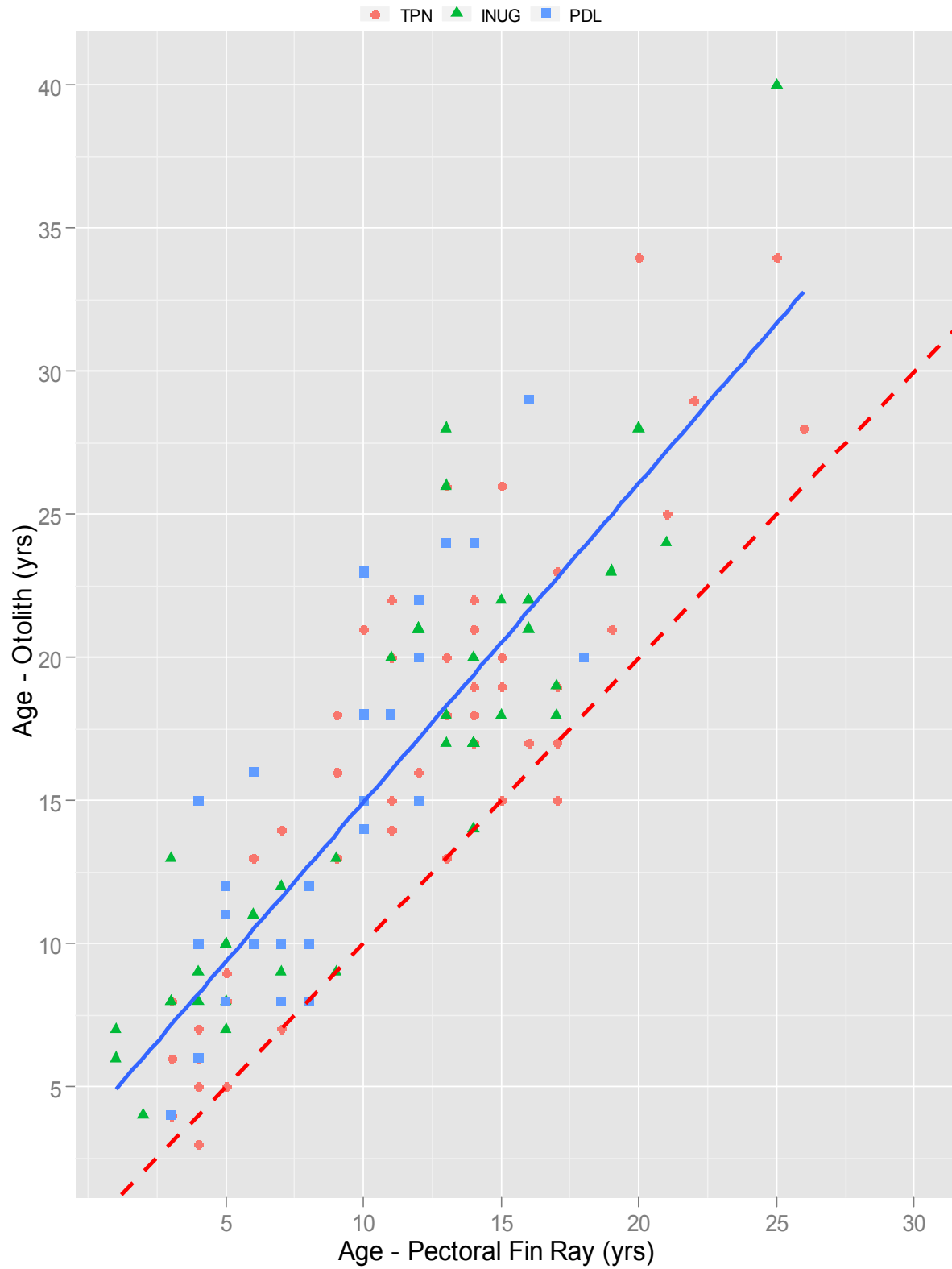


Figure 4.3- 12. Relationship between body weight (g) and age (yr) for pooled reference and exposure areas (axes shown in log space).



Figure 4.3- 13. Relationship between fork length (mm) and age (yr) for pooled reference and exposure areas (axes shown in log space).

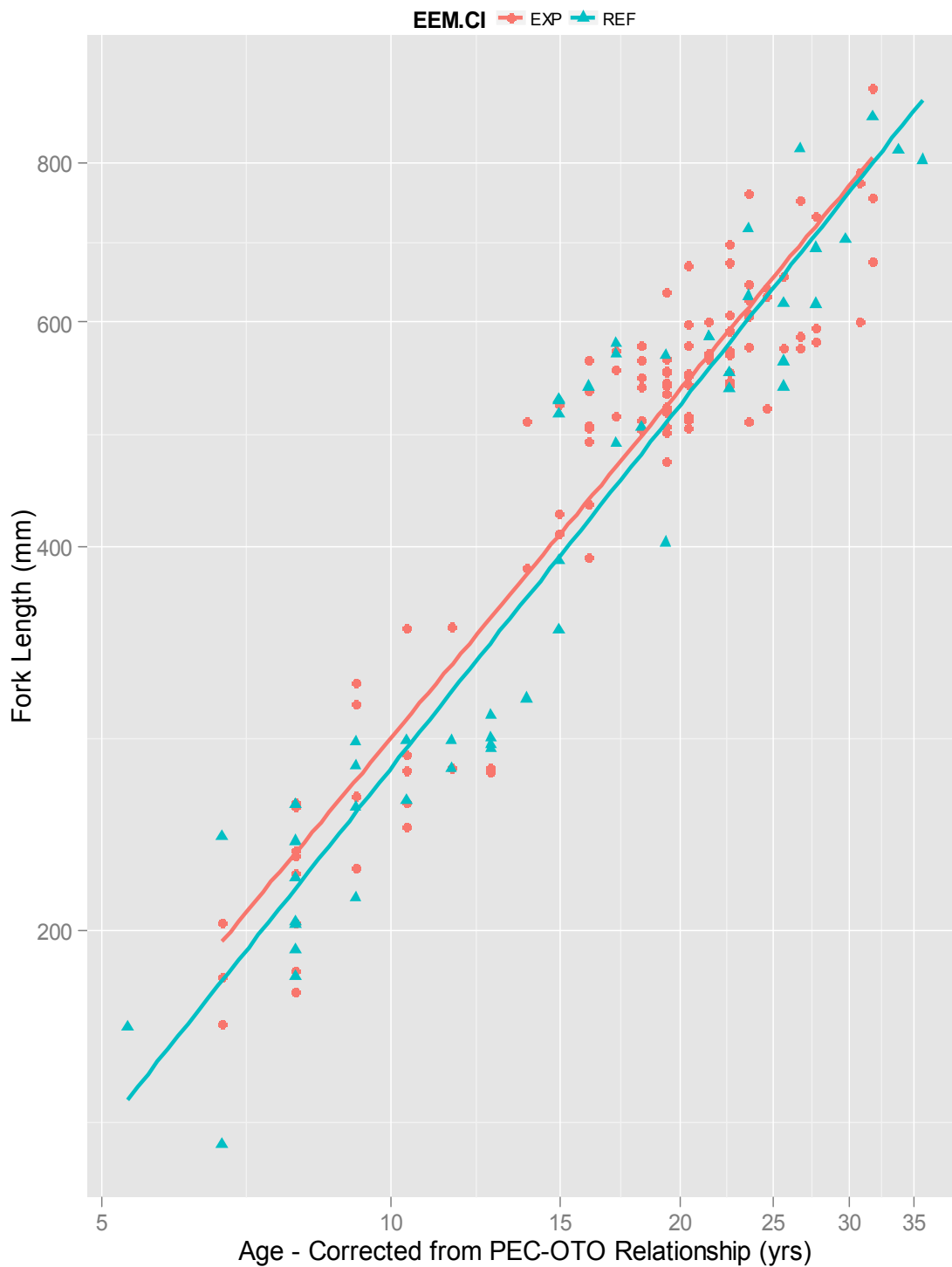


Figure 4.3- 14. Relationship between body weight (g) and fork length (mm) for pooled reference and exposure areas (axes shown in log space).

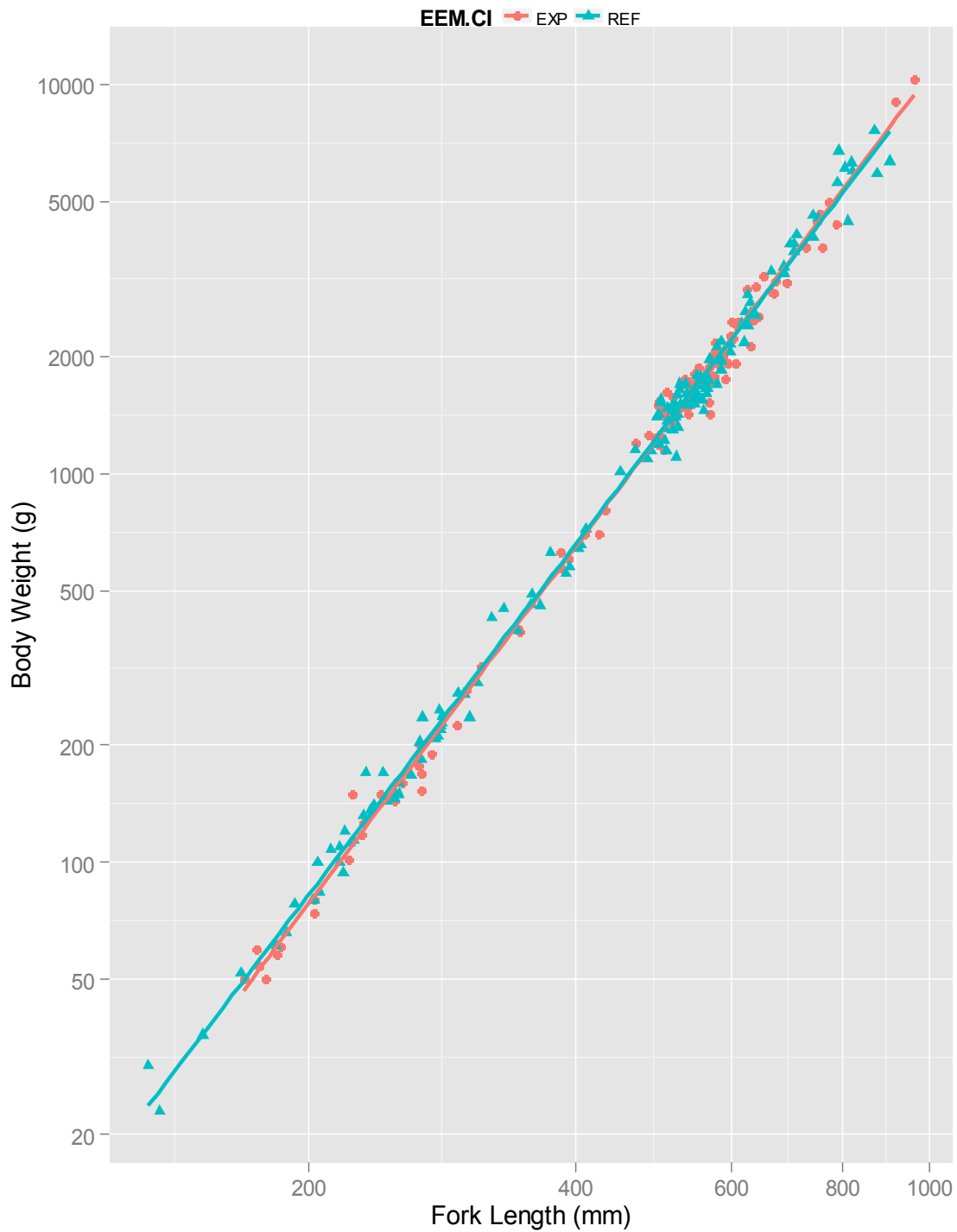


Figure 4.3- 15. Fork length (mm) empirical cumulative distribution function for pooled reference and exposure areas.

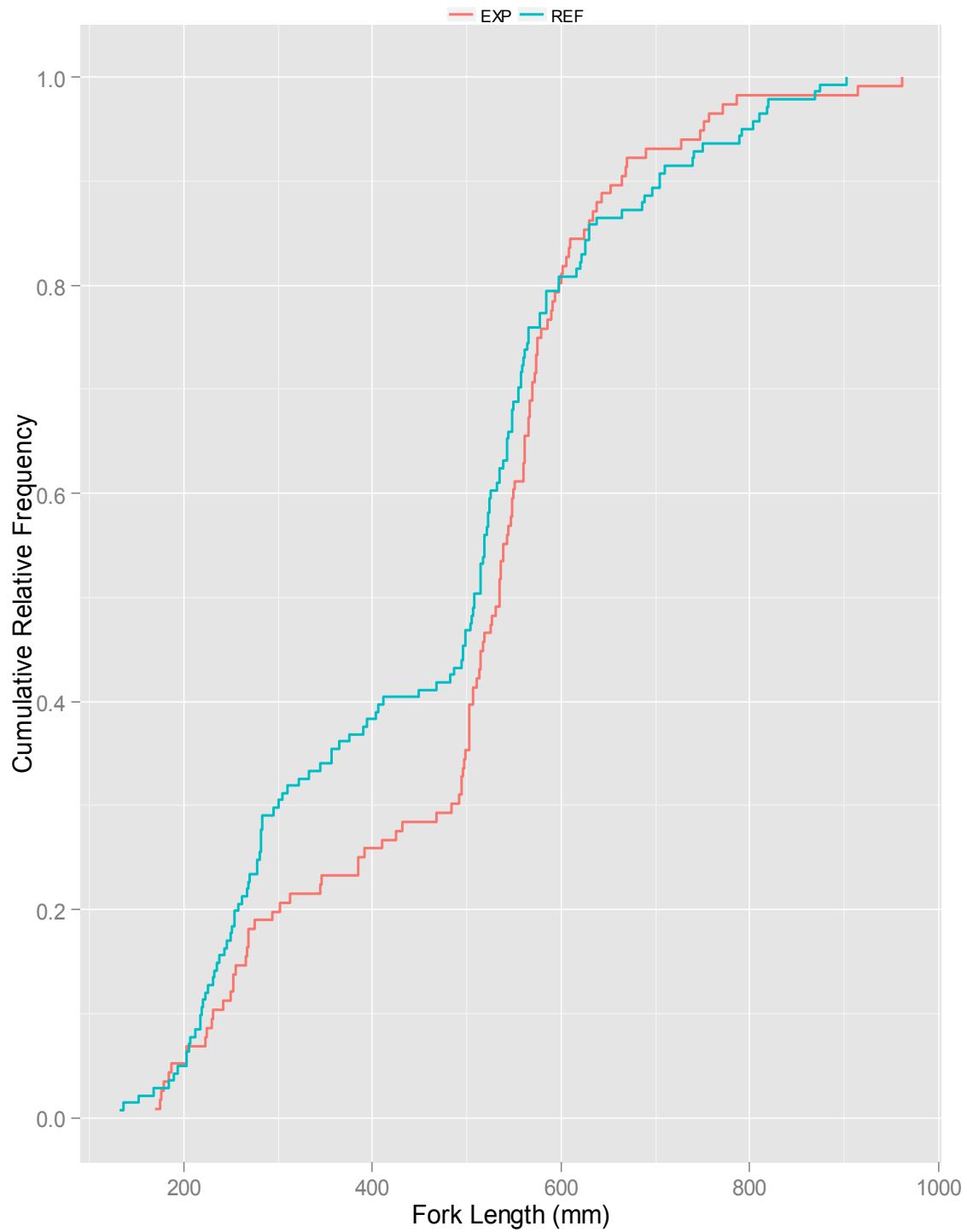


Figure 4.3- 16. Fork length (mm) density plots for pooled reference and exposure areas.

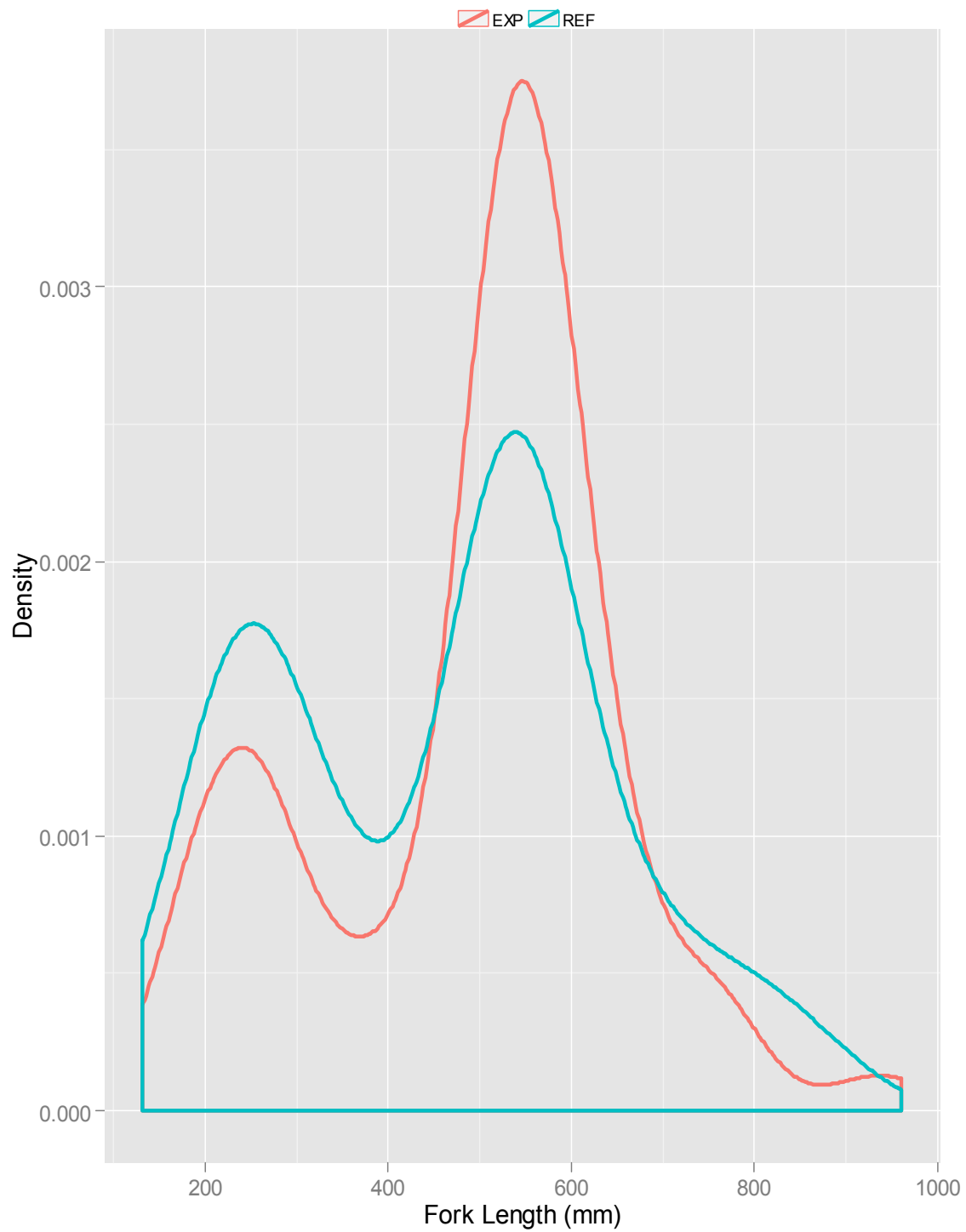


Figure 4.3- 17. Age (yr) empirical cumulative distribution function for pooled reference and exposure areas.

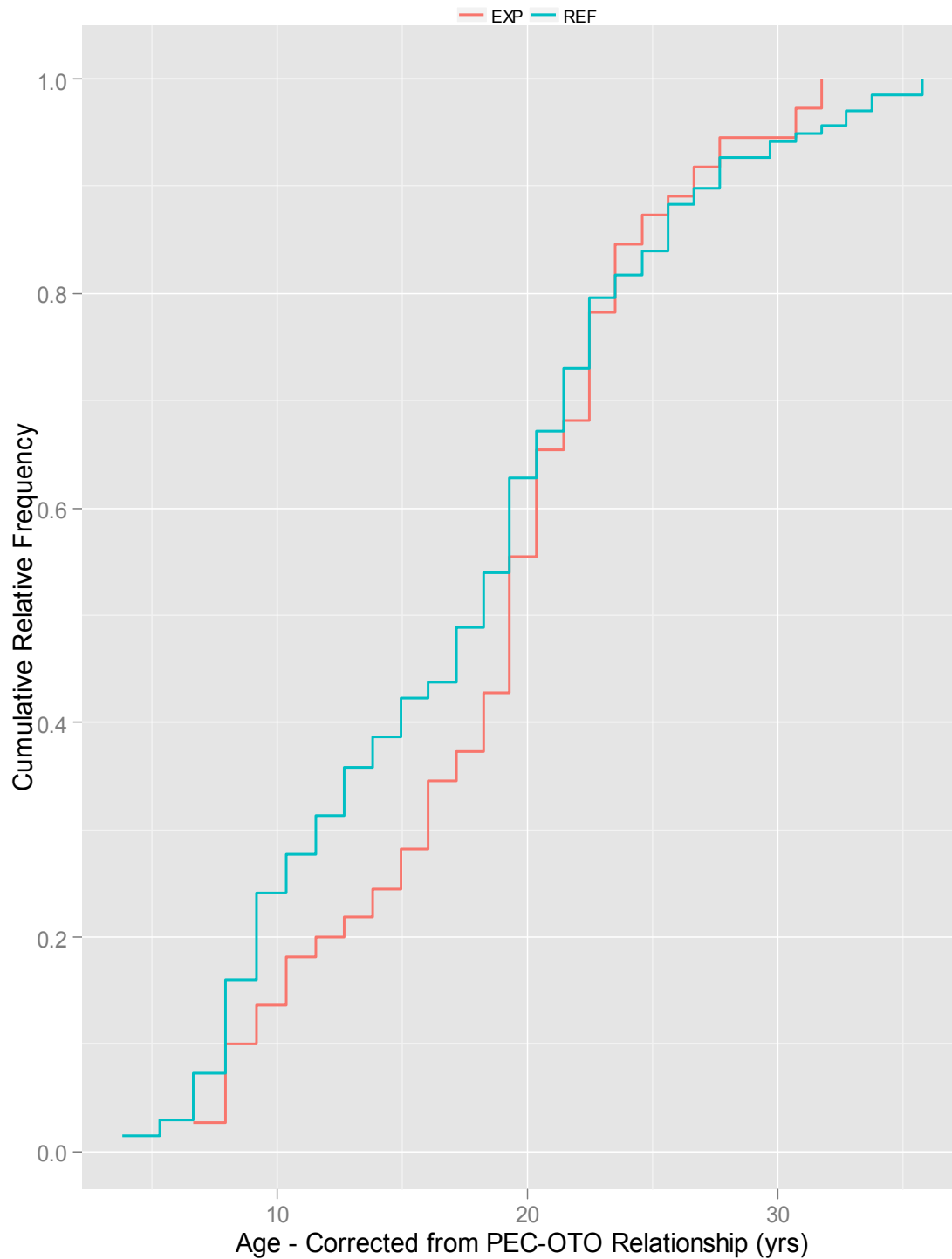


Figure 4.3- 18. Age (yr) density plots for pooled reference and exposure areas.

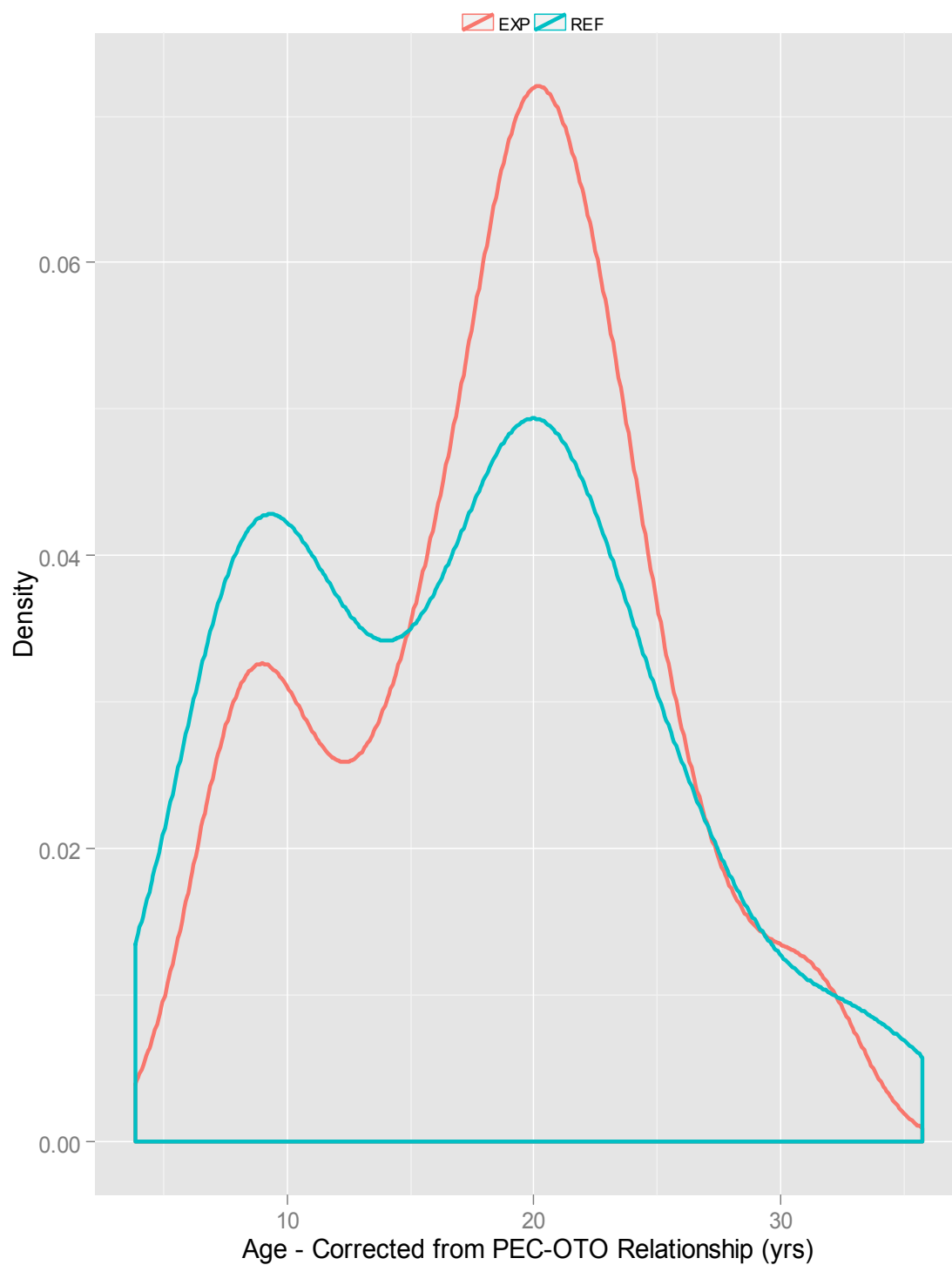


Figure 4.3- 19. Observed effect size and 95% confidence intervals for ANCOVA endpoints relative to EEM-recommended critical effect sizes (CES).

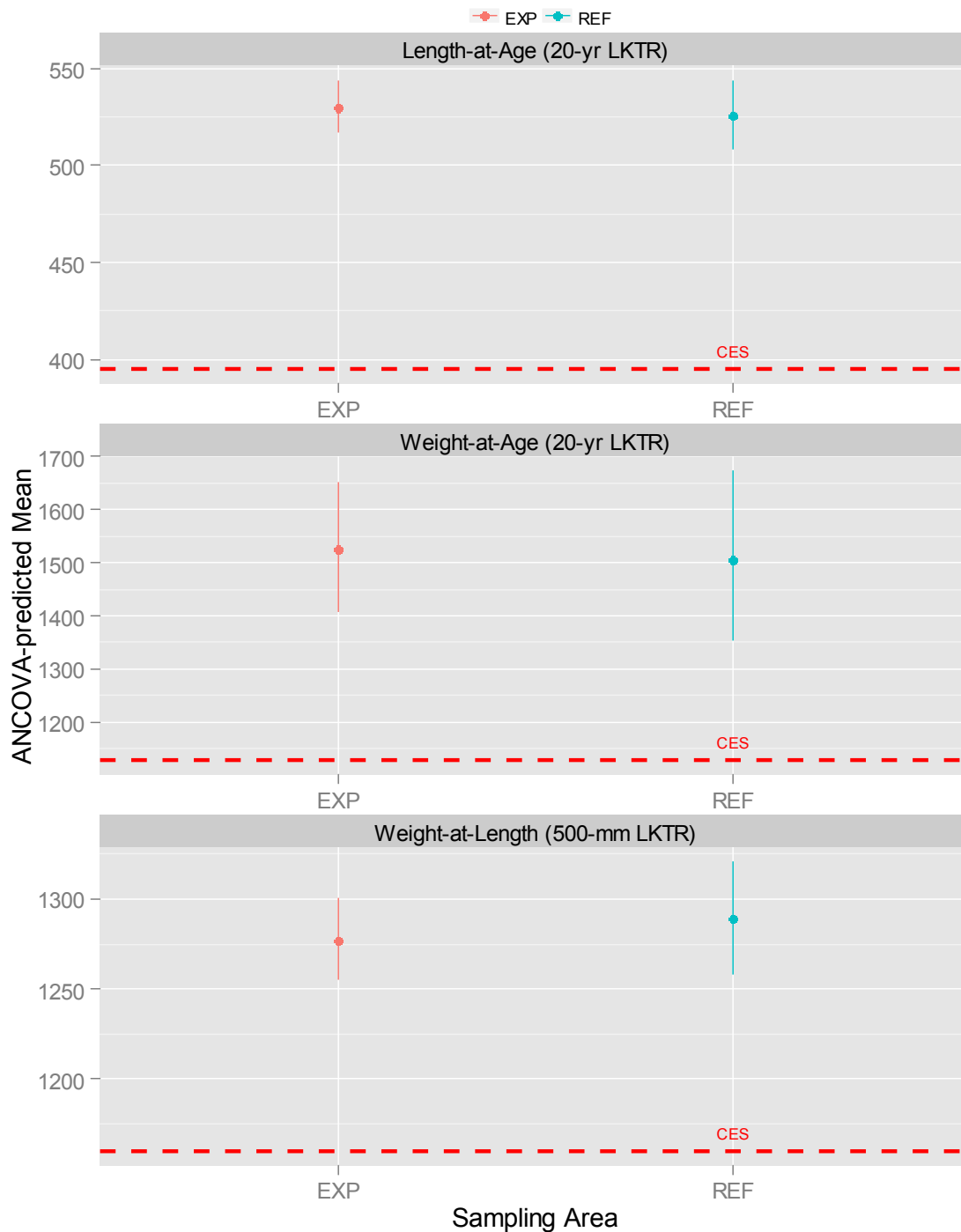


Figure 4.3- 20. Response ratio (i.e., exposure response/reference) empirical cumulative distribution function simulated for weight-at-age relative to the EEM-recommended critical effect size (CES) of 25% (i.e., response ratio of 0.75).

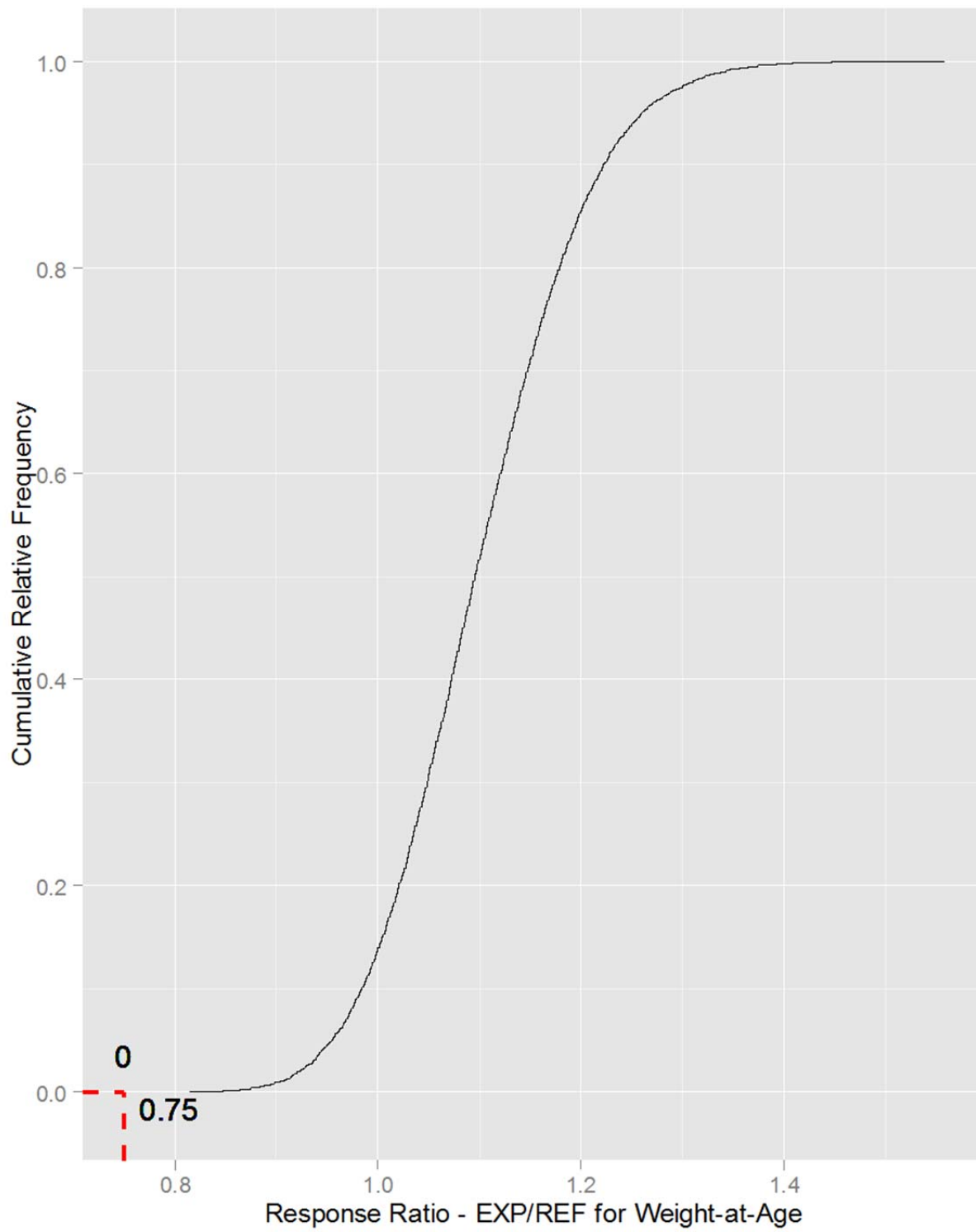


Figure 4.3- 21. Lake trout age (yr) empirical cumulative distribution function for pooled reference (with associated 95% confidence bands) and exposure areas.

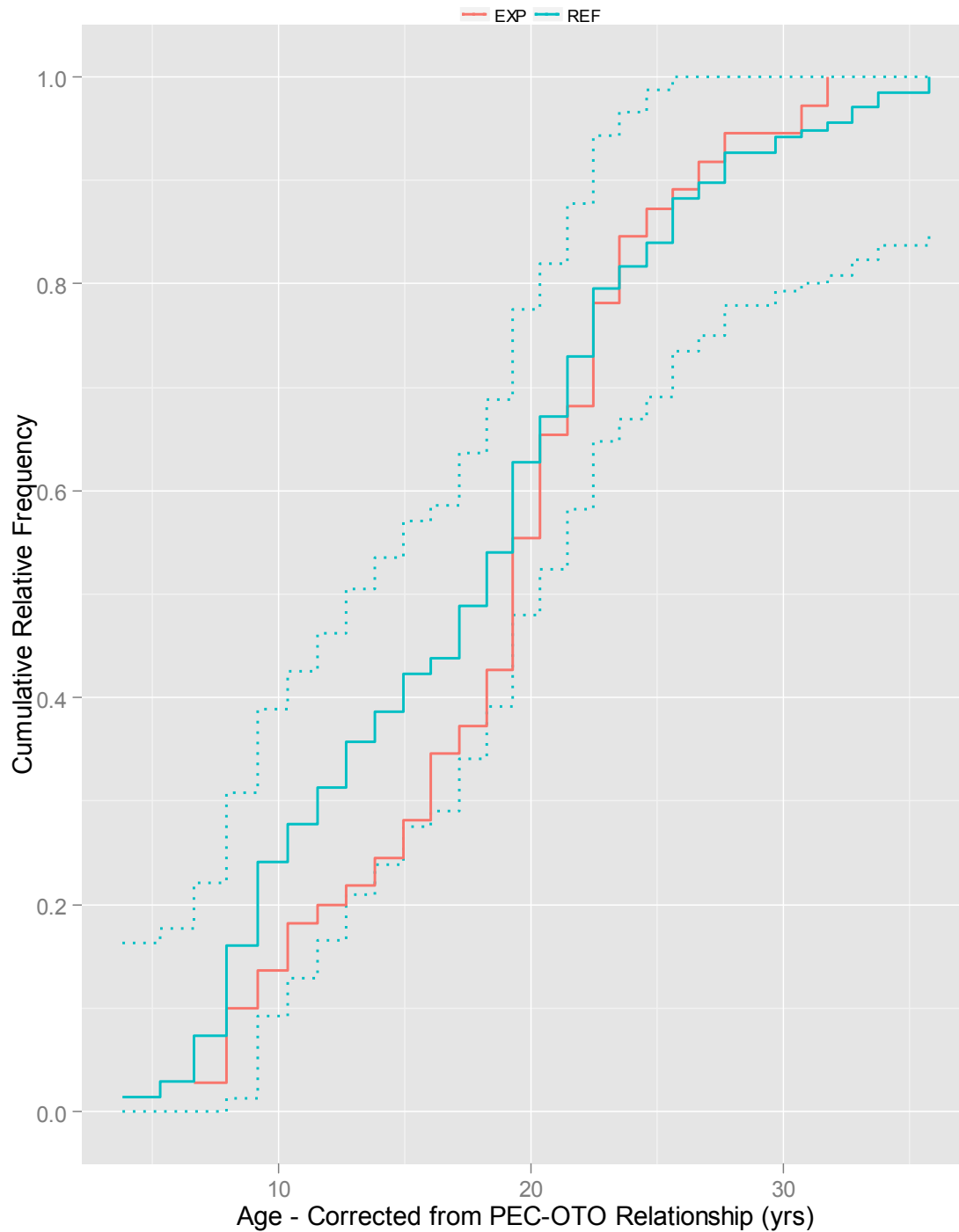


Figure 4.3- 22. Lake trout fork length (mm) empirical cumulative distribution function for pooled reference (with associated 95% confidence bands) and exposure areas.

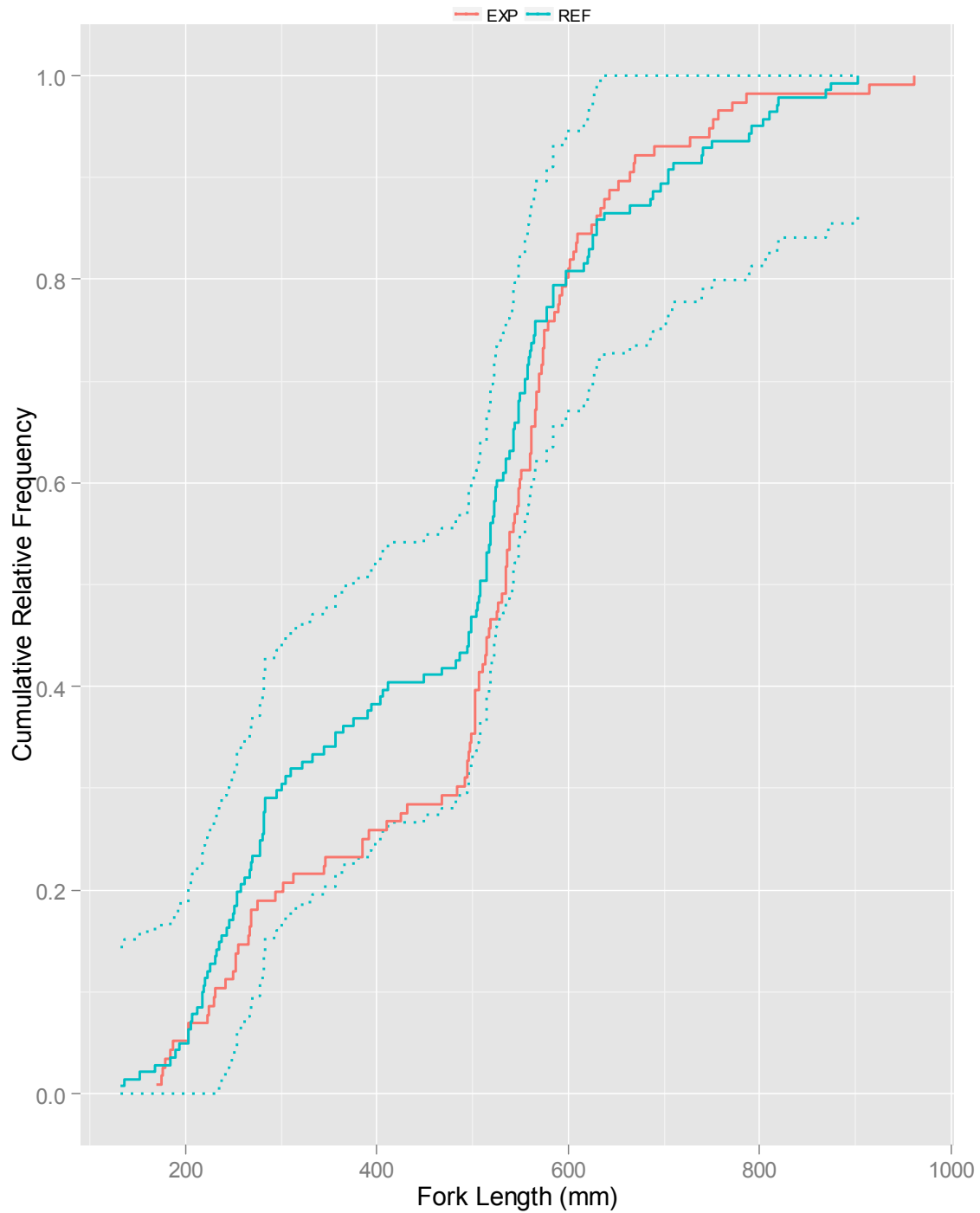


Figure 4.3- 23. Lake trout age (yr) empirical cumulative distribution function for each sampling area.

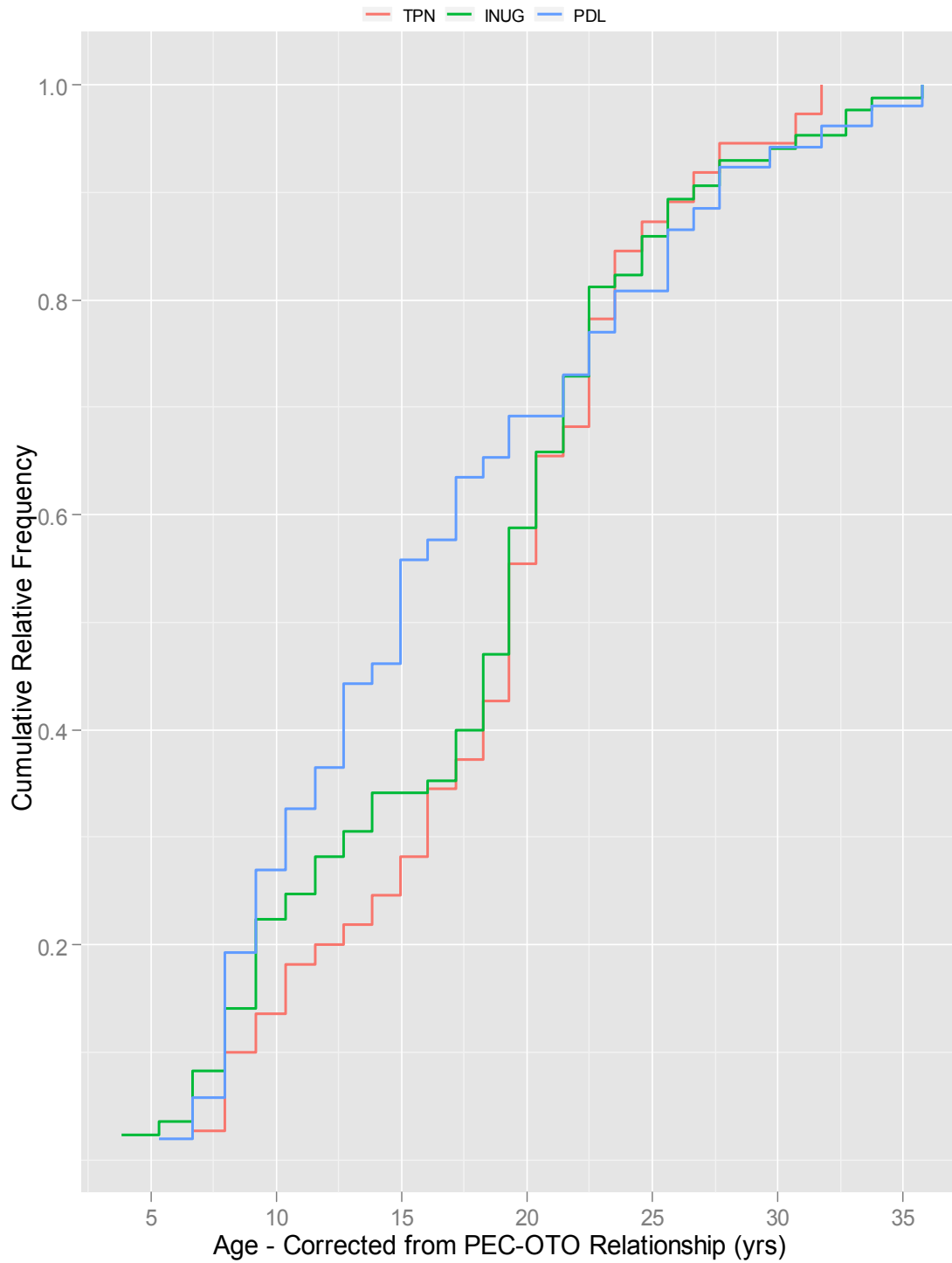


Figure 4.3- 24. Lake trout age (yr) density plots for each sampling area.

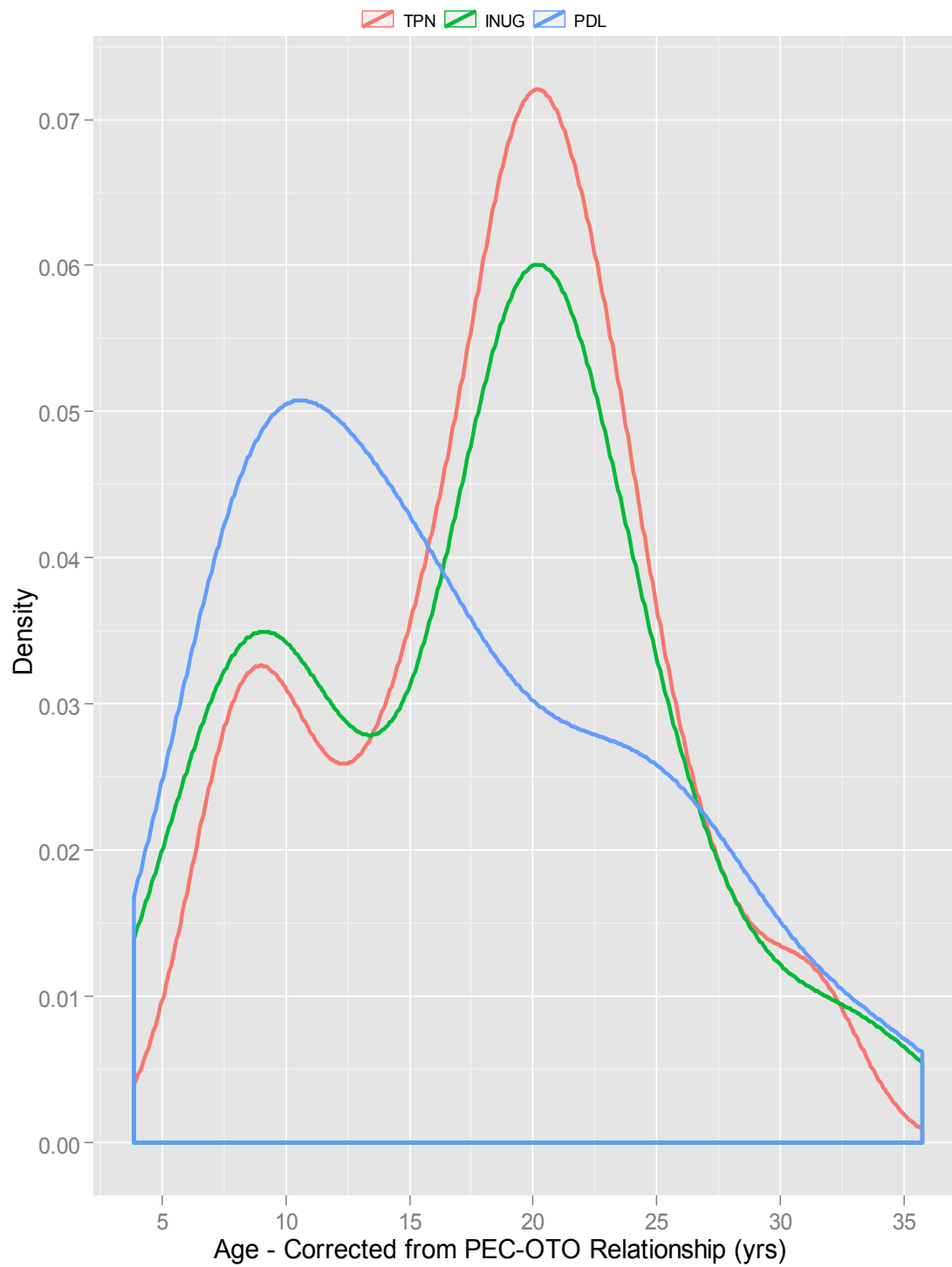


Figure 4.3- 25. Lake trout fork length (mm) empirical cumulative distribution function for each sampling area.

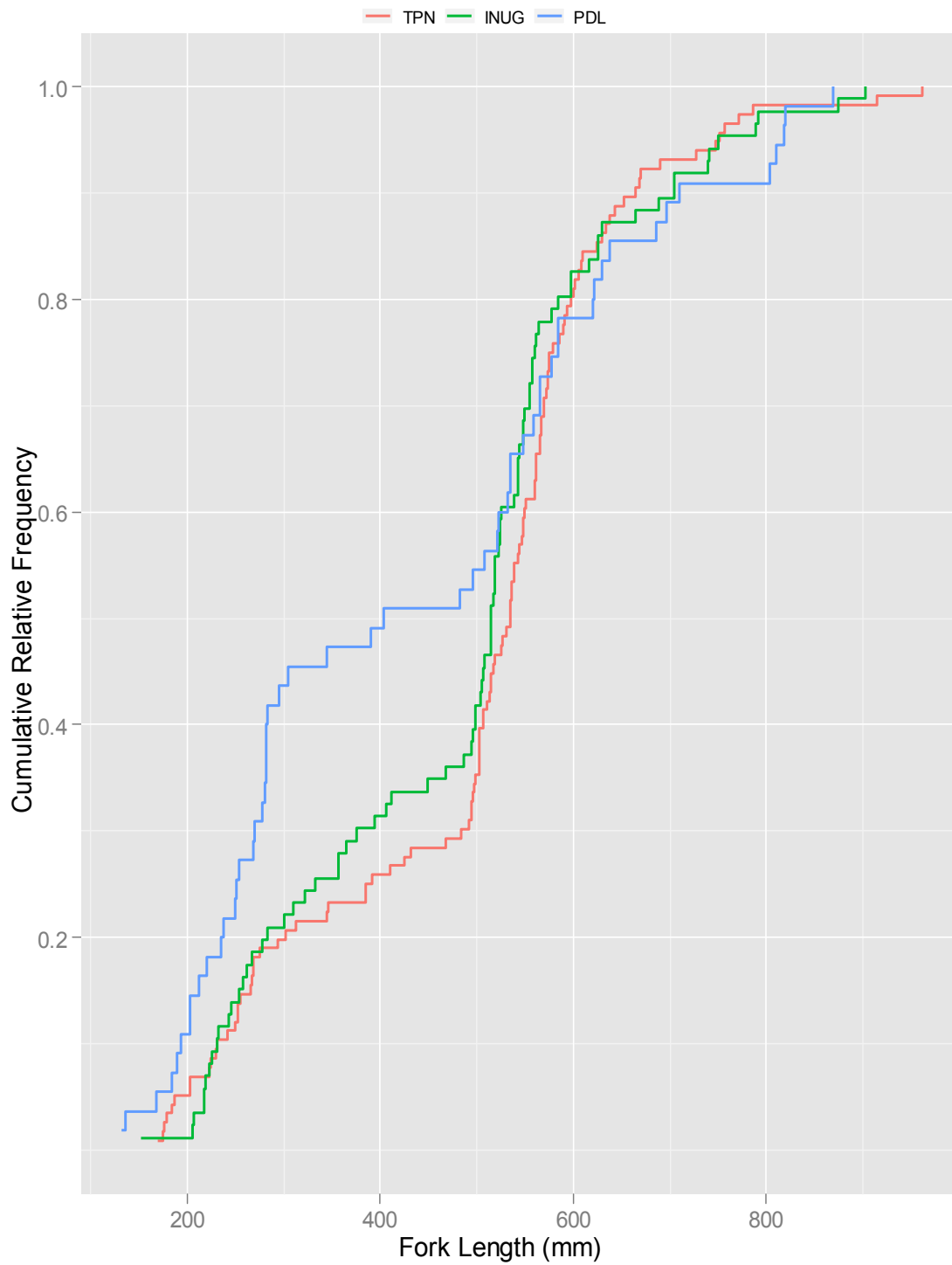
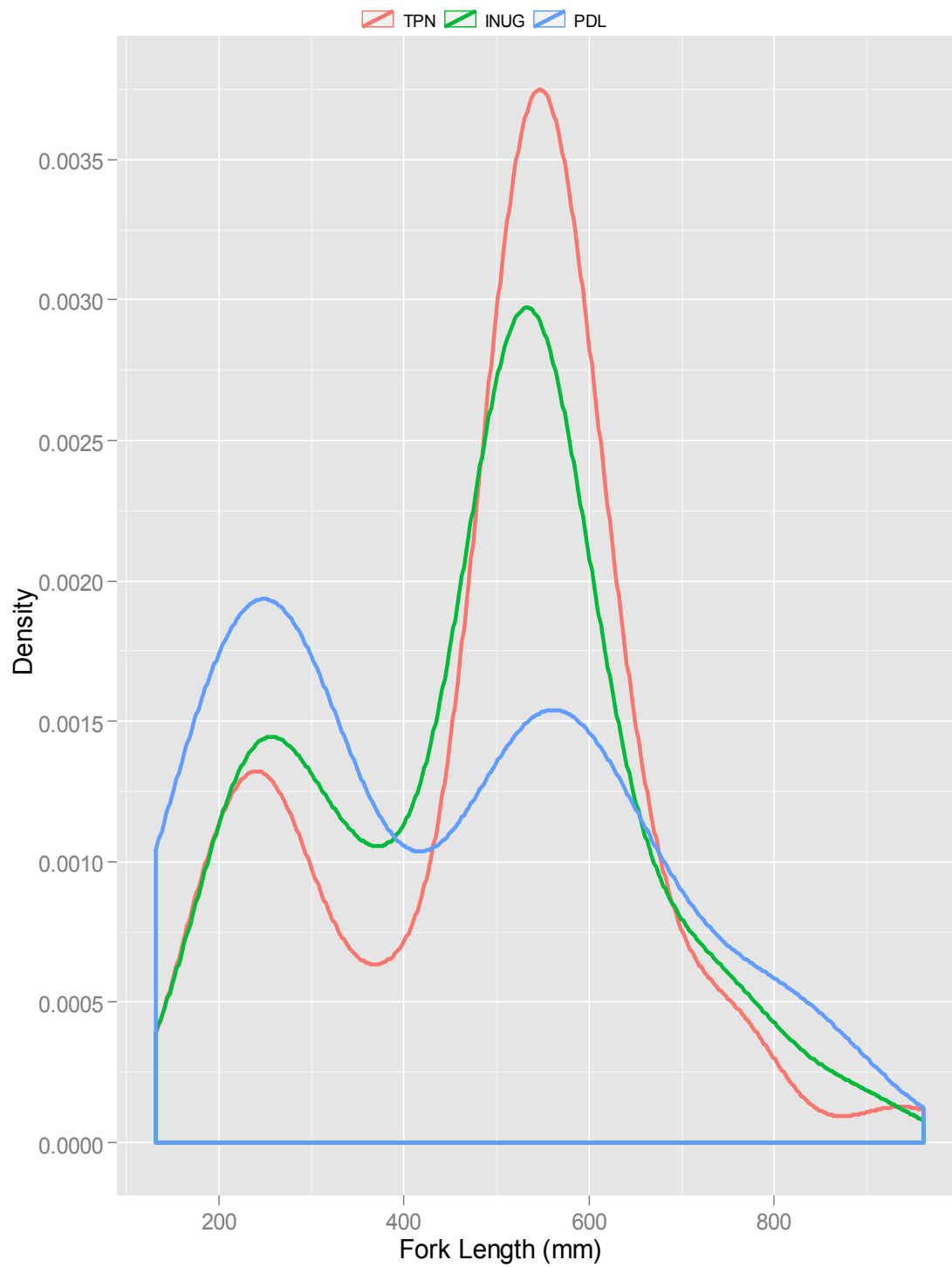


Figure 4.3- 26. Lake trout fork length (mm) density plots for each sampling area.



5. BENTHIC INVERTEBRATE COMMUNITY SURVEY

5.1. Sample Collection

Benthic invertebrates were collected from each of the three sampling areas (INUG, PDL, and TPN) from August 13–19th, 2011. Sampling locations (UTM coordinates), depth and dates are presented in **Table 5.1-1**. Sampling locations are illustrated in **Figure 3.3-7** (corresponding with sediment quality sampling), i.e., benthic invertebrate samples were collected at the same time and locations as sediment chemistry samples (see **Section 3.3.3**).

Benthic invertebrates were collected and analyzed according to the guidelines of the sediment and benthos sampling SOP (**Appendix D**) and according to the QA/QC procedures presented in the Study Design document (Azimuth, 2011). Generally, a Petite Ponar grab sampler (0.023 m²) was used to collect five replicate samples per area. Three independent grabs (subsamples) per replicate were sieved (500-µm) and composited to form a single sample. Only those grab samples that met the acceptability criteria outlined in the SOP were retained for analysis. Depths ranged from 6.5 to 9.2 m. Field data sheets from August 2011 are presented in **Appendix E** (sediment and benthos data are recorded on the same data sheets).

Table 5.1- 1. Benthic invertebrate sampling areas (UTM coordinates, NAD83).

Area Type ¹	Area-Replicate ²	Sampling Date	Depth (m)	Latitude (dd.mm.ss.s)	Longitude (dd.mm.ss.s)
REF	INUG-1	14-Aug-11	6.9	65 03 09.2	96 23 23.5
	INUG-2	14-Aug-11	6.8	65 03 09.7	96 23 25.9
	INUG-3	14-Aug-11	7.6	65 03 08.8	96 23 29.4
	INUG-4	14-Aug-11	8.5	65 03 07.3	96 23 31.7
	INUG-5	14-Aug-11	9.2	65 03 06.8	96 23 33.1
REF	PDL-1	13-Aug-11	6.5	65 06 16.8	96 13 11.6
	PDL-2	13-Aug-11	6.8	65 06 18.1	96 13 07.4
	PDL-3	13-Aug-11	6.9	65 06 09.4	96 13 00.4
	PDL-4	13-Aug-11	7.6	65 06 18.5	96 13 01.9
	PDL-5	13-Aug-11	6.7	65 06 17.5	96 13 08.3
EXP	TPN-1	19-Aug-11	7.0	65 02 08.1	96 06 11.2
	TPN-2	19-Aug-11	9.0	65 02 07.4	96 06 11.5
	TPN-3	19-Aug-11	9.2	65 02 06.9	96 06 09.1
	TPN-4	19-Aug-11	8.6	65 02 04.4	96 06 02.0
	TPN-5	19-Aug-11	8.4	65 02 03.3	96 05 55.0

Notes: ¹ Area types are as follows: EXP = exposure; REF = reference; ² Area IDs are as follows: TPN=Third Portage Lake - North; INUG=Inuggugayualik Lake; PDL=Pipedream Lake.

5.2. QA/QC

Standard procedures were used to collect biota samples. All sampling gear was thoroughly rinsed between sampling areas to ensure that there was no inadvertent introduction (i.e., cross-contamination) of biota from one area to another.

Field replicates (5 per area) were collected for benthos to determine natural variability and heterogeneity. Replicates were collected at least 20 m apart from one another, within the defined sampling areas, as described in **Section 5.1**.

Benthos samples were preserved in the field with a 10% buffered formalin solution and sent to Zaranko Environmental Assessment Services (ZEAS) (Nobleton, ON) for taxonomic identification and analysis. Benthic invertebrate raw data are presented in **Appendix K**.

Upon arrival at ZEAS, samples were immediately logged and inspected to ensure adequate preservation to a minimum level of 10% buffered formalin and checked for correct labeling. Benthos samples were sorted at a magnification of between 7 and 10 times with the use of a stereomicroscope. To expedite sorting prior to processing, all samples were stained with a protein dye that is absorbed by aquatic organisms but not by organic material, such as detritus and algae.

Prior to sorting, samples were washed free of formalin in a 500- μ m sieve. Benthos were enumerated and sorted into major taxonomic groups, (i.e., order and family), placed in glass bottles and re-preserved in 80% ethanol for more detailed taxonomic analysis by senior staff. Each bottle was labeled internally with the survey name, date, area and replicate number.

ZEAS incorporates the following set of QA/QC procedures in all benthic projects undertaken by the company to ensure the generation of high quality and reliable data:

- Samples were logged upon arrival, inspected, and enumerated;
- Samples were checked for proper preservation;
- Samples were stained to facilitate sorting;
- Taxonomic identifications were based on the most updated and widely used keys;
- 10% of the samples were re-sorted, and re-counted, targeting >90% recovery;
- Precision and accuracy estimates were calculated;
- A voucher collection was compiled;
- Sorted sediments and debris were re-preserved in 10% formalin and are retained for up to three months. For samples subject to subsampling, sorted and unsorted fractions were re-preserved separately.

Laboratory replicate counts were performed on 10% of all benthic samples. Replicate samples were chosen at random and processed at different times from the original analysis to reduce bias. Of the re-sorted samples, 0/21 and 1/14 of organisms were missed (**Table 5.2.1**). These results suggest that the vast majority of animals observed in benthic samples by the taxonomist were recovered.

Table 5.2- 1. Percent recovery of benthic invertebrate samples.

Area-Replicate	Number of Organisms Recovered	Number of Organisms in Re-sort	Percent Recovery
INUG-5-A	21	21	100.0%
PDL-4-A	13	14	92.9%
<i>Average % Recovery</i>			96.4%

Note: All fifteen samples were sorted in their entirety.

5.3. Data Assessment

5.3.1. Benthic Invertebrate Community Endpoints

The targeted benthic community endpoints (along with their abbreviated variable names in some tables and figures) were:

- *Total density (N.Tot.m2)* – number of invertebrates per m².
- *Evenness (Simp.E.prop)* – calculated using the method (based on Smith and Wilson, 1996) described in the TGD (Environment Canada, 2012).
- *Taxa richness (R.Tot)* – based on taxa counts identified to the lowest practical taxonomic level (i.e., lower than or equal to family level).
- *Similarity index (Bray.Cur)* – calculated using the method (based on Legendre and Legendre, 1983) described in the TGD (Environment Canada, 2012).

These endpoints are summarized in **Table 5.3-1** and **Figure 5.3-1**.

Summary tables for other supporting endpoints are presented in **Appendix L**; this includes:

- *Family density* – the number of individuals of each family expressed per unit area (#/m²). Values are reported for each family at each replicate station and as the mean and standard error of each family for the area.
- *Family proportion* – the percentage (%) density for each family at each replicate station and the mean and standard error of each family for the area.
- *Family presence/absence* – a matrix indicating the presence and absence of each family at each replicate station.
- *Raw family data* – a tabular listing of the number of individuals identified per family for each replicate station.
- *Raw lowest practical level data* – a tabular listing of the number of individuals identified at the lowest practical level for each replicate station.

5.3.2. Supporting Environmental Endpoints

The targeted supporting environmental endpoints (along with their abbreviated variable names in tables and figures) were:

- *Total organic carbon (TOC)* – in units of percent (%).
- *Sediment particle size (Fines)* – in units of percent (%); combination of silt and clay (fraction size <4 µm – 0.063 mm).
- *Depth (Depth.m)* – in units of meters (m).

These endpoints are summarized in **Table 5.3-2** and **Figure 5.3-2**.

5.3.3. Statistical Analysis

As described in **Section 2.2** and shown in **Figure 2.2-1**, a hierarchical approach was used to test for statistical differences among sampling areas. Initial analyses were conducted to test for differences between the pooled reference areas (i.e., INUG and PDL pooled as “REF” for reference) and TPN (as “EXP” for exposure); this is hereafter referred to as “C-I” (or “CI-based”). If statistical differences were identified, then analyses would test for differences among areas (i.e., testing to see if TPN is different from both INUG and PDL) (referred to hereafter as “area-based”).

The analyses described herein follow the statistical procedures recommended by the EEM technical guidance document (Environment Canada, 2012) to assess potential differences in the benthic invertebrate community between reference and exposure areas. Analysis of variance (ANOVA) is the standard method for testing for statistical differences among sampling areas (although ANCOVA can be used if there is a known covariate). All statistical analyses were conducted using R software (v 2.15).

Key assumptions of ANOVA are as follows (Environment Canada, 2012):

- Data for reference and exposure areas are normally distributed; and
- Variances are equal between reference and exposure area; and
- Errors are independently distributed.

The steps for CI-based ANOVA were as follows:

- 1) Response variables were plotted, highlighting C-I groups, to see basic relationships (e.g., see **Figures 5.3-1 and 5.3-2**).
- 2) Formal tests (Shapiro-Wilk normality test and Bartlett test of homogeneity of variance) and graphical methods (e.g., examination of residuals from ANOVA model conducted with various transformations) to determine the need for transformations.
- 3) Run ANOVA model testing for pooled C-I differences in the response endpoints. The model form in R was: Response ~ CI, where Response is a numeric response variable and CI is a factor with 2 levels (Control [pooled reference] and Impact [exposure]).
- 4) Examine final ANOVA model residuals graphically to check for underlying patterns that would violate model assumptions of normality and homogeneity of variance.
- 5) Effect size information calculated relative to reference area means.

This process was repeated for area-based ANOVA testing (i.e., the second tier of the hierarchical strategy discussed in **Section 2.2** to make inference for the MC-I design) to determine whether the TPN exposure area was different from both reference areas (i.e., independently, not pooled).

The CI-based ANOVA results are provided below by response endpoint.

Total Density (N.Tot.m2)

- N.Tot.m2 was not transformed for the ANOVA.
- Reference and exposure area means +/- standard error are shown in **Figure 5.3-3**.
- ANOVA results are shown in **Table 5.3-3**. The difference between pooled reference area and the TPN exposure area was not statistically significant.
- Observed effect size information is provided in **Table 5.3-4**. The exposure area had 39% lower total density than the pooled reference area.

Total Richness (R.Tot)

- R.Tot was not transformed for the ANOVA.
- Reference and exposure area means +/- standard error are shown in **Figure 5.3-3**.
- ANOVA results are shown in **Table 5.3-3**. The difference between pooled reference area and the TPN exposure area was statistically significant.
- Observed effect size information is provided in **Table 5.3-4**. The exposure area had 24% lower total richness than the pooled reference area.

Evenness (Simp.E.prop)

- Simp.E.prop was not transformed for the ANOVA.
- Reference and exposure area means +/- standard error are shown in **Figure 5.3-3**.
- ANOVA results are shown in **Table 5.3-3**. The difference between pooled reference area and the TPN exposure area was not statistically significant.
- Observed effect size information is provided in **Table 5.3-4**. The exposure area had 12% higher Simpson's Evenness than the pooled reference area.

Similarity (Bray.Cur)

- Bray.Cur was not transformed for the ANOVA.
- Reference and exposure area means +/- standard error are shown in **Figure 5.3-3**.
- ANOVA results are shown in **Table 5.3-3**. The difference between pooled reference area and the TPN exposure area was statistically significant.
- Observed effect size information is provided in **Table 5.3-4**. The exposure area had a 41% higher Bray Curtis Similarity Index than the pooled reference area.

Total organic carbon (TOC)

- TOC was not transformed for the ANOVA.
- Reference and exposure area means +/- standard error are shown in **Figure 5.3-4**.
- ANOVA results are shown in **Table 5.3-3**. The difference between pooled reference area and the TPN exposure area was statistically significant.

- Observed effect size information is provided in **Table 5.3-4**. The exposure area had 50% lower TOC than the pooled reference area.

Fines (Fines)

- Fines was not transformed for the ANOVA.
- Reference and exposure area means +/- standard error are shown in **Figure 5.3-4**.
- ANOVA results are shown in **Table 5.3-3**. The difference between pooled reference area and the TPN exposure area was statistically significant.
- Observed effect size information is provided in **Table 5.3-4**. The exposure area had 29% lower fine sediments than the pooled reference area.

Depth (Depth.m)

- Depth.m was not transformed for the ANOVA.
- Reference and exposure area means +/- standard error are shown in **Figure 5.3-4**.
- ANOVA results are shown in **Table 5.3-3**. The difference between pooled reference area and the TPN exposure area was statistically significant.
- Observed effect size information is provided in **Table 5.3-4**. The exposure area was 15% deeper than the pooled reference area.

The area-based ANOVA results are provided below by response endpoint (note that only those endpoints showing significant differences in CI-based analyses were included in area-based analyses). ANOVAs were first conducted to test for among-area differences (all area-based ANOVA results shown in **Table 5.3-5**) then to test whether there was a difference between the TPN exposure area and the particular reference area (i.e., INUG or PDL) with the nearest mean (only the ultimate results [effect or not] are reported for the area-based ANOVAs in **Table 5.3-6**, along with effect size information).

Total Density (N.Tot.m2)

- No differences in CI-based analyses, so not included in area-based assessment.

Total Richness (R.Tot)

- R.Tot was not transformed for the ANOVA.
- Area means and raw data are shown in **Figure 5.3-1**.
- Significant differences were not detected among areas (**Table 5.3-5**), nor was there a significant difference between TPN and PDL.
- Observed effect size information is provided in **Table 5.3-6**. The TPN exposure area had 20% lower total richness than the PDL reference area.

Evenness (Simp.E.prop)

- No differences in CI-based analyses, so not included in area-based assessment.

Similarity (Bray.Cur)

- Bray.Cur was not transformed for the ANOVA.
- Area means and raw data are shown in **Figure 5.3-1**.
- Significant differences were detected among areas (**Table 5.3-5**), and there was a significant difference between TPN and PDL.
- Observed effect size information is provided in **Table 5.3-6**. The exposure area had a 38% higher Bray Curtis Similarity Index than the pooled reference area.

Total organic carbon (TOC)

- TOC was not transformed for the ANOVA.
- Area means and raw data are shown in **Figure 5.3-2**.
- Significant differences were detected among areas (**Table 5.3-5**), but there was no significant difference between TPN and PDL.
- Observed effect size information is provided in **Table 5.3-6**. The exposure area had 37% lower TOC than the PDL area.

Fines (Fines)

- Fines were not transformed for the ANOVA.
- Area means and raw data are shown in **Figure 5.3-2**.
- Significant differences were detected among areas (**Table 5.3-5**), and there was a significant difference between TPN and PDL.
- Observed effect size information is provided in **Table 5.3-6**. The exposure area had 27% lower fine sediments than the PDL reference area.

Depth (Depth.m)

- Depth.m was not transformed for the ANOVA.
- Area means and raw data are shown in **Figure 5.3-2**.
- Significant differences were detected among areas (**Table 5.3-5**), but there was no significant difference between TPN and INUG.
- Observed effect size information is provided in **Table 5.3-6**. The TPN exposure area was 8% deeper than the INUG reference area.

Based on the preceding results (i.e., differences detected between reference and exposure areas for both benthic invertebrate community endpoints and supporting environmental variables), a

series of supplemental analyses were conducted (**Section 5.4**) to assess causality (i.e., if the differences are attributable to effluent exposure or to natural causes).

5.3.4. Post-hoc Assessment of Power and Confidence

This section examines results of CI-based (i.e., pooled reference area) statistical analyses with the intent of providing insights into the degree of confidence of their conclusions. This essentially focuses on understanding the degree of uncertainty (e.g., using 95% confidence intervals) for observed effect size estimates in the context of the EEM-recommended critical effect size (CES). *Post-hoc*¹² power analyses can also provide insights for planning subsequent EEM cycles.

This section focuses on the benthic invertebrate community endpoints only; further exploration of the role of the supplemental environment endpoints is conducted in **Section 5.4** (i.e., where a much larger CREMP data set can be used to better characterize the influence of supporting environmental variables on the differences observed in the EEM data). Consequently, while caution should be used regarding making inferences as to the cause to the observed differences between pooled reference and exposure areas (i.e., as the results described in **Section 5.3.3** show differences in both the benthic invertebrate community endpoints and supplemental environmental endpoints, the nature and cause of the biological differences will be explored in greater detail in **Section 5.4**), the ability of the study design to detect such effects relative to the CES is still relevant for planning future studies.

The following approaches were taken using the results of the CI-based ANOVA analyses:

- 1) Comparisons of observed effect sizes and their 95% confidence intervals to the EEM-recommended critical effect sizes (CES) to determine the confidence that the observed effects do not exceed the CES. This was conducted by:
 - a. Randomly generate 500,000 new effect size estimates using the actual effect size and its standard error.
 - b. Compare each of the generated effect sizes to the CES and record the proportion of those not exceeding the CES.
- 2) Estimation of statistical power of detecting the CES based on the executed study design. This was conducted using the two methods described in the textbox.
- 3) Estimation of the minimum detectable effect size for the targeted statistical power of 0.9 (i.e., $\beta = 0.1$) based on the executed study design. This was conducted using the two methods described in the textbox.

¹² Note that *post-hoc* power analysis is often associated with the controversial practice of estimating the power of a study using the sample size and observed effect size. While the EEM TGD uses the term “*post-hoc*”, the nature of the power analyses would actually classify them as *a priori* since they are being used to inform subsequent EEM cycles.

- 4) Estimation of minimum sample size required to detect the CES based on the targeted statistical power. This was conducted using the two methods described in the textbox.

Key results for the benthic invertebrate community endpoints were as follows:

- *Observed Effect Sizes* – **Figure 5.3-5** shows the mean and standard errors of each benthic invertebrate community endpoint relative to the EEM-recommended CES values. The simulated effect size distribution and EEM CES are shown in **Figure 5.3-6**, along with the confidence estimates regarding exceeding the CES. Confidence was high (i.e., > 0.88) that the observed effects sizes were below the CES for all endpoints except for the Similarity Index (Bray.Cur), for which the mean value exceeded the CES (confidence was 0.44 that the CES was not exceeded).
- *Power to Detect CES* – an R function (see textbox) was used to estimate statistical power. Power to detect the CES was fairly high for all three endpoints (**Table 5.3-7**), ranging from near 0.8 to nearly 1.
- *Minimum Effect Size for Target Power* – an R function (see textbox) was used to estimate minimum effect size. Minimum detectable effect sizes were higher than observed effect sizes for all benthic invertebrate endpoints (**Table 5.3-7**).
- *Minimum Sample Size for CES with Target Power* – an R function (see textbox) was used to estimate minimum sample size. The minimum sample sizes were higher than actual sample sizes for Evenness and Similarity (**Table 5.3-7**).

Power Analyses for ANOVA

Method 1 – The function “pwr.t2n.test” in the R “pwr” package, which estimates power for the t-test for unequal sample sizes, was used (n.b., t-test produces the same result as ANOVA when testing two groups [C-I]). This procedure solves for whichever of the following are missing: effect size ($d = [EXP-REF]/SD$, where $SD = \sqrt{ANOVA \text{ Mean Square Error}}$), n for EXP, n for REF, α , β . This allows all the key power questions to be addressed.

Method 2 – Simulated data sets based on the actual properties of the data (e.g., ref mean, SD, n) were used to verify that Method 1 was producing accurate results.

As discussed earlier, these results do not take potential confounding relationships with supporting environmental variables into consideration. Consequently, the above results should be viewed with caution.

Table 5.3- 1. Summary statistics for benthic invertebrate community endpoints by sampling area, EEM Data.

Primary Benthic Endpoints							Supporting Benthic Endpoints						
Primary Benthic Endpoints				Supporting Benthic Endpoints			Primary Benthic Endpoints				Supporting Benthic Endpoints		
Total Density (#/m ²)	Taxa Richness (# taxa)	Simpson's Evenness Index	Bray-Curtis Similarity Index	Simpson's Diversity Index	Family Richness (# families)		Total Density (#/m ²)	Taxa Richness (# taxa)	Simpson's Evenness Index	Bray-Curtis Similarity Index	Simpson's Diversity Index	Family Richness (# families)	
INUG							INUG						
1	319	9	0.53	0.59	0.79	4	n	5	5	5	5	5	5
2	783	9	0.46	0.34	0.76	4	Mean	928	12.6	0.40	0.40	0.79	4.2
3	971	12	0.39	0.34	0.79	4	Median	783	12	0.41	0.34	0.79	4.0
4	1783	18	0.23	0.41	0.76	5	StDev	535	3.9	0.11	0.11	0.03	0.4
5	783	15	0.41	0.34	0.84	4	SE	239	1.7	0.05	0.05	0.01	0.2
							Min	319	9	0.23	0.34	0.76	4.0
							Max	1783	18	0.53	0.59	0.84	5.0
PDL							PDL						
1	739	12	0.42	0.43	0.80	4	n	5	5	5	5	5	5
2	899	12	0.40	0.41	0.79	5	Mean	881	11.2	0.40	0.42	0.77	4.8
3	1203	10	0.35	0.48	0.71	6	Median	899	12	0.41	0.43	0.79	5.0
4	638	8	0.41	0.35	0.70	3	StDev	215	2.3	0.03	0.05	0.06	1.3
5	928	14	0.44	0.43	0.84	6	SE	96	1.0	0.02	0.02	0.03	0.6
							Min	638	8	0.35	0.35	0.70	3.0
							Max	1203	14	0.44	0.48	0.84	6.0
TPN							TPN						
1	899	10	0.32	0.42	0.69	5	n	5	5	5	5	5	5
2	855	10	0.31	0.45	0.68	4	Mean	554	9.0	0.45	0.58	0.73	3.6
3	261	6	0.64	0.75	0.74	3	Median	536	10	0.40	0.51	0.74	3.0
4	536	11	0.40	0.51	0.77	3	StDev	320	2.0	0.16	0.17	0.05	0.9
5	217	8	0.60	0.77	0.79	3	SE	143	0.9	0.07	0.07	0.02	0.4
							Min	217	6	0.31	0.42	0.68	3.0
							Max	899	11	0.64	0.77	0.79	5.0

Table 5.3- 2. Summary statistics for benthic invertebrate community supporting environmental variables endpoints by sampling area, EEM Data.

INUG				INUG			
	Fines (%)	TOC (%)	Depth (m)		Fines (%)	TOC (%)	Depth (m)
1	88.5	2.93	6.9	n	5	5	5
2	86.4	2.99	6.8	Mean	92.9	3.80	7.8
3	95.0	4.84	7.6	Median	95.0	4.01	7.6
4	97.3	4.01	8.5	StDev	5.1	0.82	1.0
5	97.3	4.22	9.2	SE	2.3	0.37	0.5
				Min	86.4	2.93	6.8
				Max	97.3	4.84	9.2
PDL				PDL			
1	86.3	2.44	6.5	n	5	5	5
2	85.1	2.64	6.8	Mean	87.4	2.54	6.9
3	84.2	1.77	6.9	Median	86.3	2.64	6.8
4	94.2	2.76	7.6	StDev	4.0	0.49	0.4
5	87.5	3.09	6.7	SE	1.8	0.22	0.2
				Min	84.2	1.77	6.5
				Max	94.2	3.09	7.6
TPN				TPN			
1	53.4	1.80	7.0	n	5	5	5
2	85.5	3.71	9.0	Mean	64.0	1.59	8.4
3	53.2	1.25	9.2	Median	57.7	1.25	8.6
4	57.7	0.66	8.6	StDev	13.9	1.29	0.9
5	70.3	0.51	8.4	SE	6.2	0.58	0.4
				Min	53.2	0.51	7.0
				Max	85.5	3.71	9.2

Table 5.3- 3. Summary of CI-based ANOVA tests for benthic invertebrate community and supporting environmental endpoints, EEM Data.

Endpoint	Source of Variation	Sum of Squares	df	Mean Square	F	p-value	Significant at p<0.1?
<i>Benthic Community Endpoints</i>							
Total Density	Between Groups (C-I)	410029	1	410029	3.054	0.104	No
N.Tot.m2	Within Groups (Residual:	1745266	13	134251			
	Total	2155295	14				
Total Richness (R.Tot)	Between Groups (C-I)	28.033	1	28.0333	3.542	0.082	Yes
	Within Groups (Residual:	102.9	13	7.9154			
	Total	130.93	14				
Evenness (Simp.E.prop)	Between Groups (C-I)	0.008	1	0.008	0.715	0.413	No
	Within Groups (Residual:	0.152	13	0.012			
	Total	0.160	14				
Similarity (Bray.Cur)	Between Groups (C-I)	0.094	1	0.094	7.247	0.018	Yes
	Within Groups (Residual:	0.169	13	0.013			
	Total	0.263	14				
<i>Supporting Sediment Endpoints</i>							
TOC	Between Groups (C-I)	8.353	1	8.353	7.583	0.016	Yes
	Within Groups (Residual:	14.321	13	1.102			
	Total	22.674	14				
Fines	Between Groups (C-I)	2277.3	1	2277.3	29.272	<0.001	Yes
	Within Groups (Residual:	1011.4	13	77.8			
	Total	3288.7	14				
Depth	Between Groups (C-I)	3.960	1	3.960	5.140	0.041	Yes
	Within Groups (Residual:	10.017	13	0.771			
	Total	13.977	14				

Notes: "df" = degrees of freedom; "F" = statistical F ratio; "C-I" = control-impact (or REF-EXP).

Table 5.3- 4. Summary of CI-based effect size information for benthic invertebrate community and supporting environmental endpoints, EEM Data.

Endpoint	Area	Mean	Ref SD	Obs. ES	% ES	ES/Ref SD	Effect?
<i>Benthic Community Endpoints</i>							
Total Density	Reference	905	385				
N.Tot.m2	Exposure	554		-351	-39%	-0.9	No
Total Richness	Reference	11.9	3.107				
(R.Tot)	Exposure	9		-2.9	-24%	-0.9	Yes
Evenness	Reference	0.404	0.077				
(Simp.E.prop)	Exposure	0.454		0.05	12%	0.6	No
Similarity	Reference	0.412	0.079				
(Bray.Cur)	Exposure	0.580		0.17	41%	2.1	Yes
<i>Supporting Sediment Endpoints</i>							
TOC	Reference	3.2	0.9				
	Exposure	1.6		-1.6	-50%	-1.7	Yes
Fines	Reference	90.2	5.2				
	Exposure	64.0		-26.1	-29%	-5.0	Yes
Depth	Reference	7.4	0.9				
	Exposure	8.4		1.1	15%	1.2	Yes

Notes: "Ref SD" = pooled REF standard deviation; "Obs. ES" = observed effect size; "% ES" = 100*(EXP Mean - REF mean)/REF mean.

Table 5.3- 5. Summary of Area-based ANOVA tests for benthic invertebrate community and supporting environmental endpoints, EEM Data.

Endpoint	Source of Variation	Sum of Squares	df	Mean Square	F	p-value	Effect?
Benthic Community Endpoints							
Total Richness (R.Tot)	Between Groups (Areas)	32.9	2	16.5	2.016	0.176	No
	Within Groups (Residuals)	98.0	12	8.2			
	Total	130.90	14				
Similarity (Bray.Cur)	Between Groups (Areas)	0.095	2	0.047	3.380	0.068	Yes
	Within Groups (Residuals)	0.168	12	0.014			
	Total	0.263	14				
Supporting Sediment Endpoints							
TOC	Between Groups (Areas)	12.309	2	6.155	7.126	0.009	Yes
	Within Groups (Residuals)	10.364	12	0.864			
	Total	22.673	14				
Fines	Between Groups (Areas)	2351.3	2	1175.65	15.050	<0.001	Yes
	Within Groups (Residuals)	937.39	12	78.12			
	Total	3288.7	14				
Depth	Between Groups (Areas)	5.985	2	2.993	4.494	0.035	Yes
	Within Groups (Residuals)	7.992	12	0.666			
	Total	13.977	14				

Notes: "df" = degrees of freedom; "F" = statistical F ratio; "C-I" = control-impact (or REF-EXP); "Effect" = difference among areas ($p < 0.1$).

Table 5.3- 6. Summary of Area-based effect size information for benthic invertebrate community and supporting environmental endpoints, EEM Data.

Endpoint	Area	Mean	Ref SD	Obs. ES	% ES	ES/Ref SD	Effect?
<i>Benthic Community Endpoints</i>							
Total Richness (R.Tot)	PDL	11.2	2.28				
	TPN	9		-2.2	-20%	-1.0	No
Similarity (Bray.Cur)	PDL	0.420	0.047				
	TPN	0.580		0.16	38%	3.4	Yes
<i>Supporting Sediment Endpoints</i>							
TOC	PDL	2.5	0.5				
	TPN	1.6		-0.9	-37%	-1.9	No
Fines	PDL	87.4	4.0				
	TPN	64.0		-23.4	-27%	-5.9	Yes
Depth	INUG	7.8	1.0				
	TPN	8.4		0.6	8%	0.6	No

Notes: "Ref SD" = standard deviation of Ref area tested; "Obs. ES" = observed effect size; "% ES" = 100*(EXP Mean - REF mean)/REF mean; "Effect" = difference between TPN and closest REF area (p<0.1).

Table 5.3- 7. Power analysis information and results for benthic invertebrate community endpoints tested using ANOVA.

Endpoint	Obs. ES	Ref SD	CES	n (EXP/REF)	α	β	Confidence Obs. ES<CES	Power for CES	Minimum ES	Minimum EXP n for CES and Power
<i>Benthic Community Endpoints</i>										
Total Density N.Tot.m2	-351	385	-770	5/10	0.1	0.1	0.88	0.98	620	4
Total Richness (R.Tot)	-2.9	3.1073	-6.2146	5/10	0.1	0.1	0.98	0.98	4.77	4
Evenness (Simp.E.prop)	0.05	0.0775	0.155	5/10	0.1	0.1	0.96	0.80	0.183	7
Similarity (Bray.Cur)	0.17	0.0791	0.1582	5/10	0.1	0.1	0.44	0.78	0.193	7

Notes: "Obs. ES" = observed effect size; "CES" = critical effect size (from EEM TGD) = 2 * Ref SD (note that it is shown here with the same change direction as the Obs. ES); "n" = sample size; "Ref SD" = standard deviation of pooled reference area; "Minimum ES" reported in Obs. ES units, not as multiple of Ref SD; Minimum sample size (REF n = 2 * EXP n).

Figure 5.3- 1. Benthic invertebrate community endpoint results by sampling area, EEM Data.

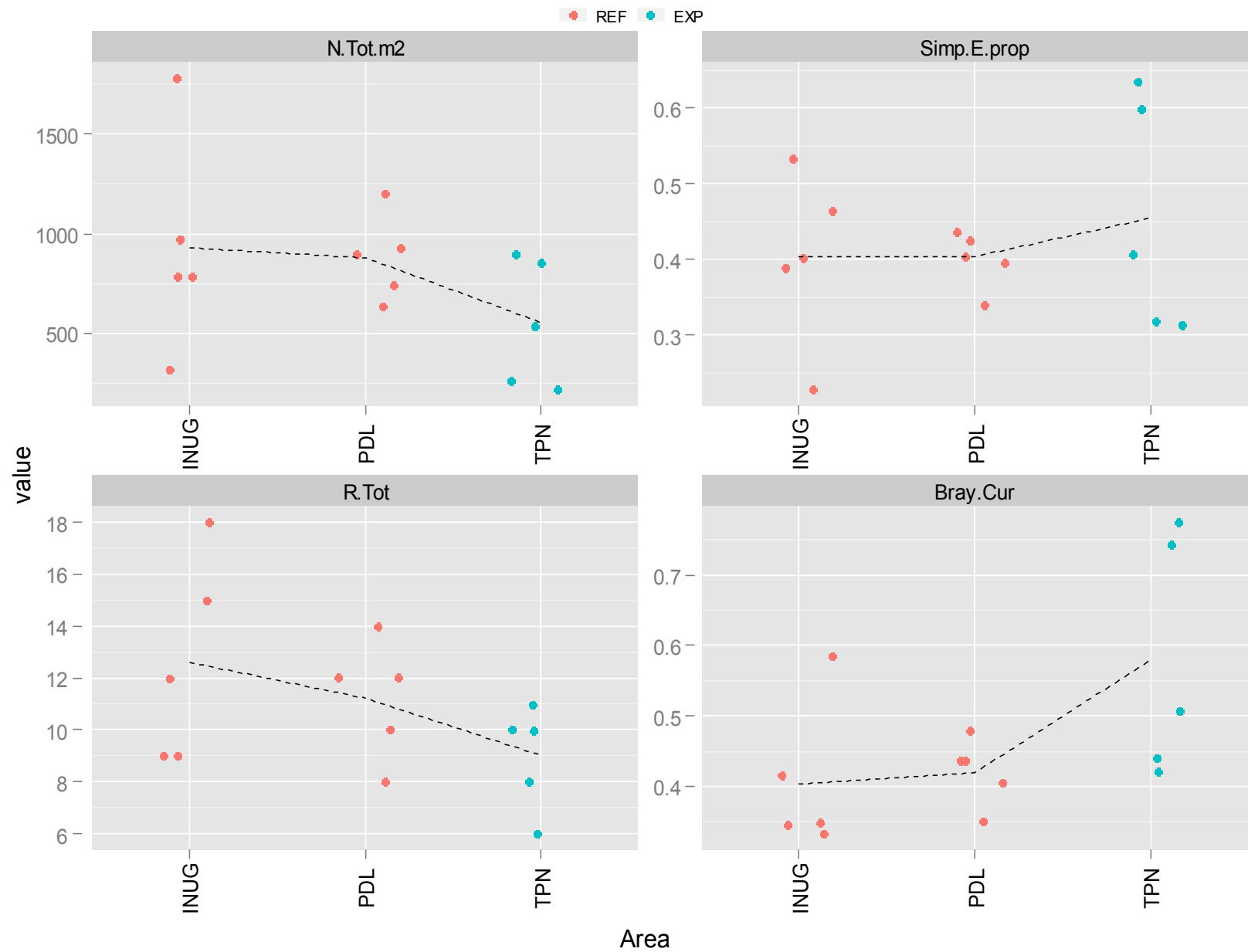


Figure 5.3- 2. Benthic invertebrate community supporting environmental endpoint results by sampling area, EEM Data.

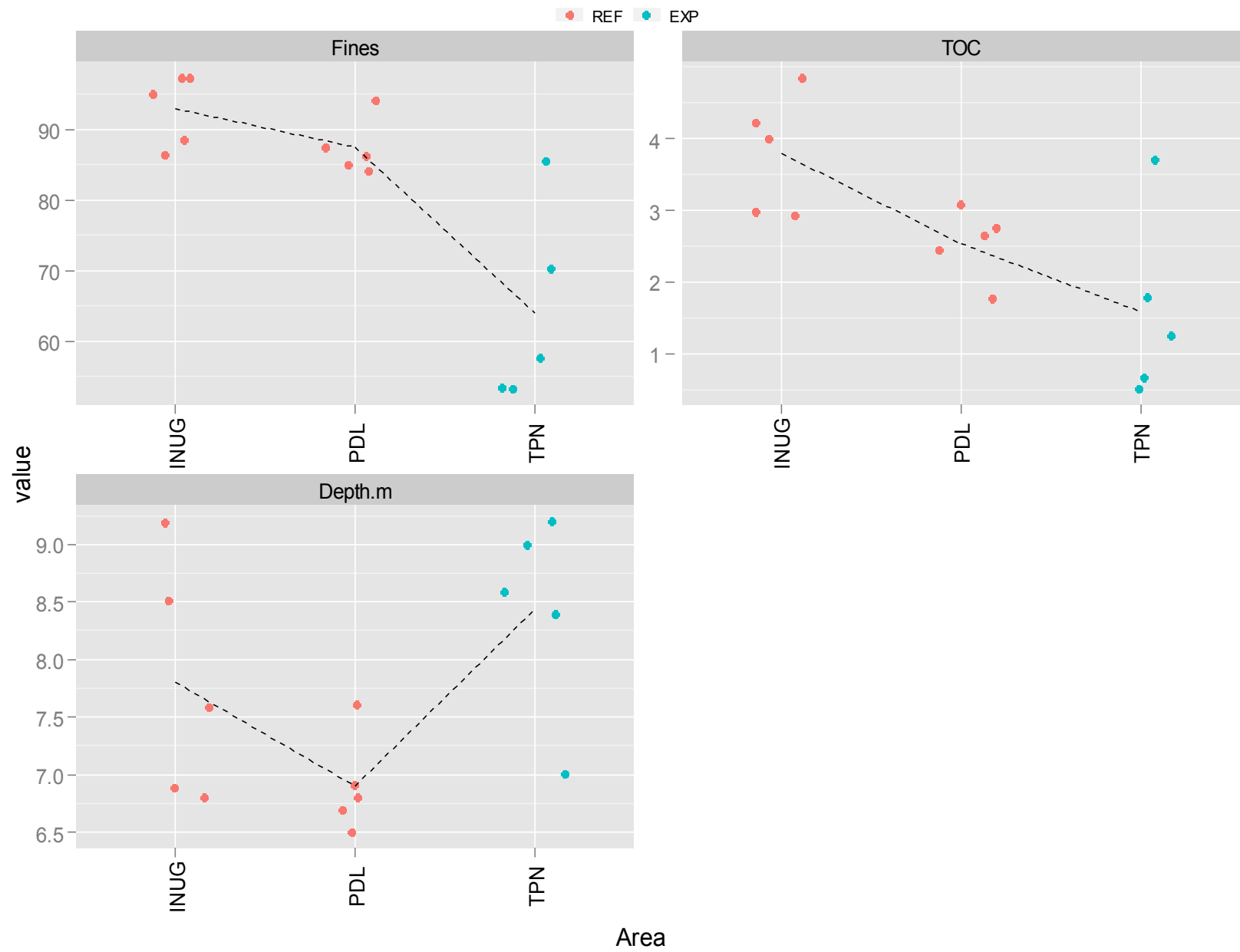


Figure 5.3- 3. Benthic invertebrate community endpoint means and standard errors for pooled reference area and exposure area, EEM Data.

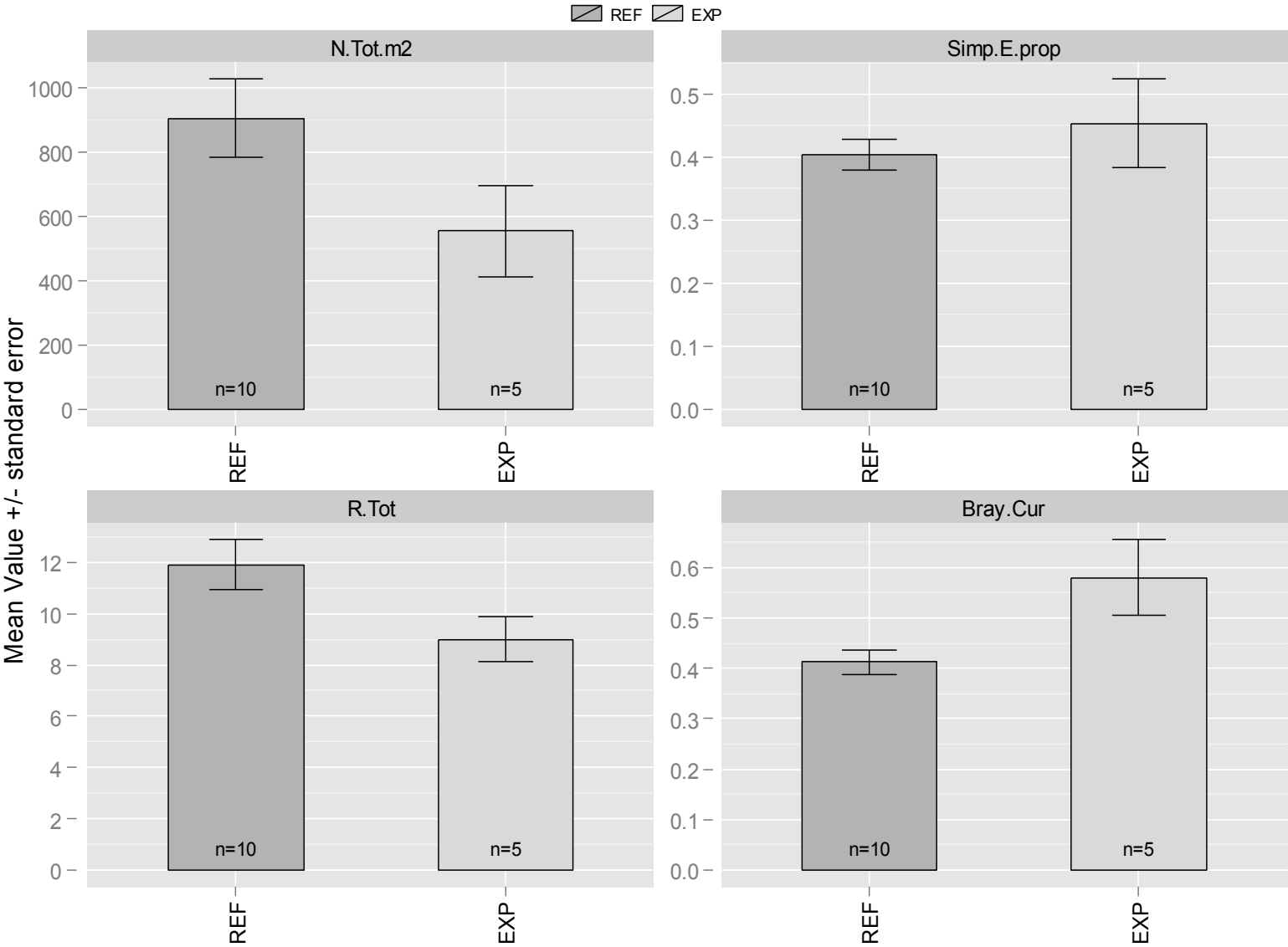


Figure 5.3- 4. Benthic invertebrate community supporting environmental endpoint means and standard errors for pooled reference area and exposure area, EEM Data.

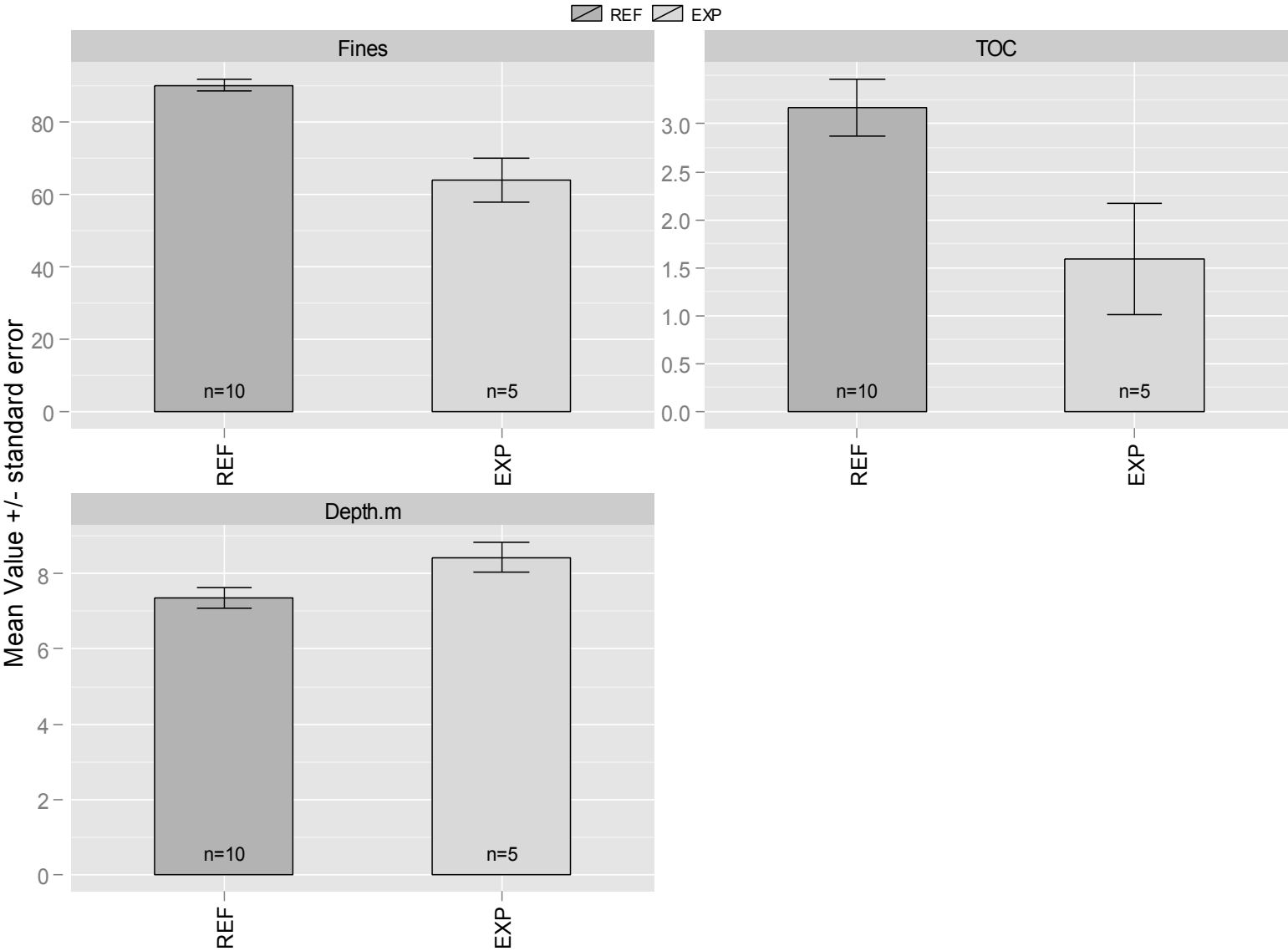


Figure 5.3- 5. Benthic invertebrate community endpoint means and standard errors for pooled reference areas plotted along with the EEM-recommended critical effect size (+/- 2 SD), EEM Data.

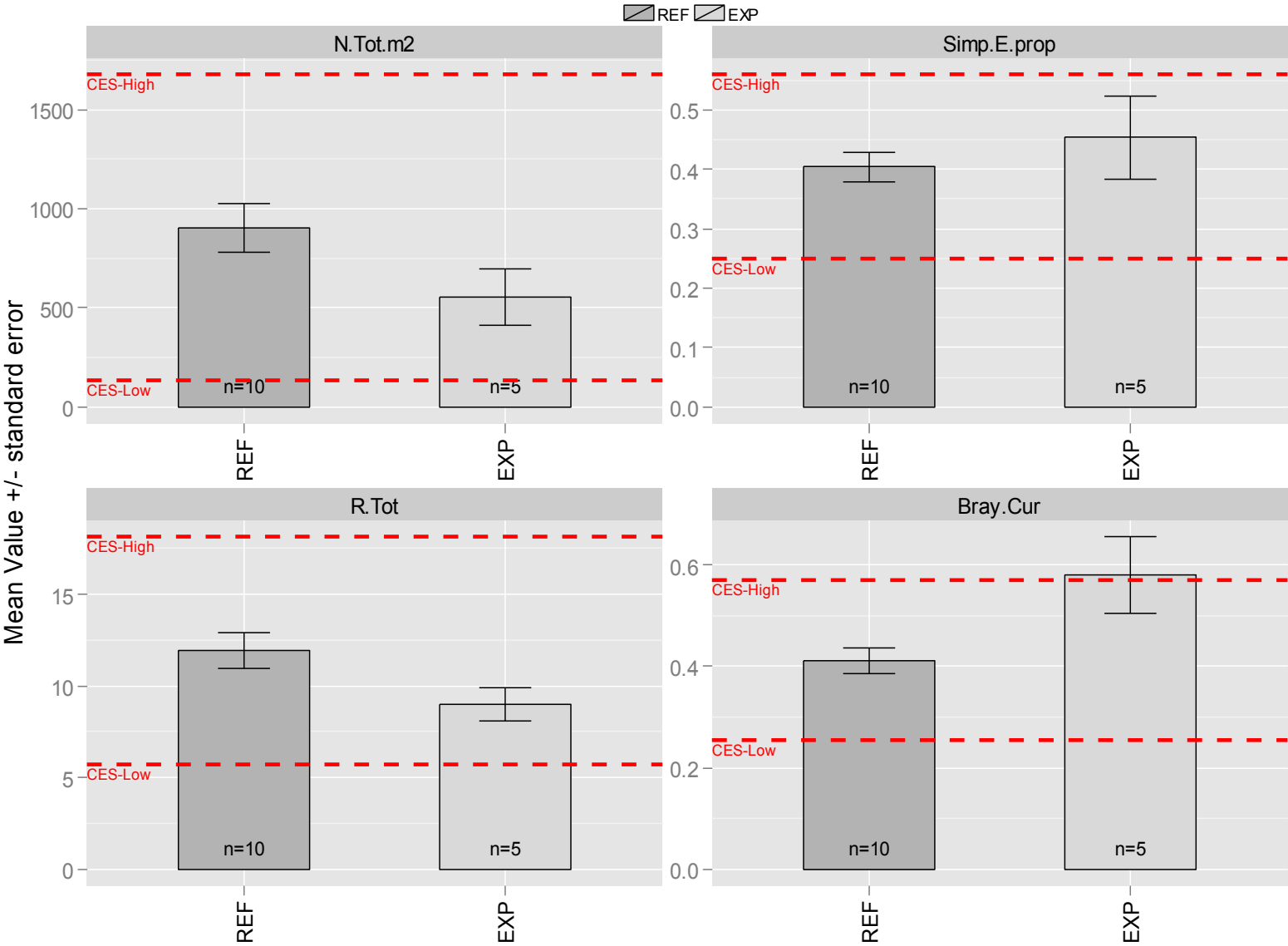
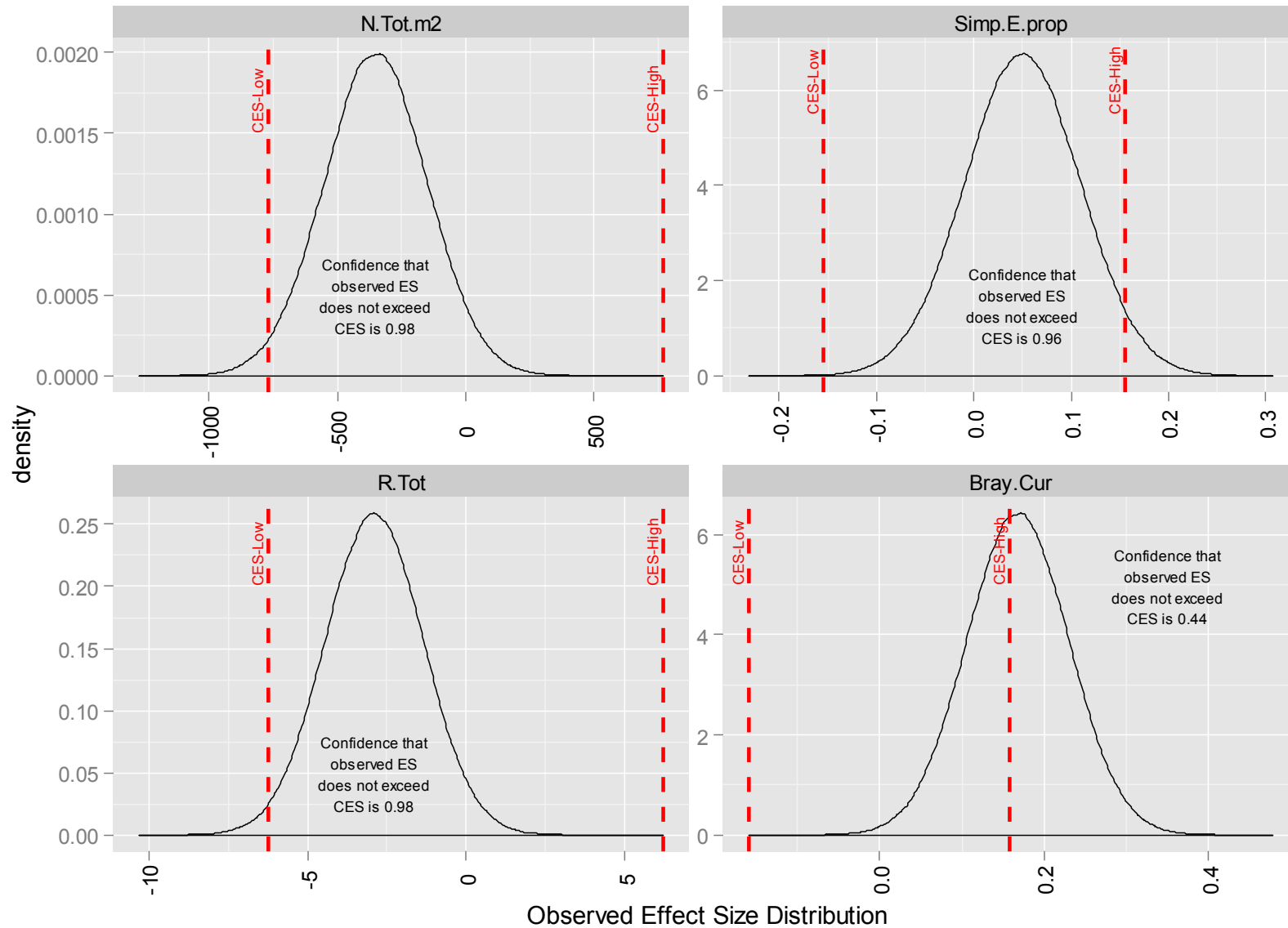


Figure 5.3- 6. Simulated effect size distributions (in original units of each endpoint) for benthic invertebrate community endpoints showing the confidence estimates of exceeding the EEM-recommended critical effect size (± 2 SD), C-I ANOVA on EEM Data.



5.4. Supplementary Analyses

This section presents the results of a number of different statistical analyses conducted to put the observed differences in EEM benthic invertebrate community endpoints into context (see **Figure 5.4-1**). This included testing whether the initial conclusions were more broadly applicable (i.e., when a data set with more reference areas was used) and testing to discern the causes of the observed differences.

Most of the supplementary analyses use the CREMP data set, which provides EEM-related insights over a wider spatial and temporal range. While the EEM and CREMP data sets were collected using virtually the same methods (e.g., sampling locations, grab type, sieve mesh, etc.), the one key difference between the data sets is the number of subsample grabs (i.e., CREMP has two and EEM has three) composited for each of the five stations per sampling area.

This section covers the following components

- Comparability of EEM and CREMP data sets.
- Summary of applicable CREMP data.
- Conduct CI-based ANOVA with CREMP data.
- Report on before-after-control-impact (BACI) model conducted with CREMP data.
- Assessment of the importance of supporting environmental variables as covariates with the benthic invertebrate community endpoints using CREMP data.
- Analysis of MC-I design with important supporting environmental variables using CREMP data.
- Reference condition approach (RCA) using CREMP data.

Each of the above analyses will be presented and discussed in the following sections. The overall results will be assessed using a weight-of-evidence approach in **Section 5.5**.

5.4.1. Comparability of 2011 EEM and 2011 CREMP Data Sets

As described above, the main difference between the two data sets is the number of subsample grabs (i.e., CREMP has two and EEM has three) composited for each of the five stations per sampling area. In an effort to reduce duplication of monitoring programs at Meadowbank, methods (e.g., Elliot equation and taxa-area curves) outlined in the EEM TGD (Environment Canada, 2012) were used to assess the number of subsamples required to adequately characterize total density and family-level richness. Based on tests of these two variables at four different locations, the highest subsample estimate was 2.4. However, there were anomalies in one of the subsamples at the station where the 2.4 result was obtained for total density. Consequently, both the original study design document and the addendum included recommendations for maintaining consistency with the CREMP by collecting two subsample grabs per station. Notwithstanding, the project's Technical Advisory Panel (TAP) recommended that benthic invertebrate samples be comprised of three subsamples to be consistent with the default for EEM and allow for a more comprehensive assessment of the two data sets.

While the benefits of having more data (more reference areas and more years) are somewhat obvious, this section explores whether the two-grab CREMP data set poses any constraints to detecting statistical differences between reference and exposure areas. The main concern would be whether the two-grab samples inflate estimates of within-area variability (i.e., due to insufficient characterization) relative to the EEM data, thus reducing statistical power.

Side-by-side EEM (all data = TPN, PDL, INUG in 2011) vs. CREMP (subset of data to include only TPN, PDL, INUG in 2011) raw data (by area; **Figure 5.4-2**) and means and standard errors for each benthic invertebrate community endpoint (except for Simpson's Evenness, which is not calculated in the CREMP) are shown by pooled reference area and exposure area in **Figure 5.4-3**. Given the concerns highlighted above regarding statistical power, power analysis (see text box in **Section 5.3.4**) was used to compare the two data sets (**Table 5.4-1**). In two of three cases, the minimum detectable effect size was actually smaller (i.e., better) for the CREMP data set. Power to detect the ± 2 SD CES was somewhat lower for the CREMP data set for the Similarity Index, but that result was mainly due to a smaller reference area standard deviation (and hence a smaller SD-based CES). The CREMP and EEM results are very similar and there is no reason to believe that using the CREMP data would limit our ability to make defensible inferences regarding potential differences between reference and exposure areas. Over and above the obvious cost savings in terms of laboratory and statistical analyses, the more extensive (spatially and temporally) CREMP data set will ultimately help make better inferences regarding potential effluent-related effects to the benthic invertebrate community.

5.4.2. Summary of CREMP Data

The CREMP has been conducted at Meadowbank since 2006. The first two years of monitoring were conducted prior to any major development activity at the site, thus representing baseline conditions. For the CREMP, tracking spatial and temporal differences related to mining activities relied on categorizing sampling areas using two factors:

- *Type* – this concept relates to an area's spatial proximity to the planned mine development, whether built or not; categories include near-field, mid-field, far-field, and reference (see text for more details).
- *Status* – this concept has two levels: control (not exposed to mine-related activity) or impact (exposed to mine-related activity). The term “impact” is taken from the BACI statistical study design approach and does not mean that an actual impact has taken place; rather, it designates a time period when potential mine-related impacts may occur for an area.

The construction phase of the project started in 2008 and the operations phase in 2010. Discharge of dewatering effluent started in spring 2009. **Table 5.4-2** provides a temporal look at the status of the CREMP monitoring areas (excluding those in Baker Lake). Based on their continued “control” status, the following CREMP areas are considered suitable reference areas for the TPN exposure area (see **Figure 3.3-7** for a map of these sampling areas):

- INUG and PDL (these are the EEM reference areas)
- South Basin of Third Portage Lake (TPS)

- Tehek Lake Far-field (TEFF)

Simpson's evenness is not calculated in the CREMP (and there was no statistically significant difference for this endpoint in the EEM data), so the focus of subsequent analyses is on total density, total richness and similarity index. Monitoring results for the benthic invertebrate community endpoints are presented in **Figure 5.4-4** (raw results) and **Figure 5.4-5** (area means and standard error). Monitoring results for the supporting environmental endpoints are presented in **Figure 5.4-6** (raw results) and **Figure 5.4-7** (area means and standard error). The figures show a vertical line at TPN representing the dividing line¹³ when the area changed from "control" to "impact" status.

5.4.3. CI-based ANOVA – 2011 CREMP Data

The statistical analyses of the MC-I EEM data were conducted following a hierarchical approach (see **Section 2.2** for rationale). In this section, the CI-based (i.e., pooled reference area) approach is conducted using the 2011 CREMP data comprised of the same areas used in the EEM study (TPN, PDL and INUG), but with two additional reference areas (TPS and TEFF).

The general procedures were the same as those reported in **Section 5.3.3**. Plots of the data by sampling area are shown in **Figures 5.4-3 through 5.4-6**; plots of pooled reference area vs. exposure area for the benthic invertebrate community endpoints (except Evenness) are shown in **Figure 5.4-8**.

Results from the CI-based ANOVA of the CREMP data are provided in **Tables 5.4-3 and 5.4-4**. Key findings were:

- 1) No significant differences were detected between pooled reference area and the TPN exposure area for any benthic invertebrate community endpoint.
- 2) All observed effect sizes for benthic invertebrate community endpoints were within 1 standard deviation of the pooled reference area mean (i.e., below the EEM-recommended CES).
- 3) Significant differences were observed for Fines and Depth.

The lack of significant differences in these results suggest that the differences detected in the EEM CI-based ANOVA analyses are sensitive to which reference areas are used in the CI comparison. This highlights the challenges of making meaningful inferences regarding potential mine-related effects without a broader understanding of natural variability in unimpaired areas.

¹³ Note that the dividing line is shown between 2008 and 2009, while dewatering discharges to TPN started in spring 2009. The reason for this is that benthic invertebrate community samples are collected in August each year.

5.4.4. CREMP Before-After-Control-Impact (BACI) Model

CREMP monitoring has been conducted since 2006 at a number of lakes surrounding the Meadowbank Mine. As such, it includes pre-development baseline data and thus provides context regarding spatial and temporal variability in the benthic community. The CREMP data set was used as part of the TSS Effects Assessment Study (Azimuth, 2012b) that was initiated in response to concerns regarding short-term and long-term adverse effects related to sediment releases during dike construction. In addition, the TSS EAS study also included a BACI analysis of the TPN benthic invertebrate community to provide insights into the potential for adverse effects related to TSS in dewatering effluent. This section reports on the results of that study at a fairly high level; details are available in the report (Azimuth, 2012b).

The BACI model was run as a mixed effects model using the lmer package in R. Model structure was as follows:

$$\text{lmer}(y \sim \text{Area} + \text{Year} + \text{Effect} + (1|\text{Area}:\text{Year}))$$

For a mixed-effects BACI model with Year and Effect as fixed effects and Area-Year as the random effect, overall significance of the Effect term would be tested by defining the appropriate *F* ratio as $\text{MS}[\text{Effect}]/\text{MS}[\text{Area}*\text{Year}]$. This is consistent with the discussion in **Section 2.2**.

The BACI model focused on single-year and multi-year effects to benthos at four key “impact” (i.e., exposed to mine-related activities) locations compared to spatial-temporal controls (i.e., area-year combinations not exposed to mine-related activities). The results reported for TPN in the 2011 TSS EAS (Azimuth, 2012b) were as follows:

“TPN (2009, 2010, 2011 and 2009-2011) – ... this area was potentially exposed to elevated TSS during dewatering of the NW arm of Second Portage Lake starting in March 2009. During dewatering, AEM was required to discharge water that was lower than TSS thresholds to minimize the potential for adverse effects in Third Portage Lake north basin. Based on CREMP water quality (Azimuth, 2010a; 2011b; 2012a) and Bay-Goose dike construction monitoring (Azimuth, 2010b), TSS values in the north basin were very low (< 1 mg/L), suggesting that adverse effects to the benthic community would be unlikely. This was confirmed by the CREMP benthos results, which had no significant adverse effects. The only significant or marginal changes observed were increases for richness (2010; Table 3-8) and Simpson’s Diversity (2009-2011; Table 3-10).”

Results for the short-term (2011 only) and long-term (2009-2011) are presented in **Tables 5.4-5 and 5.4-6**, respectively. The large size of the 95% confidence intervals of the effect size estimates reflect the high degree of natural variability in this system.

Box plots showing how the various effect terms compared to the spatial-temporal controls and other impacts tested (not discussed further herein) are presented in **Figures 5.4-9 and 5.4-10**.

Using the more appropriate BACI model, there were no statistically significant adverse effects found for the primary benthic invertebrate community endpoints. Note that there was an increase in total richness in 2010, but it is hard to imagine that being considered as a difference relevant to EEM.

5.4.5. Influence of Supporting Environmental Variables – CREMP Data

Sampling of supporting environmental variables in the CREMP has evolved over time. While depth measurements have always been taken at each station, the program started out with a single composite sample for sediment quality (i.e., chemistry, TOC and grain size). In 2009, the program was expanded to three replicate samples for sediment quality, taken synoptically with the first three benthic invertebrate community stations. In 2011, the program was expanded again to collect sediment quality synoptically at all five benthic invertebrate community stations. Consequently, there is a much larger data set for Depth than there is for TOC or Fines.

The following approach was used to assess the influence of supporting environmental variables on benthic invertebrate community endpoints using the CREMP data (i.e., the subset of the data described in **Section 5.4.2**):

- 1) Benthic endpoints were plotted against each supporting environmental variable (2011 data for TPN was highlighted with large points).
- 2) Linear regression was used to test the strength of the relationship; significant relationships ($p < 0.1$) were plotted on the graphs.
- 3) Influential outliers were removed and the regression analyses repeated; significant relationships were plotted.

The results are tabulated in **Table 5.4-7** (all benthic endpoints) and shown in **Figure 5.4-11** (Total Density), **Figure 5.4-12** (Total Richness) and **Figure 5.4-13** (Similarity Index). After removal of highly influential outliers (see data on plots), the results show statistically significant relationships with TOC and Fines. For Total Density and Total Richness, the relationships with TOC were quite strong and explained 24% (in both cases) of the variability in the data. These results show that TOC and Fines should be considered when testing for potential mine-related differences between reference and exposure areas.

5.4.6. C-I Testing with Environmental Covariates – CREMP Data

The preceding section highlighted the importance of environmental variables in explaining patterns in benthic invertebrate community endpoints. Given the confounding environmental gradients seen in some of the CI ANOVA analyses, it is important to try to account for environmental covariates when testing for CI differences. Consequently, the 2009 to 2011 CREMP data set (i.e., with four reference areas: INUG, PDL, TPS, TEFF; and the TPN exposure area) was used in a linear modeling framework with the following variables:

- Response variables –benthic invertebrate community endpoints (i.e., total density, total richness and similarity index)

- Environmental covariates – based on the preceding section, this effort focuses on TOC and fines.
- Cofactors – Year (as a fixed effect testing for inter-year differences among areas) and CI (as a fixed effect testing for differences potentially related to effluent discharge) were used.

Model alternatives using these variables include linear regression and multiple linear regression with and without interactions (i.e., analogous to ANCOVA). The various models were examined for each response variable using the “stepAIC” function from the MASS package for R. This function uses Akaike’s information criterion (AIC) to decide the best (lowest AIC) model.

Results are as follows by response variable:

- *Total density* – the best model explaining patterns in total density contained TOC and Year only (combining to explain 33% of the variability). Neither Fines nor CI (by itself explaining <1% of the variability) contributed sufficiently to be included.
- *Total richness* – the best model explaining patterns in total richness contained TOC and Year only (combining to explain 30% of the variability). Neither Fines nor CI (by itself explaining virtually 0% of the variability) contributed sufficiently to be included.
- *Similarity index* – the best model explaining patterns in similarity index contained Fines only (explaining 11% of the variability).

In summary, after accounting for natural variation in TOC and Fines, the factor CI (which would indicate differences potentially related to effluent discharge) does not explain results for the benthic invertebrate community. The lack of a statistically significant relationship between any of the response variables and CI suggests that the differences seen in the EEM data set are likely to be due to the differences in physical environmental variables.

5.4.7. Reference Condition Approach – CREMP Data

The fundamental premise of RCA is that a suitably large set of existing data can be used to characterize unimpacted conditions in terms of a variety of biological attributes. As described in the EEM TGD, some applications rely on using a database of biological and environmental endpoints that allows development of predictive models (i.e., predicting biological attributes based on environmental variables). Given that the CREMP study design attempts to reduce the influence of environmental factors (e.g., by targeting depositional habitat within a narrow depth range), habitat-based predictive models aren’t necessary to apply the RCA at Meadowbank.

The following steps were taken to conduct the RCA analysis:

- 1) All CREMP area-year combinations representing unimpacted conditions (i.e., all combinations with “impact” status in **Table 5.4-2** were excluded) were compiled to represent the reference condition (n = 27).
- 2) TPN data from 2009 through 2011 were used to represent the exposure area (n = 3).
- 3) Means were calculated for each of eight biological attribute variables (density and richness of the major taxa groups at Meadowbank: insects, mollusks, oligochaetes, and

“other” [e.g., flatworms, mites, fairy shrimp] taxa) using the replication stations for each area-year combination.

- 4) As described in EEM TGD (Chapter 4.4.3), non-metric multidimensional scaling (nMDS) was used to reduce the dimensionality of the multivariate data set to two main axes (i.e., each area-year combination gets a score on each axis, thus providing its relative location in XY space).
- 5) Reference area-year XY scores were plotted on an ordination biplot along with 90%, 95% and 99% probability ellipses (i.e., each representing the probability that reference area-year combinations lie within the ellipse).
- 6) Exposure area-year XY scores were then plotted on the biplot to determine where they were situated in relation to the probability ellipses.

RCA results are presented in **Figure 5.4-14** and show that all three TPN exposure area-year combinations fall within the 90% probability ellipse. According to the EEM TGD, Reynoldson et al. (1985, as cited in Environment Canada, 2012) concluded that sites located in ordination space within the 90% probability ellipse would be considered as equivalent to reference and therefore unstressed. Thus, the RCA results suggest that no TPN exposure area-year combinations would be considered different from unimpaired conditions.

Table 5.4- 1. Comparability of EEM and CREMP benthic community data sets using power analysis.

Endpoint	Program	Obs. ES	Ref SD	CES	n (EXP/REF)	α	β	Power for CES	Minimum ES	Minumum EXP n for CES and Power
<i>Benthic Community Endpoints</i>										
Total Density log10(N.Tot.m2)	EEM	-351	385	-770	5/10	0.1	0.1	0.98	620	4
	CREMP	-415	471	-942	5/10	0.1	0.1	0.99	715	4
Total Richness (R.Tot)	EEM	-2.9	3.11	-6.2	5/10	0.1	0.1	0.98	4.77	4
	CREMP	-2	2.75	-5.5	5/10	0.1	0.1	0.98	4.36	4
Similarity (Bray.Cur)	EEM	0.17	0.0791	0.158	5/10	0.1	0.1	0.78	0.193	7
	CREMP	0.16	0.0579	0.116	5/10	0.1	0.1	0.57	0.187	13

Notes: "Obs. ES" = observed effect size, reported in log10 units for N.Tot.m2; "CES" = critical effect size (from EEM TGD) = 2 * Ref SD (reported here in the same direction as the Obs.ES); "n" = sample size; "Ref SD" = standard deviation of pooled reference area; "Minimum ES" reported in Obs. ES units, not as multiple of Ref SD; Minimum sample size (REF n = 2 * EXP n).

Table 5.4- 2. Status of all CREMP sampling areas since the beginning of monitoring.

	Meadowbank Sampling Areas									Baker Lake Areas			
	REF	REF	NF	NF	NF	NF	MF	MF	FF	REF	REF	NF	NF
Year	INUG	PDL	TPN	SP	TPE	WAL	TPS	TE	TEFF	BAP	BES	BBD	BPJ
2006	C		C	C	C	C	C	C					
2007	C		C	C	C	C	C	C					
2008	C		C	I (Aug)	C	C	C	I (Aug)		C		I	I
2009	C	C	I (Mar)	I	I (Aug)	C	C	I	C	C		I	I
2010	C	C	I	I	I	C	C	I	C	C		I	I
2011	C	C	I	I	I	C	C	I	C	C	C	I	I

Notes:

Area designations: C = Control; I = Impact; REF = reference; NF = near-field; MF = mid-field; FF = far-field; Blank denotes that the area was not part of the monitoring program that year.

Meadowbank Sampling Area abbreviations: INUG = Inuggugayualik Lake; PDL = Pipedream Lake; TPN = Third Portage North; SP = Second Portage Lake; TPE = Third Portage East; WAL = Wally Lake; TPS = Third Portage South; TE = Tehek Lake; TEFF = Tehek Lake Far Field.

Table 5.4- 3. Summary of CI-based ANOVA tests for benthic invertebrate community and supporting environmental endpoints, CREMP Data.

Endpoint	Source of Variation	Sum of Squares	df	Mean Square	F	p-value	Effect?
Benthic Community Endpoints							
Total Density	Between Groups (C-I)	578923	1	578923	2.600	0.121	No
N.Tot.m2	Within Groups (Residual:	5121448	23	222672			
	Total	5700371	24				
Total Richness	Between Groups (C-I)	6.8	1	7	1.035	0.320	No
(R.Tot)	Within Groups (Residual:	150.2	23	7			
	Total	157.0	24				
Similarity	Between Groups (C-I)	0.016	1	0	0.936	0.343	No
(Bray.Cur)	Within Groups (Residual:	0.390	23	0			
	Total	0.406	24				
Supporting Sediment Endpoints							
TOC	Between Groups (C-I)	7.37	1	7.4	6.791	0.016	No
	Within Groups (Residual:	24.95	23	1.1			
	Total	32.31	24				
Fines	Between Groups (C-I)	2127	1	2127	12.803	0.002	Yes
	Within Groups (Residual:	3821	23	166			
	Total	5948	24				
Depth	Between Groups (C-I)	4.8	1	4.8	8.063	0.009	Yes
	Within Groups (Residual:	13.7	23	0.6			
	Total	18.5	24				

Notes: "df" = degrees of freedom; "F" = statistical F ratio; "C-I" = control-impact (or REF-EXP); "Effect" = significant (p<0.1) difference between pooled REF and EXP.

Table 5.4- 4. Summary of CI-based effect size information for benthic invertebrate community and supporting environmental endpoints, CREMP Data.

Endpoint	Area	Mean	Ref SD	Obs. ES	% ES	ES/Ref SD	Effect?
<i>Benthic Community Endpoints</i>							
Total Density	Reference	945	502				
N.Tot.m2	Exposure	565		-380	-40%	-0.8	No
Total Richness	Reference	9.3	2.64				
(R.Tot)	Exposure	8		-1.3	-14%	-0.5	No
Similarity	Reference	0.497	0.117				
(Bray.Cur)	Exposure	0.560		0.06	13%	0.5	No
<i>Supporting Sediment Endpoints</i>							
TOC	Reference	2.9	1.0				
	Exposure	1.6		-1.4	-46%	-1.4	No
Fines	Reference	87.0	12.7				
	Exposure	64.0		-23.0	-26%	-1.8	Yes
Depth	Reference	7.3	0.8				
	Exposure	8.4		1.1	15%	1.5	Yes

Notes: "Ref SD" = pooled REF standard deviation; "Obs. ES" = observed effect size; "% ES" = 100*(EXP Mean - REF mean)/REF mean; "Effect" = significant (p<0.1) difference between pooled REF and EXP.

Table 5.4- 5. Results of statistical analyses of benthic invertebrate community descriptors for the 2011 CREMP data set; short-term effect.

	Total Abundance (#/m ²)	Taxa Richness (# taxa/ sample)	Simpson's Diversity (unitless) ⁵	Bray Curtis Distance (unitless) ⁵
Data Transformation ¹	Log10	None	None	None
Advanced Transformation ²	Log10(x-10)	NA	NA	NA
Tests relative to controls				
C-SP2011 Differences?	No	Marginal	No	No
p-value	0.78	0.14	0.17	0.46
"No Effect" Mean ³	647	7.1	0.72	0.63
Effect Size	-85	3.1	0.08	0.18
95% Upper CI ⁴	-458	-1.2	-0.04	-0.23
95% Lower CI ⁴	1068	7.5	0.21	1.01
C-TE2011 Differences?	No	No	No	No
p-value	0.87	0.89	0.29	0.95
"No Effect" Mean	791	6.5	0.66	0.60
Effect Size	-68	-0.3	0.07	-0.01
95% Upper CI ⁴	-559	-4.9	-0.07	-0.33
95% Lower CI ⁴	1516	4.3	0.20	0.68
C-TPE2011 Differences?	No	No	No	No
p-value	0.42	0.87	0.62	0.41
"No Effect" Mean	2357	9.7	0.72	0.77
Effect Size	-774	-0.3	0.03	-0.19
95% Upper CI ⁴	-1803	-4.4	-0.09	-0.49
95% Lower CI ⁴	2205	3.8	0.15	0.44
C-TPN2011 Differences?	No	No	Marginal	No
p-value	0.38	0.52	0.09	0.48
"No Effect" Mean	779	6.7	0.72	0.69
Effect Size	-282	1.3	0.10	-0.15
95% Upper CI ⁴	-608	-3.0	-0.02	-0.44
95% Lower CI ⁴	703	5.5	0.23	0.46

Notes:

1. Initial transformation options discussed in **Section 2**.
2. Advanced transformations determined using Box-Cox method (Venables & Ripley 2002).
3. "No Effect" Mean estimated by not including the Effect coefficient when estimating Area-Year mean.
4. Confidence intervals estimated using 2 * Std. Error of Effect Size estimate.
5. Model assumptions not met, see text for details.

Table 5.4- 6. Results of statistical analyses of benthic invertebrate community descriptors for the 2006 to 2011 CREMP data set; long-term effect.

	Total Abundance (#/m ²)	Taxa Richness (# taxa/ sample)	Simpson's Diversity (unitless) ⁵	Bray Curtis Distance (unitless) ⁵
Data Transformation ¹	Log10	None	None	Log10
Advanced Transformation ²	Log10(x+50)	NA	NA	NA
Tests relative to controls				
C-SP'08-'11 Differences?	Marginal	No	Marginal	No
p-value	0.10	0.61	0.10	0.87
"No Effect" Mean ³	836	8.05	0.71	0.60
Effect Size	-379	-0.8	0.07	0.02
95% Upper CI ⁴	-625	-4.3	-0.01	-0.17
95% Lower CI ⁴	101	2.6	0.15	0.29
C-TE'08-'11 Differences?	No	No	Marginal	No
p-value	0.29	1.00	0.07	0.89
"No Effect" Mean ³	1018	7.41	0.65	0.60
Effect Size	-335	-0.01	0.08	0.01
95% Upper CI ⁴	-710	-3.70	-0.01	-0.18
95% Lower CI ⁴	432	3.68	0.17	0.30
C-TPE'09-'11 Differences?	Yes	No	Yes	No
p-value	0.02	0.97	0.02	0.11
"No Effect" Mean ³	2866	10.02	0.70	0.70
Effect Size	-1484	-0.06	0.10	-0.16
95% Upper CI ⁴	-2139	-3.20	0.02	-0.32
95% Lower CI ⁴	-277	3.09	0.17	0.05
C-TPN'09-'11 Differences?	No	No	Yes	No
p-value	0.67	0.24	0.01	0.33
"No Effect" Mean ³	963	7.08	0.69	0.63
Effect Size	-130	1.96	0.11	-0.10
95% Upper CI ⁴	-556	-1.43	0.03	-0.26
95% Lower CI ⁴	692	5.34	0.19	0.13

Notes:

1. Initial transformation options discussed in **Section 2**.
2. Advanced transformations determined using Box-Cox method (Venables & Ripley 2002).
3. "No Effect" Mean estimated by not including the Effect coefficient when estimating Area-Year mean.
4. Confidence intervals estimated using 2 * Std. Error of Effect Size estimate.
5. Model assumptions not met, see text for details.

Table 5.4- 7. Linear regression results testing for relationships between benthic invertebrate community endpoints and supporting environmental variables, CREMP Data.

Data	Dependent Variable	Independent Variable	n	slope	intercept	p	significant	adj. R ²	Figure #
CREMP - all	Log10(N.Tot.m2)	Depth.m	111	0.033	2.74	0.122	no	0.01	5.4-11
CREMP - out	Log10(N.Tot.m2)	Depth.m	105	0.017	2.85	0.41	no	0.00	5.4-11
CREMP - all	Log10(N.Tot.m2)	TOC	56	0.096	2.72	0.001	yes	0.17	5.4-11
CREMP - out	Log10(N.Tot.m2)	TOC	53	0.111	2.65	<0.001	yes	0.24	5.4-11
CREMP - all	Log10(N.Tot.m2)	Fines	54	0.005	2.55	0.05	yes	0.05	5.4-11
CREMP - all	R.Tot	Depth.m	111	0.39	6.9	0.079	yes	0.02	5.4-12
CREMP - out	R.Tot	Depth.m	108	0.34	7.24	0.19	no	0.01	5.4-12
CREMP - all	R.Tot	TOC	56	1.17	6.43	<0.001	yes	0.24	5.4-12
CREMP - all	R.Tot	Fines	56	0.076	3.43	0.005	yes	0.12	5.4-12
CREMP - out	R.Tot	Fines	50	0.013	8.29	0.56	no	0.00	5.4-12
CREMP - all	Bray.Cur	Depth.m	111	0.12	0.44	0.3	no	0.00	5.4-13
CREMP - out	Bray.Cur	Depth.m	108	0.016	0.4	0.23	no	0.00	5.4-13
CREMP - all	Bray.Cur	TOC	56	-0.022	0.58	0.011	no	0.02	5.4-13
CREMP - all	Bray.Cur	Fines	56	-0.004	0.81	0.03	yes	0.16	5.4-13

Notes: significance based on $p < 0.1$; "out" = with outliers removed; N.Tot.m2 = total density ($\#/m^2$); R.Tot = taxa richness; Bray.Cur = Bray Curtis Index; Depth.m = depth (m); TOC = total organic carbon; Fines = clay + silt (%).

Figure 5.4- 1. Schematic of data analysis for benthic invertebrate community endpoints showing supplemental analyses.

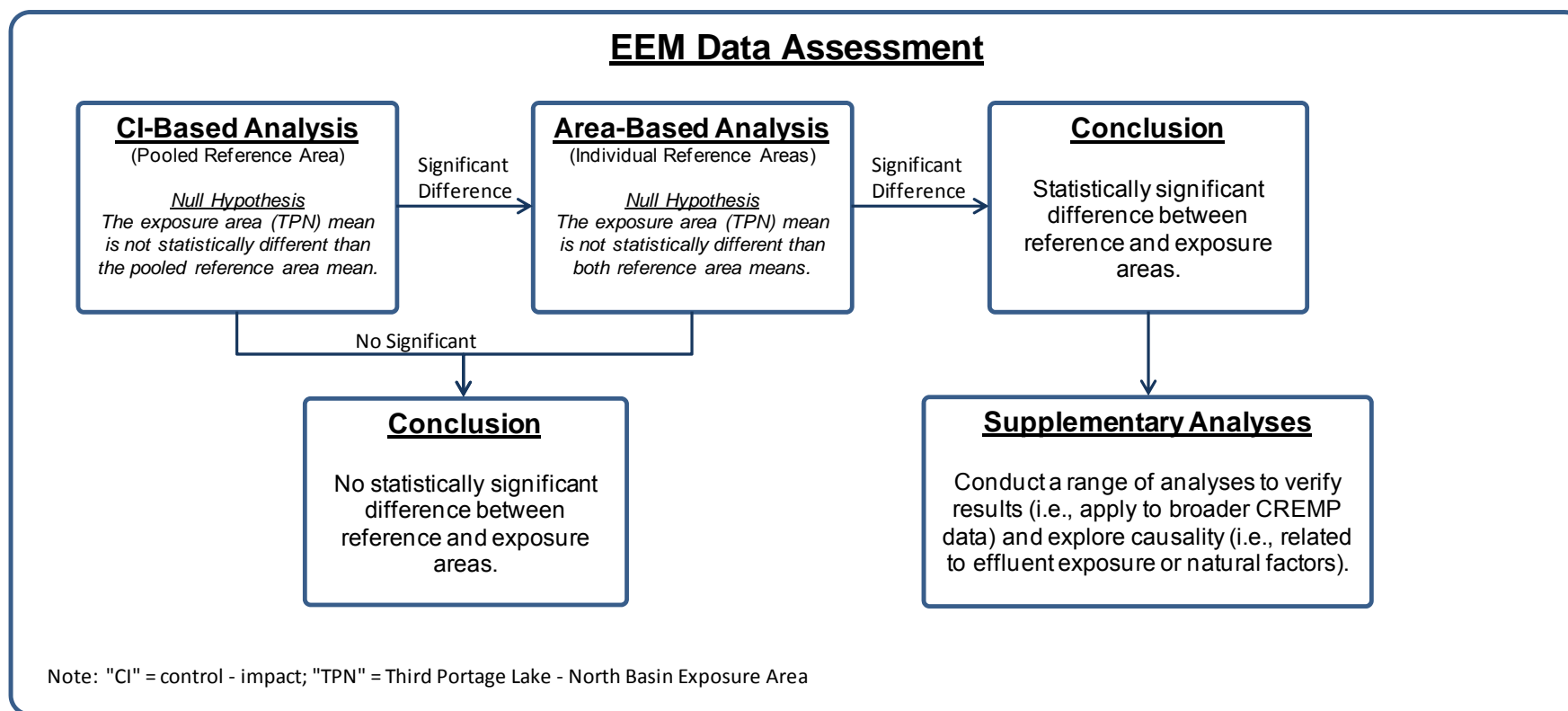


Figure 5.4- 2. Benthic invertebrate community endpoint results by area and program (CREMP, EEM).

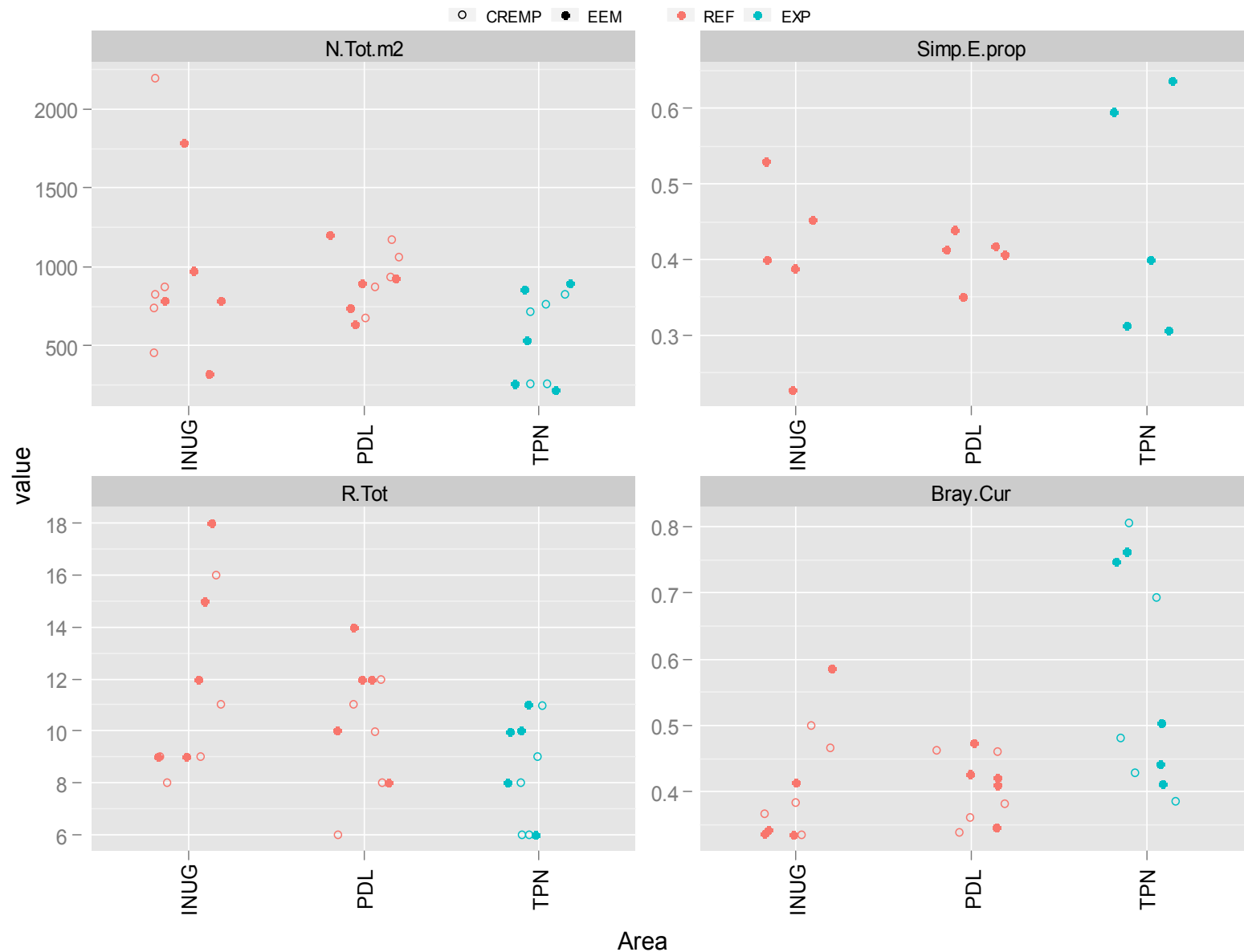


Figure 5.4- 3. Benthic invertebrate community endpoint means and standard errors for pooled reference area and exposure area by program (CREMP, EEM).

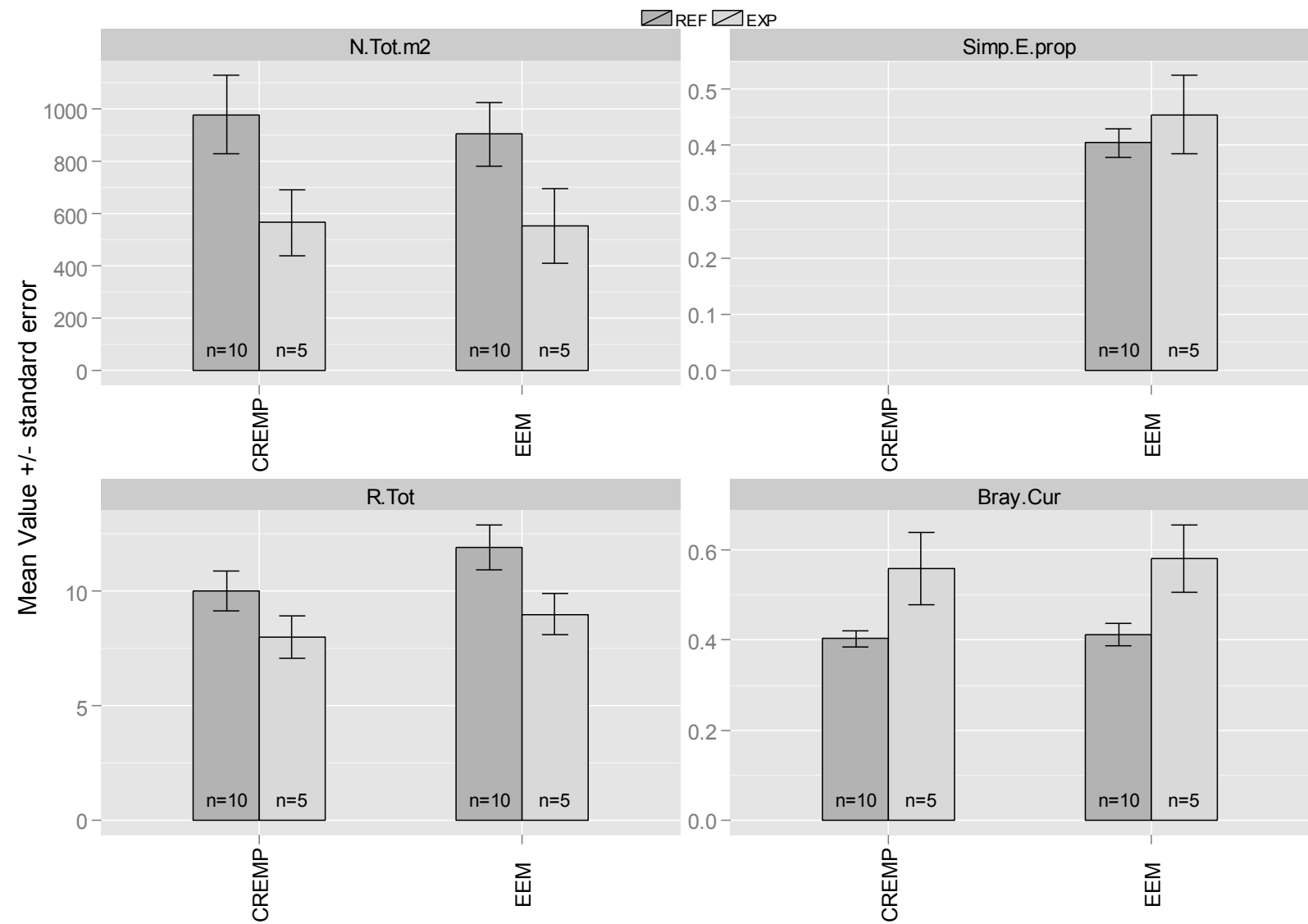


Figure 5.4- 4. Benthic invertebrate community endpoint results by year (dashed line connects annual means) and sampling area, CREMP Data.

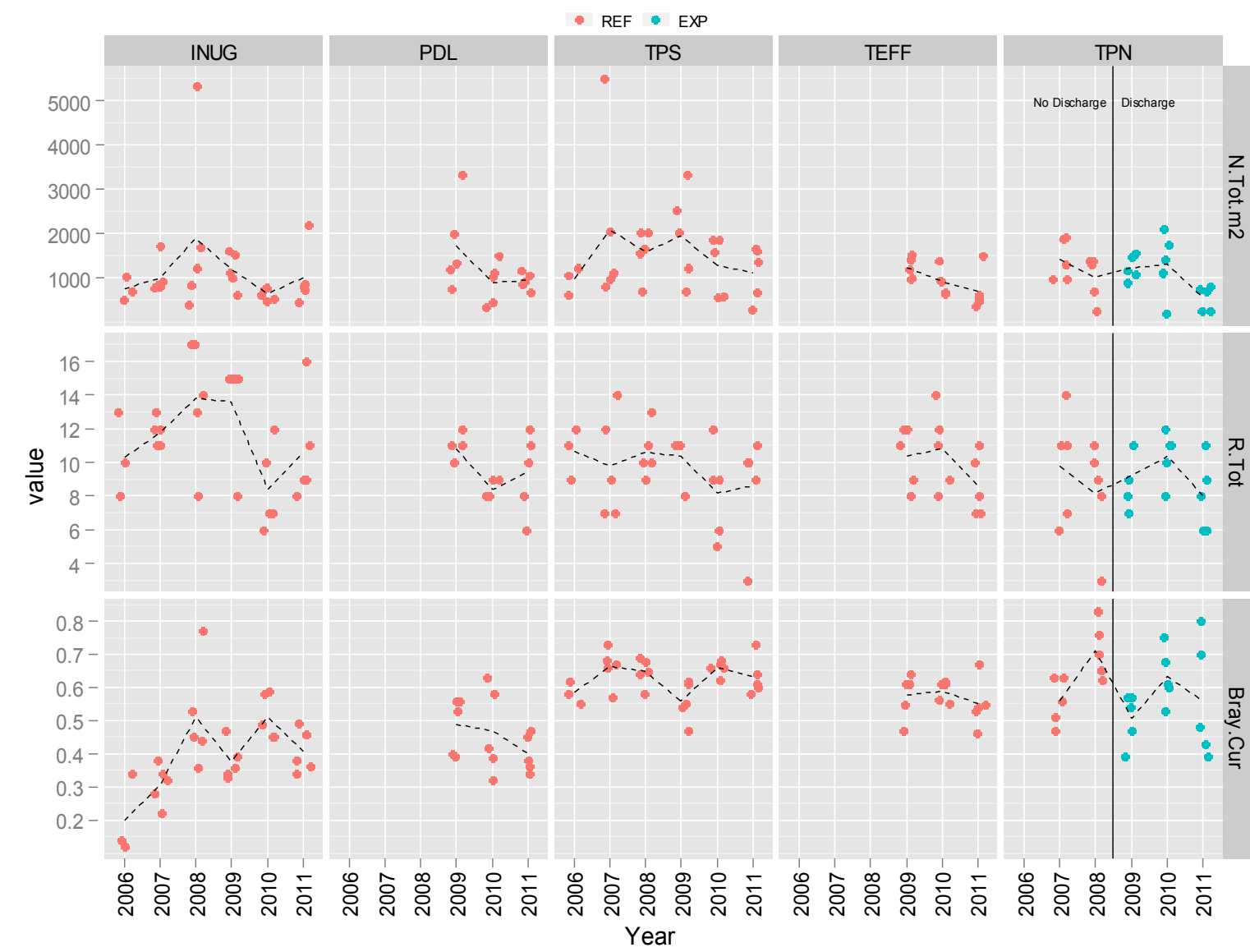


Figure 5.4- 5. Benthic invertebrate community endpoint means and standard errors by year and sampling area, CREMP Data.

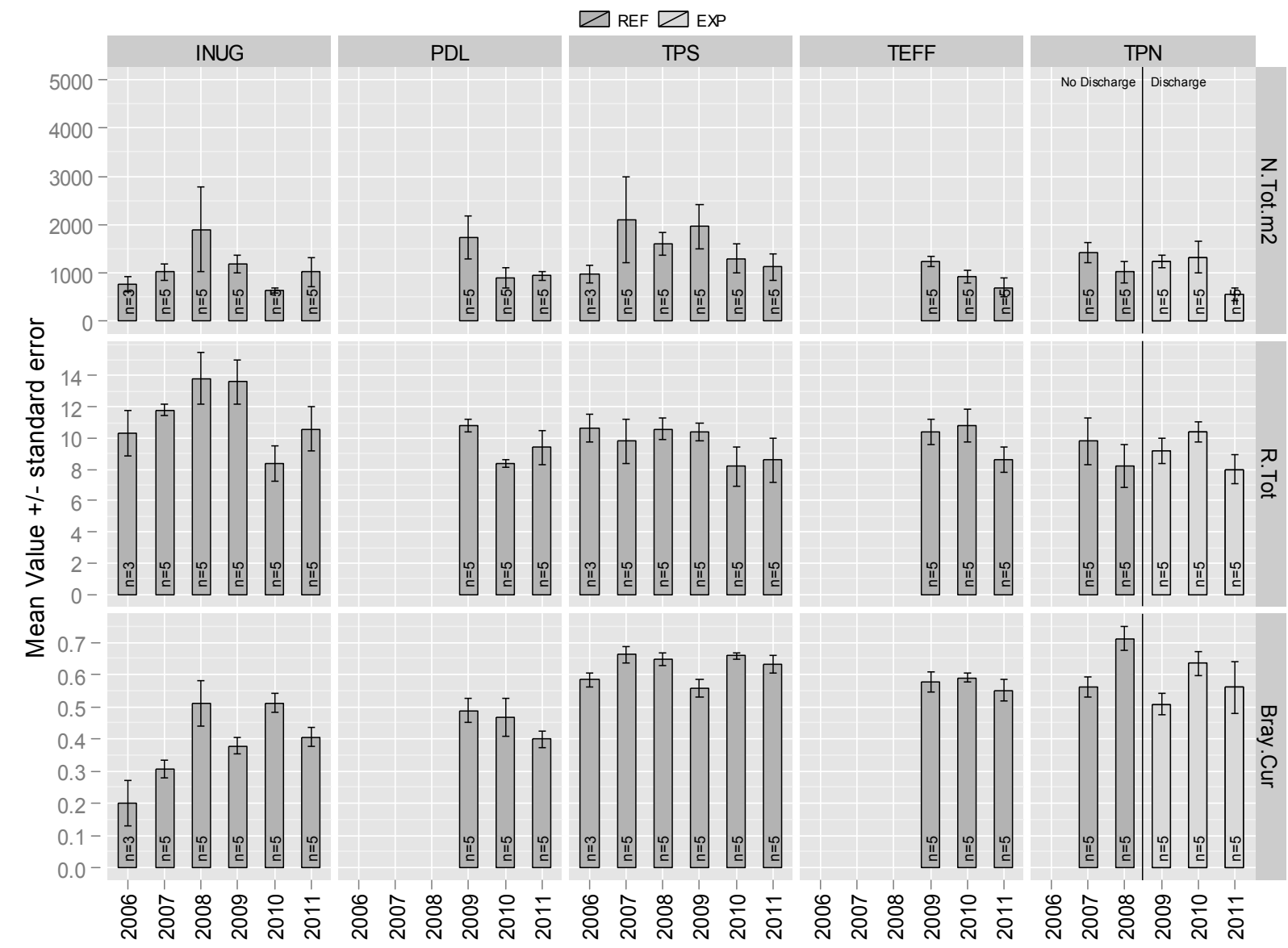


Figure 5.4- 6. Benthic invertebrate community supporting environmental endpoint results by year (dashed line connects annual means) and sampling area, CREMP Data.



Figure 5.4- 7. Benthic invertebrate community supporting environmental endpoint means and standard errors by sampling area, CREMP Data.

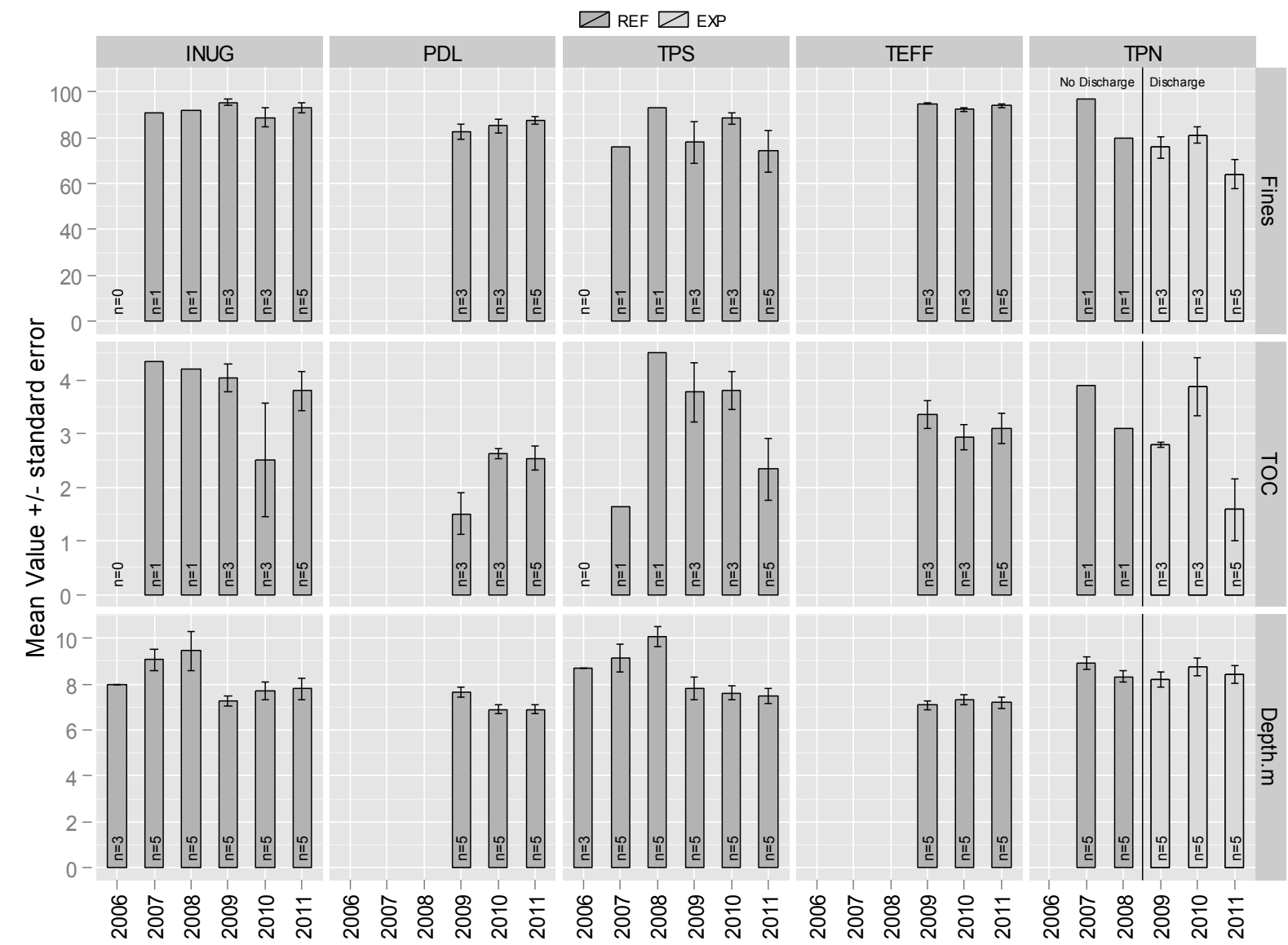


Figure 5.4- 8. Benthic invertebrate community endpoint means and standard errors by pooled reference area and exposure area, 2011 CREMP Data.

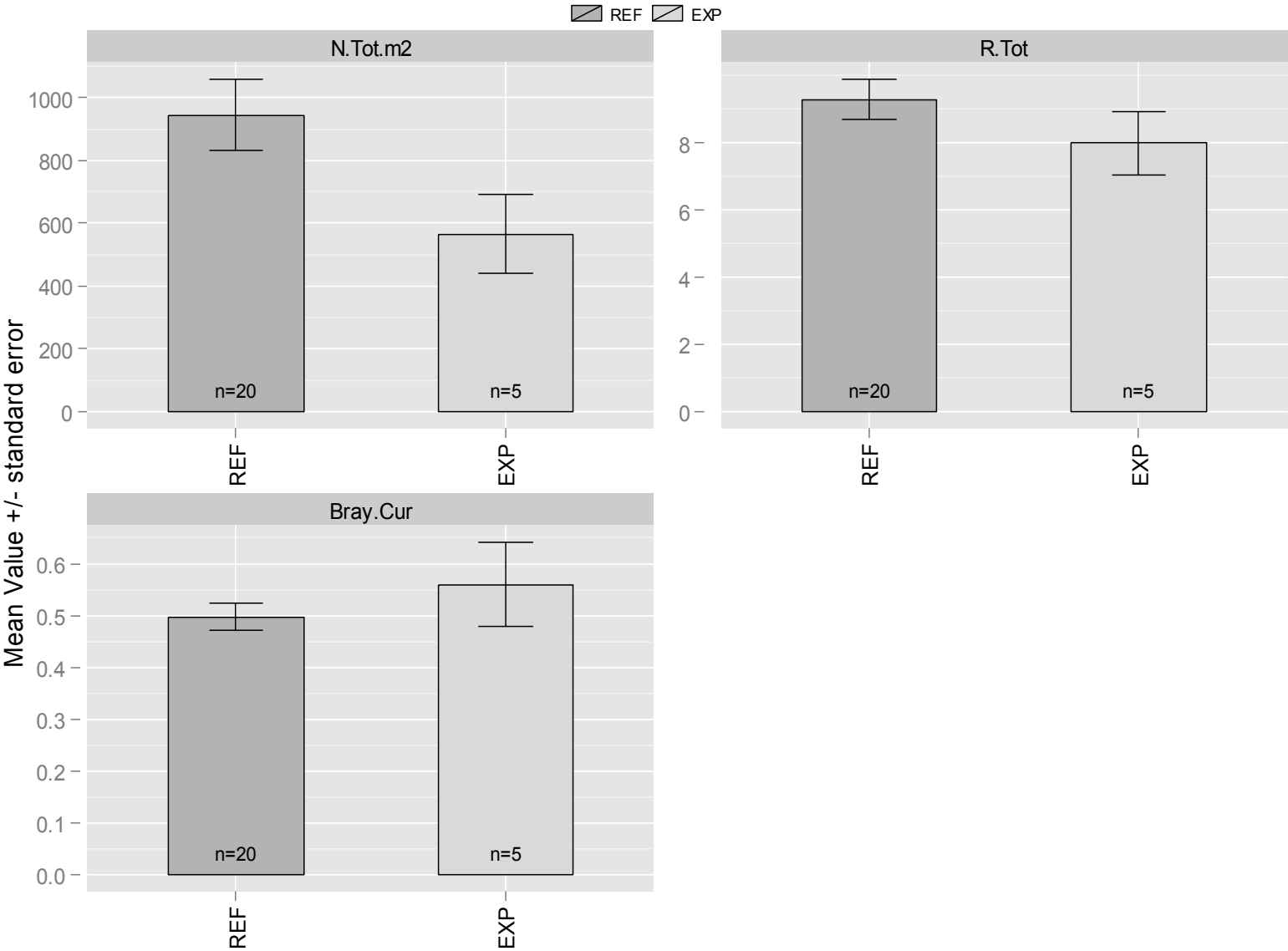


Figure 5.4- 9. Boxplots of benthic community metrics by short-term effect grouping for the CREMP BACI Model.

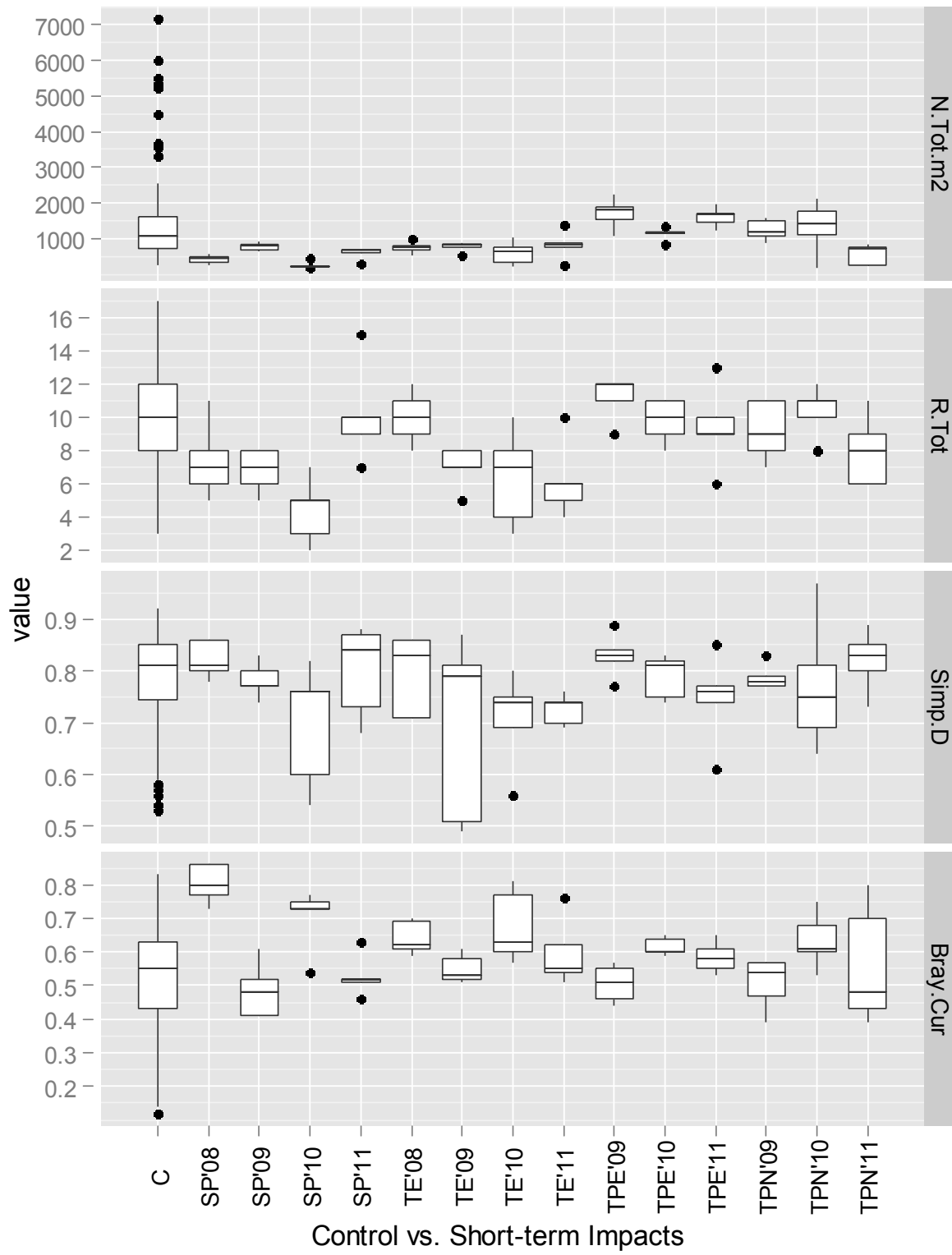


Figure 5.4- 10. Boxplots of benthic community metrics by long-term effect grouping for the CREMP BACI Model.

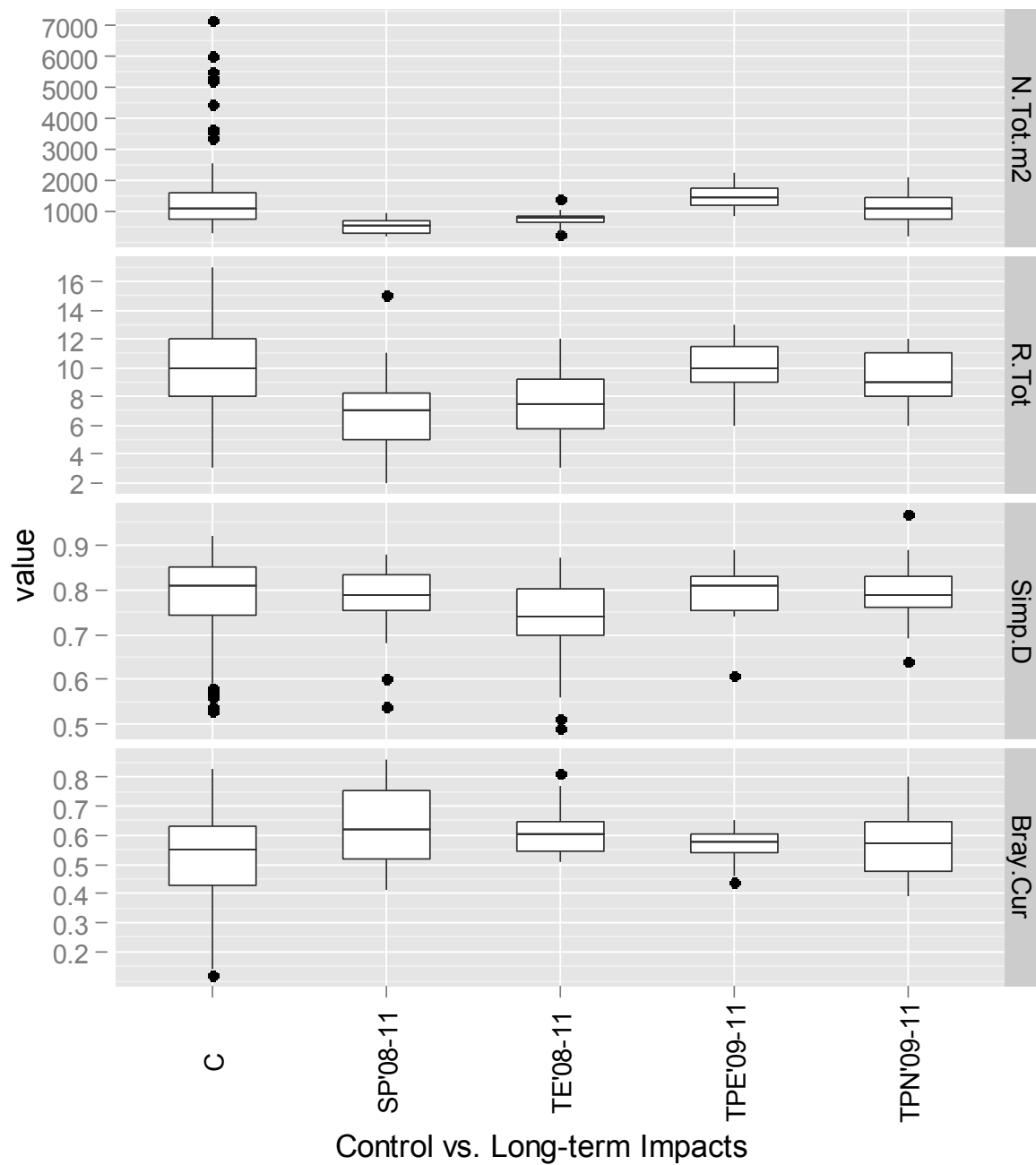


Figure 5.4- 11. Relationships (significant regressions shown in blue) between Total Density and Depth, TOC and Fines, CREMP Data.

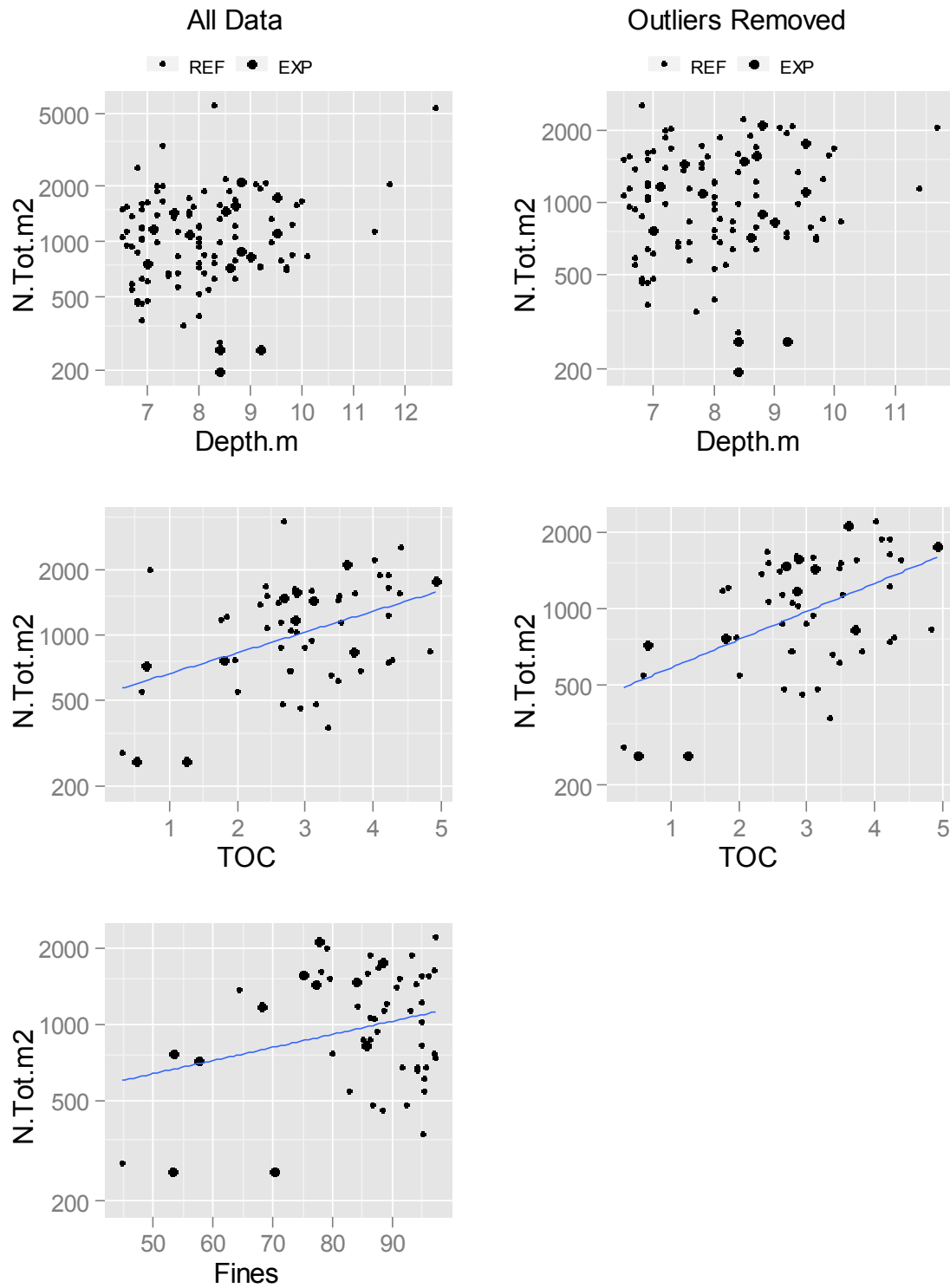


Figure 5.4- 12. Relationships (significant regressions shown in blue) between Total Richness and Depth, TOC and Fines, CREMP Data.

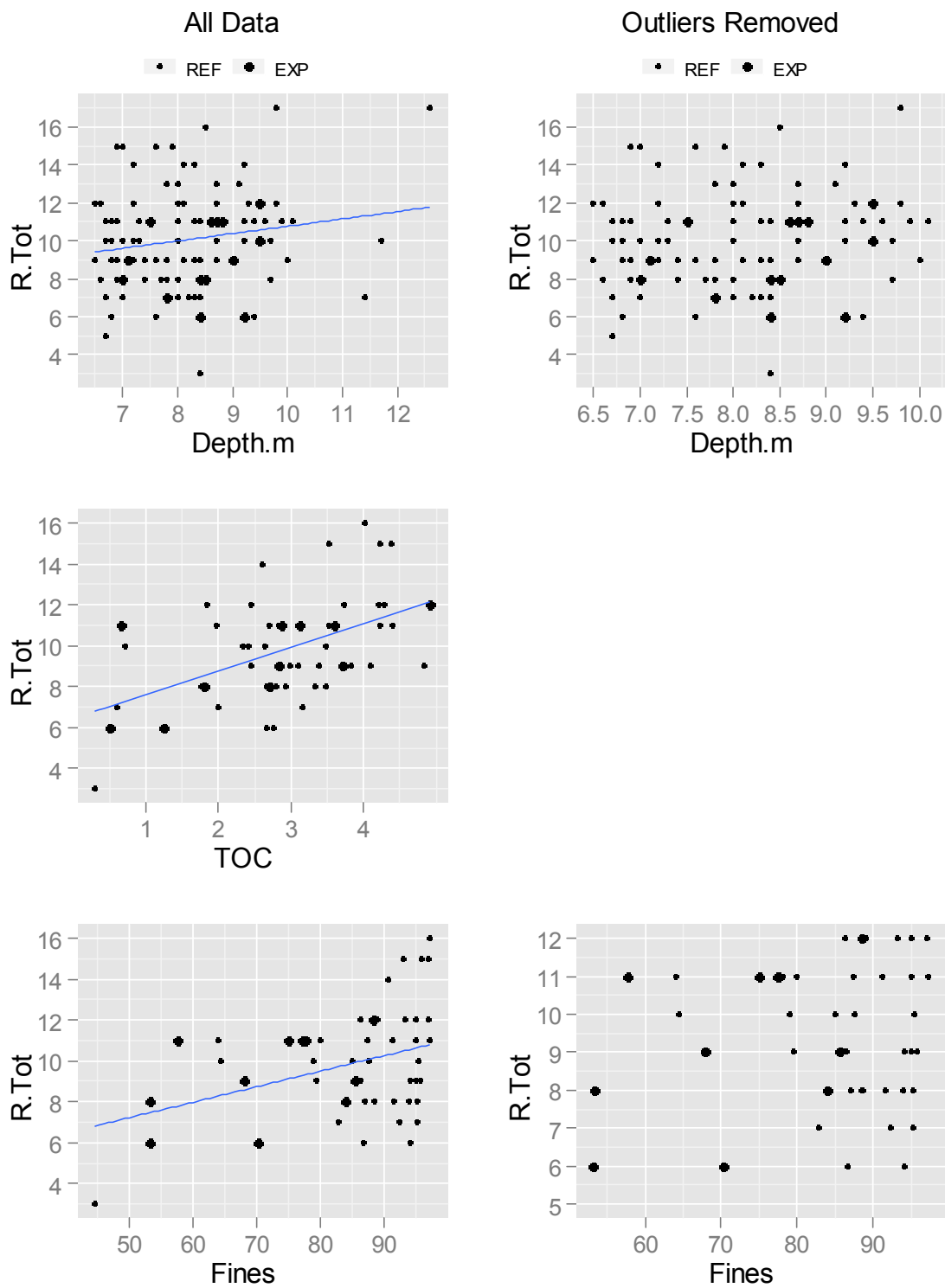


Figure 5.4- 13. Relationships (significant regressions shown in blue) between Similarity Index (Bray.Cur) and Depth, TOC and Fines, CREMP Data.

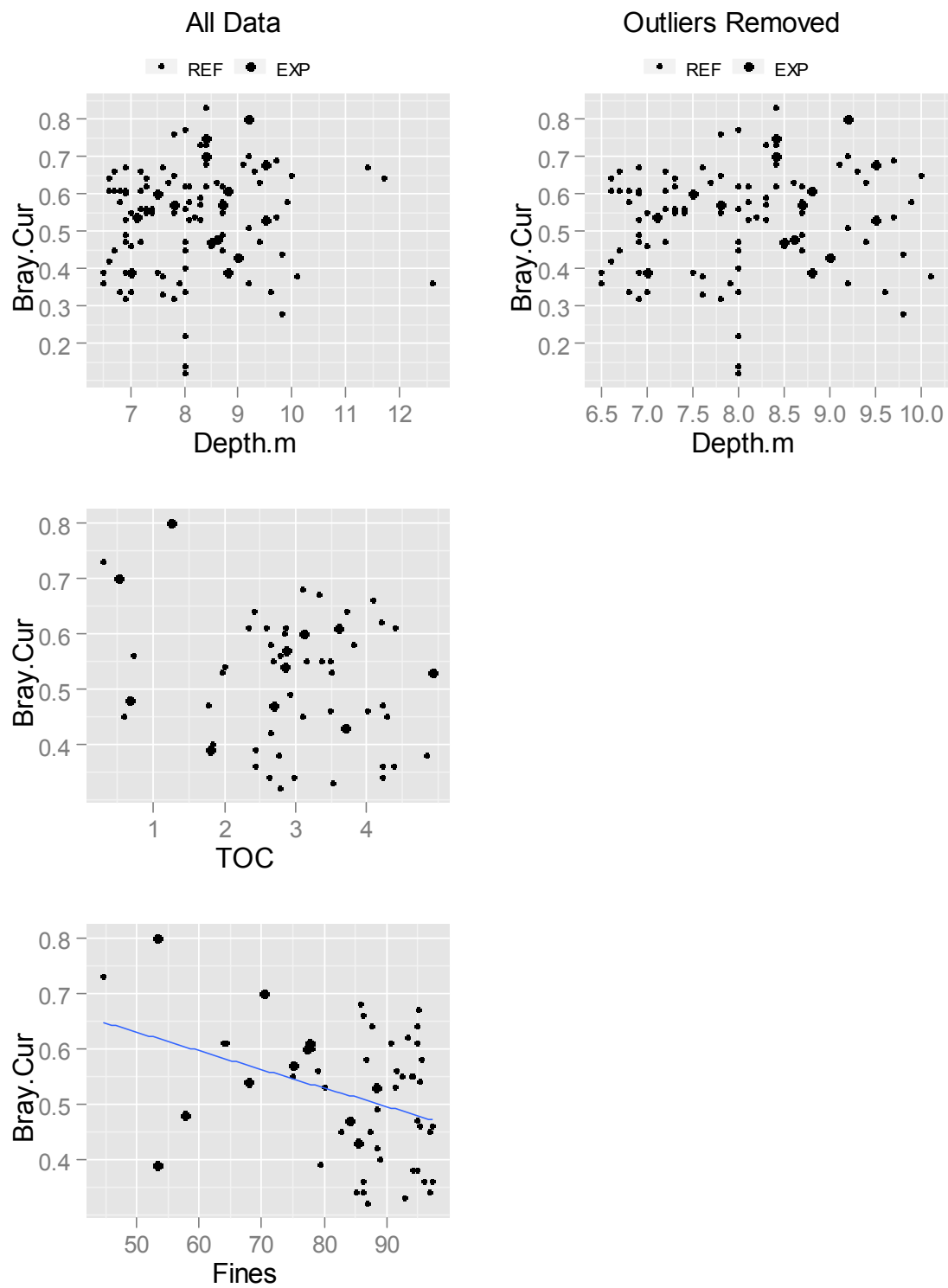
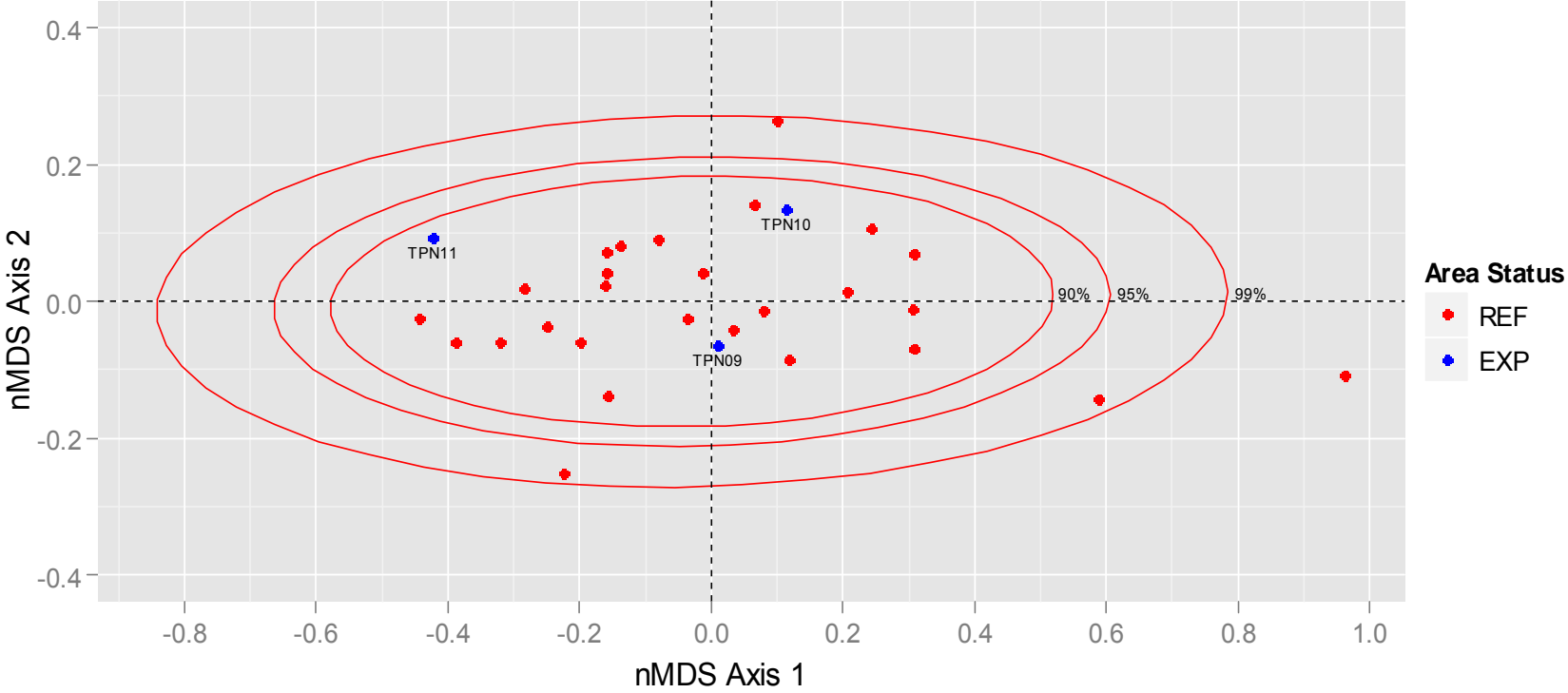


Figure 5.4- 14. Reference condition approach (RCA) results comparing TPN exposure area-year combinations to all CREMP control area-year combinations based on multivariate analysis of density and richness by major taxa group, CREMP Data.



5.5. Weight-of-Evidence Assessment

A weight-of-evidence framework was used to evaluate the extensive analyses conducted in support of this EEM study. The goal of this approach was to critically evaluate the existing data to determine the most likely cause of the observed differences in benthic invertebrate community metrics described in **Section 5.3.3 (Figure 5.5-1)**. The weight-of-evidence approach included the following:

- *Analysis name* – included to clearly differentiate between subtly different statistical analyses that provide complementary insights.
- *Data set description* – describes the program (EEM vs CREMP), years and areas included in the analysis.
- *Effect definition* – describes the specific comparisons being made in the analysis and the critical p-value defining an effect.
- *Significant differences in benthic invertebrate community endpoints* – this column generally provides a synopsis of the actual results, with one or more of: (1) a “SS” or “NS” indicating whether a difference was statistically significant, (2) the direction of response relative to reference (i.e., < or >) and (3) the rough magnitude of response relative to the respective CES (i.e., <, =, >). Other information may also be included as appropriate for the analysis (e.g., relationships with supporting environmental variables described as low, moderate or strong). Shaded cells highlight cases where statistically significant differences were observed.
- *Notes on Supporting Environmental Endpoints* – similar to above.
- *Comments regarding interpretation* – this column provides a narrative regarding key constraints or uncertainties that should be considered when interpreting the overall results.

The weight-of-evidence is presented in **Table 5.5-1**. While significant differences in benthic invertebrate community endpoints were found between reference and exposure areas using the EEM 2011 data, the available evidence from the CREMP data set indicates that the cause of those differences was a confounding gradient in TOC or Fines.

These findings are also consistent with known temporal patterns in effluent exposure (**Figure 5.5-2**). Despite the highest TSS loading occurring in spring 2010, there was no apparent adverse response by the benthic invertebrate community in August 2010. Effluent TSS loadings had been reduced for nearly a year by the time the observed differences occurred. During that same period (i.e., low effluent TSS loadings), however, measured amounts of Fines and TOC were noticeably lower relative to samples from previous years (with assumed cause to be spatial heterogeneity rather than temporal changes¹⁴). While the benthic invertebrate sampling programs target fine sediments, it is not uncommon to encounter areas with coarser substrates in all the study lakes. Consequently, the most likely explanation of the lower TOC and Fines is that such an area was sampled at TPN in 2011.

Finally, these findings are also consistent with those of a long-term study (TSS Effects Assessment Study) looking at the potential effects of TSS releases during dike construction (Azimuth, 2012b). In that study, despite seeing high TSS concentrations in the receiving environment (e.g., extensive areas >10 to 15 mg/L and localized areas near the construction zone in the hundreds of mg/L), adverse effects to the benthic community appeared limited to the areas closest to the dike construction zone. Even then, the expected patterns of response and subsequent recovery were often clouded by natural variability. Based on the TSS EAS results, it is unlikely that the receiving environment TSS concentrations measured to date would result in a “signal” (i.e., response) that can be detected above the “noise” of natural variability.

¹⁴ There are a number of reasons why we would expect the differences observed in TOC and Fines to be natural rather than associated with the dewatering effluent: (1) the particulates in the dewatering effluent are expected to be within the range included in the Fines (i.e., silt and clay fractions). Heavier material would be expected to settle in the treatment plant or very close to the final discharge point; (2) the TPN sampling stations were all much closer to each other than they were to the final discharge point (i.e., in the 8 to 10 m depth range shown in **Figure 3.3-7**). Thus, we would not expect to see the magnitude of change observed in Fines and TOC based on effluent exposure.

Table 5.5- 1. Weight-of-evidence summary for benthic invertebrate community statistical analyses.

Analysis	Data Set Description	Effect Definition	Differences in Benthic Invertebrate Community Endpoints				Notes on Supporting Environmental Endpoints			Comments Regarding Interpretation
			Total Density	Total Richness	Simpson's Evenness	Similarity Index	TOC	FINES	DEPTH	
EEM CI-Based ANOVA (Section 5.3.3)	2011 EEM Data; Pooled Reference (PDL/INUG) Areas and TPN Exposure Area.	Exposure mean different than pooled reference mean (p<0.1).	NS REF>EXP <CES	SS REF>EXP <CES	NS EXP>REF <CES	SS EXP>REF >CES	SS REF>EXP	SS REF>EXP	SS EXP>REF	The pooled CI differences observed are confounded by the gradient in all three supporting variables. However, the data set to too small to accurately characterize the importance of the environmental variables in explaining the observed benthic community differences.
EEM Area-Based ANOVA (Section 5.3.3)	2011 EEM Data; Pooled Reference (PDL/INUG) Areas and TPN Exposure Area.	Exposure mean independently different than both reference means (p<0.1).	NS REF>EXP <CES	NS REF>EXP <CES	NS EXP>REF <CES	SS EXP>REF >CES	NS REF>EXP	SS REF>EXP	NS EXP>REF	The Area-based ANOVA requires TPN to be different from both reference areas. While this is a more stringent definition of "effect", limited inferences can be made about the observed difference in the Similarity Index due to the confounding environmental gradient.
CREMP CI-Based ANOVA (Section 5.4.3)	2011 CREMP Data; Pooled Reference (PDL/INUG + TPS/TEFF) Areas and TPN Exposure Area.	Exposure mean different than pooled reference mean (p<0.1).	NS REF>EXP <CES	NS REF>EXP <CES	--	NS EXP>REF <CES	NS REF>EXP	SS REF>EXP	SS EXP>REF	This data set contains a more extensive set of reference areas. No CI differences were seen in this analysis. Differences were seen in Fines and Depth, after difference in the Similarity Index due to the confounding environmental gradient.
CREMP BACI MODEL (Section 5.4.4)	2006 - 2011 CREMP Data; Looked for adverse effects to benthos related to dewatering discharges to TPN since 2009.	Significant differences in the B-A and C-I interaction term (p<0.1).	NS	NS	--	NS	--	--	--	The BACI model does a better job of taking natural variability in control area-year combinations into consideration. However, it did not consider relationships with environmental variables. Simpson's diversity was significantly higher at TPN for both the short-term and long-term effects.
CREMP Environmental Covariates (Section 5.4.5)	2009 - 2011 CREMP Data; Looked for relationships between benthic endpoints and environmental variables across the five areas.	Not Applicable.	Moderate relationship with TOC Weak with Fines	Moderate relationship with TOC	--	Moderate relationship with Fines	Moderate relationships with density and richness	Moderate relationships with density and richness	No relationships with benthic endpoints	These stronger relationships explain up to 24% of the variability in total density and total richness, and up to 16% in the Bray Curtis similarity index. These results warrant closer attention.
CREMP Analyses with Environmental Covariates (Section 5.4.6)	2009 - 2011 CREMP Data; Tested for CI differences using multiple regression models with environmental variables as covariates.	Significant "effect" factor (p<0.1) after relationships with environmental variables is taken into account.	NS Best model with TOC and Year; CI not important	NS Best model with TOC and Year; CI not	--	NS Best model with Fines; CI not	TOC gradient was most important component in model	Fines was important	--	CI differences were not significant for any benthic invertebrate community endpoint after including an enviromental covariate (TOC or Fines). "Year" was also important, suggesting that certain response patterns may be due to regional climate trends.
CREMP Reference Condition Approach (RCA) (Section 5.4.7)	2006 - 2011 CREMP Data; All control area-yrs used to establish reference, then test TPN yrs since 2009.	Comparison of impact area-yrs to reference envelope (90% probability ellipse).	NS TPN<90%	NS TPN<90%	NS TPN<90%	NS TPN<90%	--	--	--	Control area-yr combinations since 2006 were used to construct a multivariate characterization of reference conditions at the Meadowbank site. TPN-09, TPN-10 and TPN-11 were all situated inside the 90% probability ellipse, suggesting that their benthic community endpoints were within the range of natural conditions.

Notes: "Yes" or "No" refers to statistically significant differences detected; "REF" or "EXP" refer to reference or exposure areas; "CES" = critical effect size (+/- 2 SD); **Shading** = statistically significant difference in endpoint between C-I.

Figure 5.5- 1. Schematic of data analysis for benthic invertebrate community endpoints showing weight-of-evidence.

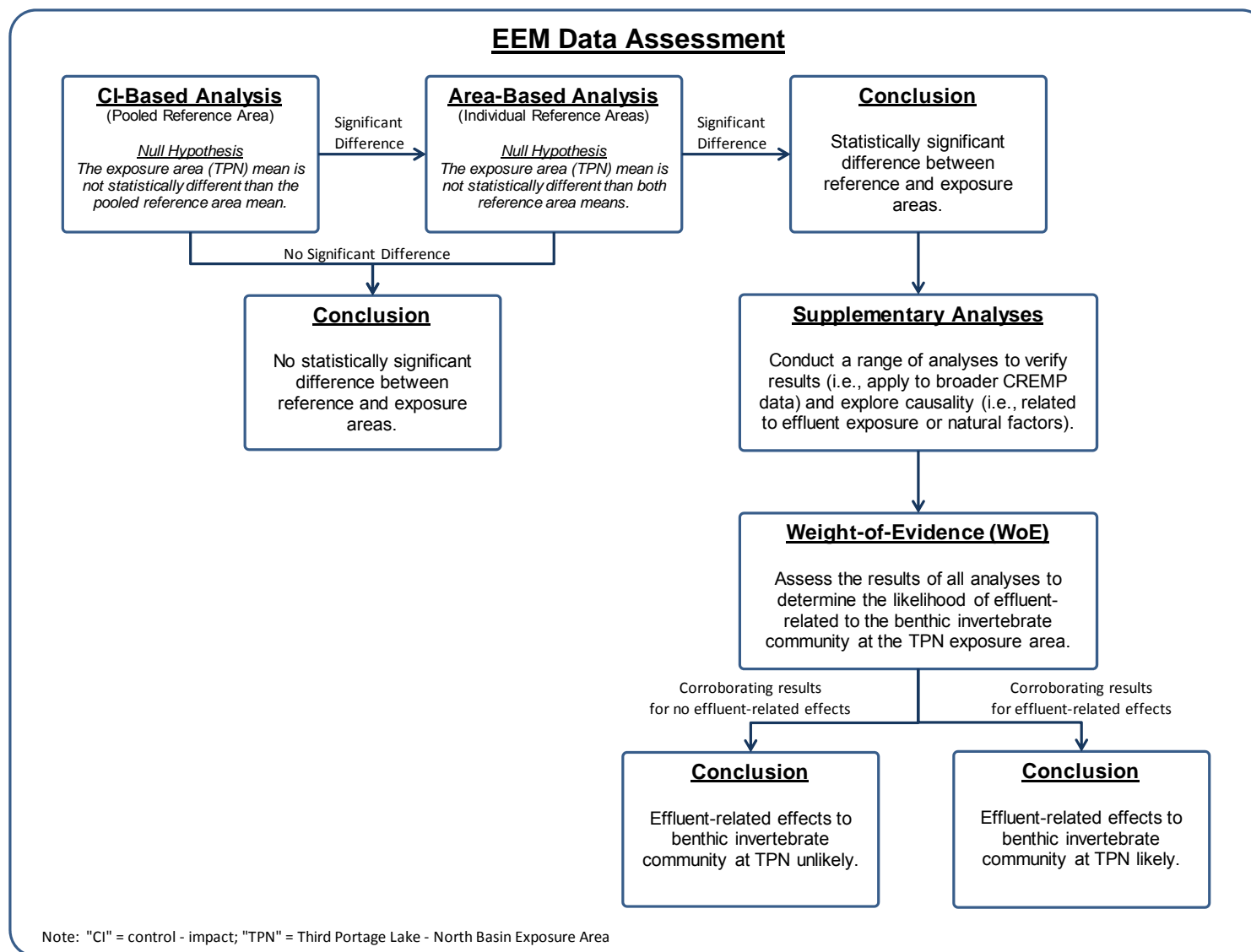
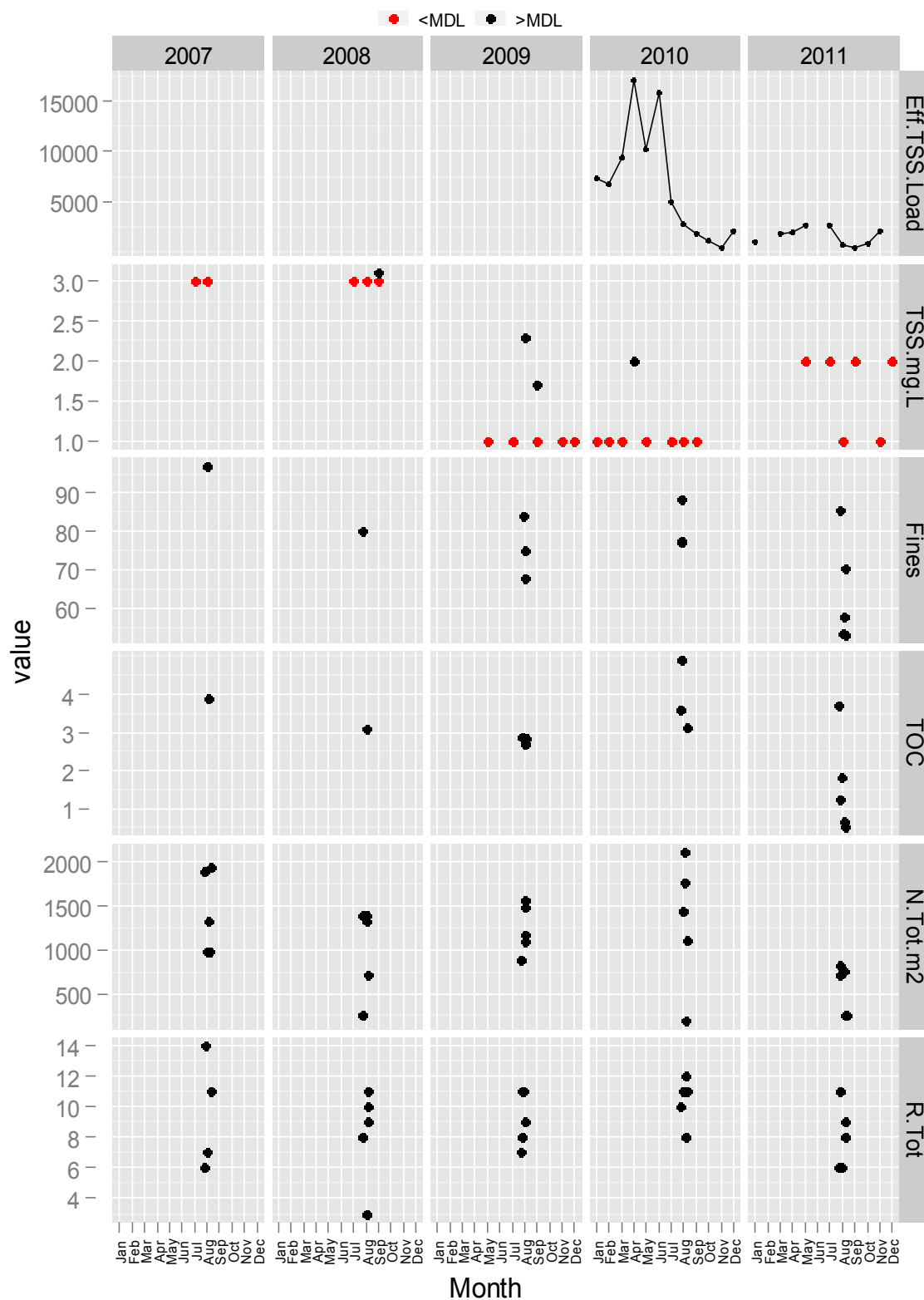


Figure 5.5- 2. Temporal patterns in effluent (TSS loading), receiving environment (TSS, Fines, TOC, Depth) variables. and biological response variables (total density, total richness).



6. CONCLUSIONS FROM BIOLOGICAL MONITORING

6.1. Fish Survey

Details of the data assessment results for the fish survey were presented in **Section 4.3**. A summary of the results by target endpoint is presented in **Table 6.1-1**. No significant differences were identified for any endpoint (see **Section 4.3.6** for details) and confidence in the results for each endpoint was high (see **Section 4.2** [data quality] and **Section 4.3.7** [statistical confidence and power assessment] for details).

These results are consistent with the receiving environment water quality results presented in **Section 3.3**.

Table 6.1- 1. Summary of lake trout data assessment results by target endpoint.

Endpoint Type	Effect Indicator	Non-lethal Endpoints for Meadowbank Fish Survey	Recommended Statistical Procedures	Effect?	Confidence in Result?
Primary	Growth (<i>Energy Use</i>)	Size-at-age (body weight against age)	ANCOVA	No	High
	Condition (<i>Energy Storage</i>)	Body weight against length	ANCOVA	No	High
	Survival	Age frequency distribution	Kolmogorov-Smirnov	No	High
		Length frequency distribution	Kolmogorov-Smirnov	No	High
Supporting	(<i>Energy Use</i>)	Size-at-age (length against age)	ANCOVA	No	High

Notes: Effect based on statistical significance (see **Section 4.3.6**); Confidence in result based on a combination of data quality (see **Section 4.2**) and statistical confidence/power assessment (see **Section 4.3.7**).

6.2. Benthic Invertebrate Community Survey

Details of the data assessment results for the benthic invertebrate community survey were presented in **Sections 5.3 and 5.4**. A weight-of-evidence approach was used to integrate the various studies to evaluate the likelihood of effluent-related effects to the benthic invertebrate community (**Section 5.5**). A simplified summary of the results by target endpoint is presented in **Table 6.2-1**. After accounting for relationships with environmental variables (i.e., TOC or Fines), there were no significant differences were identified for any endpoint (see **Section 5.4.6** for details).

Overall, confidence in these conclusions is high. As examined in **Section 5.3.4**, the study had high power to detect the CES for total density and total richness, and had moderate power ($1-\beta = \sim 0.8$) for evenness and the similarity index. In addition, study data quality objectives were also met (**Section 5.2**).

The conclusion for all benthic invertebrate community endpoints was that there were no effluent-related effects at TPN. These results are consistent with the receiving environment water quality results presented in **Section 3.3**.

Table 6.2- 1. Summary of benthic invertebrate community data assessment results by target endpoint.

Endpoint	Differences between Reference and Exposure Areas? ¹	Relationship with Environmental Variables? ²	Differences between Reference and Exposure Areas after Accounting for Relationship with Environmental Variables? ³	Confidence in result? ⁴
<i>Benthic Community Endpoints</i>				
Total Density (N.Tot.m2)	No	Yes (TOC and Fines)	No	High
Total Richness (R.Tot)	Yes	Yes (TOC)	No	High
Evenness (Simp.E.prop)	No	Not Applicable (see text)	No	High
Similarity Index (Bray.Cur)	Yes	Yes (Fines)	No	High

Notes: ¹ Section 5.3.3; ² Section 5.4.5; ³ Section 5.4.6; ⁴ Sections 5.2 and 5.3.4.

7. SUBSEQUENT BIOLOGICAL MONITORING

As per Division 2, Part 2 of Schedule 5 of the *Metal Mining Effluent Regulations*, the following timing is stipulated for the second cycle of biological monitoring studies:

- *Cycle Two Study Design* – needs to be submitted at least six months prior before undertaking the biological monitoring study (target December 2013).
- *Cycle Two Biological Monitoring Study* – no stipulated timeline, but needs to be completed to allow sufficient time for analysis, interpretation and reporting (target August 2014).
- *Cycle Two Interpretative Report* – submission by July 1, 2015 (i.e., no later than 36 months after the submission of this interpretative report).

7.1. Fish Survey

Based on the results presented herein, we recommend the following considerations for the fish survey component of the Cycle 2 Study Design:

Focus on non-lethal sampling of lake trout only.

As discussed in **Sections 2.3 and 4.3**, efforts to catch round whitefish and Arctic char to support a lethal study were far from successful this past year. In addition to the relatively low abundance of these species compared to lake trout, it is common in Arctic lakes for mature fish to take one or more years off between spawning events to “rest” (as discussed in **Section 4.3.1**). The implication of this is that even more fish need to be caught (and likely sacrificed to determine reproductive status) to support a lethal survey than would be needed at most southern mines.

Use power analysis results to reduce sample size to minimize incidental lake trout mortality.

The power analysis results summarized in **Table 4.3-12** clearly show that sample size can be reduced at all areas for all the endpoints analyzed with ANCOVA (i.e., growth [body weight and length against age] and condition [body weight against length]). The body weight-age endpoint required the highest number of fish per area at 61. A conservative target of 70 to 80 fish should meet the targeted power ($1-\beta = 0.9$) for the EEM-recommended CES values.

Sample size requirements for the age and length frequency distribution endpoints could be estimated if meaningful CES values were available. However, the challenge is that the Kolmogorov-Smirnov test is sensitive to changes in both magnitude (e.g., a change in the overall mean of the variable) and distribution shape (e.g., changes that might not affect the mean, but that would affect the overall distribution). Based on the critical D values for the K-S test (for $\alpha=0.1$, two tails and $n=100$, $D_{0.1(2),100} = 0.120$; for $n=75$, $D_{0.1(2),75} = 0.139$), the absolute difference in critical cumulative relative frequency values is only about 0.02 (or about 2%). Consequently, a reduction in sample size to 70 or 80 fish for this test would not substantially lower its power.

7.2. Benthic Invertebrate Community Survey

Based on the results presented herein, we recommend the following considerations for the benthic invertebrate community survey component of the Cycle 2 Study Design:

Change the number of subsample grabs per station from three to two to be consistent with the long-term CREMP data set.

- This issue was initially addressed in the Study Design document following EEM TGD methods (see **Section 5.4.1** for brief recap); the majority of the data supported collecting two grabs per sample (and the other results were somewhat questionable), but the TAP recommended using the default three subsamples to be consistent with EEM and to provide a more substantive data set with which to address this issue.
- The EEM and CREMP data sets were quantitatively compared using power analysis in an effort to document whether the two-grab data set had less statistical power (**Section 5.4.1**). The results confirmed those in the Study Design (i.e., statistical power of the two-grab data set was virtually the same as the three-grab data set) and support moving to two-grab subsamples in future EEM cycles.
- Not only will consistency with the CREMP reduce sampling effort, sample analysis costs, duplication of statistical analyses, and reporting time, but it will expand the available data temporally (i.e., back to 2006) and spatially (e.g., can include more CREMP control data), resulting in more robust analyses.

8. REFERENCES

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Azimuth. 2011. This reference includes three related documents, as follows:

- 1) Environmental Effects Monitoring (EEM): Cycle 1 Study Design, Meadowbank Division, Nunavut. Report prepared by Azimuth Consulting Group Inc., Vancouver, BC for Environment Canada, Edmonton, AB on behalf of Agnico-Eagle Mines Ltd., Baker Lake, NU. December, 2010.
- 2) Environmental Effects Monitoring (EEM): Cycle 1 Study Design ADDENDUM, Meadowbank Division, Nunavut. Report prepared by Azimuth Consulting Group Inc., Vancouver, BC for Environment Canada, Edmonton, AB on behalf of Agnico-Eagle Mines Ltd., Baker Lake, NU. July, 2011.
- 3) TAP comments on the Meadowbank Gold Mine July 2011 EEM Study Design Addendum and subsequent response email from Azimuth Consulting Group. August, 2011.

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- Krebs, C.J. 1985. Ecology: the experimental analysis of distribution and abundance. 3rd Edition, New York (NY): Harper and Row.
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APPENDICES



APPENDIX A

FISH PERMITS: DFO LICENSE TO FISH FOR SCIENTIFIC PURPOSES





Licence #: S-11/12-1042-NU

Gary Mann
218-2902 West Broadway
Vancouver, BC, CA V6K 2G8

Dear Gary Mann,

Enclosed is your Licence to Fish for Scientific Purposes issued pursuant to Section 52 of the Fishery (General) Regulations.

Failure to comply with any of the conditions specified on the attached licence may result in a contravention of the Fishery (General) Regulations.

Please be advised that this licence only permits those activities stated on your licence. Any other activity may require approval under the Fisheries Act or other legislation. It is the Project Authority's responsibility to obtain any other approvals.

Please ensure that you include the licence number and project title in any future correspondence and that you complete the Summary Harvest Report upon completion of activities under this licence.

Yours truly,

Chris Lewis
Fisheries Management Biologist
Eastern Arctic Area
Central and Arctic Region
Fisheries and Oceans Canada

Enclosure

Date



LICENCE TO FISH FOR SCIENTIFIC PURPOSES

S-11/12-1042-NU

Pursuant to Section 52 of the Fishery (General) Regulations, the Minister of Fisheries and Oceans hereby authorizes the individual(s) listed below to fish for scientific purposes, subject to the conditions specified.

Project Authority: Gary Mann
218-2902 West Broadway
Vancouver, BC, CA V6K 2G8
Azimuth Consulting Group

Other Personnel: Randy Baker
Sue Hertam
Mike Johnson
Laura Henderson
Claire Hrenchuk
Craig Fazakas

Objectives: Agnico-Eagle Mines Ltd. has received a NIRB Project Certificate for its Meadowbank gold project, located 70km north of Baker Lake, Nunavut. Environmental monitoring has been ongoing at this site since 1999. The purpose of the monitoring program is to avoid or mitigate negative impacts from mine activities, and to meet the conditions and commitments of the NIRB Project Certificate and DFO Authorization (NU-03-0109) for the all-weather road and project lake area. Environment Canada notified AEM that the Meadowbank Mine is subject to Metal Mining Effluent Regulations as of December 31, 2009. The focus of this study is to fulfill requirements of Cycle 1 of the EEM Study Design by evaluating the effects of metal mining effluents on fish, fish habitat and the use of fisheries resources.

CONDITIONS

Waters: Waterbodies Listed - X000:
Third Portage Lake – North basin - 65d00'51"N; 96d05'33"W
Inuggugayualik Lake - 65d01'43"N; 96d23'18"W
Pipedream Lake - 65d01'43"N; 96d23'18"W

Water Body: Waterbodies Listed - See Conditions
Point A: 0° 0' N, 0° 0' W

Species: Trout, Lake				Gear: 10 MM Mesh Gillnets and Larger				
Total Weight	Weight Live	Weight Dead	Number Alive	Number Dead	Number Tows	Number Sets	Hours	Minutes
			480	120				

Water Body: Waterbodies Listed - See Conditions
Point A: 0° 0' N, 0° 0' W

Species: Whitefish, Round				Gear: 10 MM Mesh Gillnets and Larger				
Total Weight	Weight Live	Weight Dead	Number Alive	Number Dead	Number Tows	Number Sets	Hours	Minutes
				120				



Water Body: Waterbodies Listed - See Conditions

Point A: 0° 0' N, 0° 0' W

Species: Benthos

Gear: Ponar dredge

Total Weight	Weight Live	Weight Dead	Number Alive	Number Dead	Number Tows	Number Sets	Hours	Minutes
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15

Fishing Period: August 10, 2011 to August 31, 2011

A copy of this licence must be available at the study site and produced at the request of a fishery officer.

Live fish may not be retained unless specified in the conditions of this licence.

The licence holder shall immediately cease fishing when the total fish killed or live sampled reaches any of the maximums set for any of the species listed.

Transportation:

Other approvals/permits may be necessary to collect or transport certain species, such as Marine Mammal Transportation Permits. For marine mammal parts, products and derivatives a Marine Mammal Transportation Licence is required for domestic transport and, for international transport a Canadian CITES Export Permit is also required.

Retention & Disposal of Fish Caught:

Fish not required for the purpose of dead sampling and/or retention **MUST** be returned to the water at the site of capture. Retained fish may be made available to the nearest settlement for domestic consumption or sold commercially within the Territory. Any dead fish for commercial sale beyond the Territory in which it was caught requires authorization under the Fish Inspection Regulations. Disposal of any fish remains must be in accordance with local land use regulations.

Report on Activities:

The Project Authority will submit to the Area Licensing Coordinator, Department of Fisheries and Oceans, within one month of the expiry date, a report stating:

- i) whether or not the field work was conducted; and if conducted
- ii) waterbody location, fishing coordinates, gear types used at each coordinate, numbers or amount of fish (by species) collected and/or marked and the date or period of collection.

A Summary Harvest Report template is provided by the Licensing Coordinator at time of issuance of this licence.

The Project Authority also will provide a copy of any published or public access documents which result from the project. Information supplied will be used for population management purposes by the Department of Fisheries and Oceans and becomes part of the public record.

All documents should be sent to:

Area Licensing Administrator
Fisheries and Oceans Canada
P.O. Box 358
Iqaluit, NU X0A 0H0
Email: XCA-NUpermit@dfo-mpo.gc.ca



Notification of Commencement:

Prior to the commencement of fishing the Project Authority will contact:

Area Licencing Administrator
Fisheries and Oceans Canada
Box 358
Iqaluit, NU X0A 0H0
email: XCA-NUpermit@dfo-mpo.gc.ca

2011.08.29

Eric Kan
Area Director, Eastern Arctic Area
Central and Arctic Region
Fisheries and Oceans Canada

Date

For the Minister of Fisheries and Oceans.

Pursuant to Section 52 of the Fishery (General) Regulations.



Date: August 9, 2011

To: Gary Mann
Azimuth Consulting Group
Vancouver, BC, V5V 3G1

Subject: Animal Use Protocol - Letter of Approval

Dear Gary,

Your 2011 Animal Use Protocol (AUP), number FWI-ACC-2011-057 entitled "Meadowbank Mine: Environmental Effects Monitoring Cycle 1", has been reviewed and approved by the Freshwater Institute Animal Care Committee. This AUP will expire on August 31, 2011.

Keep this signed letter of approval as well as the signed AUP approval form for your records. Please be advised that should there be a need to revise the protocol you are requested to contact the Freshwater Institute Animal Care Committee and obtain approval prior to proceeding.

In addition, you are required to submit a brief report within 30 days of completion of the project outlining the unexpected changes to the protocol, the number of animals used and any unanticipated results or mortalities. The report form is enclosed.

Feel free to contact me if you have any questions or concerns.

Sincerely,

Bernard LeBlanc

FWISL-ACC Chairperson

*Freshwater Institute Science Laboratories Animal Care Committee
Arctic Aquatic Research
Central & Arctic / Région du Centre et de l'Arctique
Fisheries and Oceans Canada / Pêches et Océans Canada
501 University Crescent
Winnipeg, Manitoba R3T 2N6
Phone: 204 983-1327
Fax: 204 984-2403*

Enclosure



Pêches et Océans
Canada

Fisheries and Oceans
Canada



Canada

APPROVAL BY ANIMAL CARE COMMITTEE MEMBERS

AUP#: ACC-2011-057

Date: August 9, 2011

Signatures of ACC Members

Bernard LeBlanc, Chair

Theresa Carmichael

Dr. Ericka Anseeuw D.V.M.

Bob Artes

Cortney Watt

Kerry Wautier

Megan Desai

Interim Approval ☐

Final Approval ☒

**APPROVAL BY THE FWI ANIMAL CARE COMMITTEE IS FOR THE PERIOD STATED ON
YOUR ANIMAL USE PROTOCOL.**



Pêches et Océans
Canada

Fisheries and Oceans
Canada

APPENDIX B

STANDARD OPERATION PROCEDURE (SOP) FOR CREMP WATER & PHYTOPLANKTON SAMPLING



Standard Operating Procedure Meadowbank Project Lakes & Baker Lake CREMP Water & Phytoplankton Sampling

GENERAL:

Project Coordinator:

Maggie McConnell
Azimuth Consulting Group
218-2902 West Broadway
Vancouver, BC, V6K 2G8
Telephone: 604-730-1220
Email: mmcconnell@azimuthgroup.ca

In case of **emergency**, contact Gary Mann (Azimuth telephone number 604-730-1220 or cell phone 604-908-0601).

LOCATION AND TIMING FOR FIELD ACTIVITIES:

Twelve (12) sampling stations have been chosen for water quality monitoring in the Meadowbank project lakes and Baker Lake. Water samples are collected monthly during open water and during ice-cover, except for October and June, when ice conditions are unsafe for sampling. The 12 stations (with their corresponding abbreviation) are:

- Third Portage Lake – North Basin (TPN) **[EEM Sampling Area]**
- Third Portage Lake – East Basin (TPE)
- Second Portage Lake (SP)
- Tehek Lake (TE)
- Inuggugayualik Lake (INUG) **[EEM Sampling Area]**

- Tehek Lake – Far-field (TEFF)
- Third Portage Lake – South Basin (TPS)
- Wally Lake (WAL)
- Pipedream Lake (PDL) **[EEM Sampling Area]**

- Baker Lake – Barge Dock (BBD)
- Baker Lake – Proposed Jetty (BPJ)
- Baker Lake – Akilahaarjuk Point (BAP)

There are two levels of sampling intensity over the course of the year, depending on whether under ice or in open water; during open water all stations are sampled, whereas only the first 5 stations listed above are sampled through-ice. Check the monthly water sampling schedule to confirm which sampled are to be collected where, BEFORE going into the field.

All samples are now collected from **3 m below the water surface**, which is indicated by “-S” at the end of the sample ID (S=Surface). Two target sample locations for each station listed above have been

randomly pre-determined and recorded in MapSource and in the hand-held GPS units (NAD 83). Confirm before going into the field.

Field activities are scheduled roughly once a month for 6 months, including winter ice sampling. Sampling through the ice will take place in May, November, and December. Open water sampling will take place in July, August, and September. Sampling will not be conducted in June and October because of thin ice conditions.

MONTHLY CREMP WATER CHEMISTRY & PHYTOPLANKTON SAMPLING:

1. Prior to leaving camp gather the appropriate type and number of sampling vessels and acid vials for preservation. Prepare appropriate labels for containers, affix them to the appropriate bottle (see below), and wrap label with packing tape. Use the following information:
 - Azimuth company name
 - Station abbreviation (e.g. TPE-36-S, INUG-24-S)
 - Date of sample collection
 - Parameters to be measured from individual bottle (TOC, total metals, etc.)

2. Gather **field collection materials:**

In the boat:

- Field collection data forms, pencils, waterproof markers & clipboard
- GPS unit, batteries
- Diaphragm water pump & 12V battery
- Flexible food-grade silicone tubing (4 meter length and 1 meter length) & weight (& extra C-clamps and cable ties)
- In-line filter (0.45 µm) and a spare
- YSI Professional Plus meter, batteries
- Secchi disk
- Hand held pH meter (if not part of YSI measurements), batteries
- Depth meter, batteries
- Rope
- Sampling gloves
- Field sample bottles & preservatives (per sample) (***this may be different for ALS Winnipeg***):
 - ▶ 2 – 1 L plastic (TSS-low and Conventionals)
 - ▶ 1 – 250 mL amber glass (TKN, Ammonia)
 - ▶ 2 – 125 mL amber glass (TOC & DOC)
 - ▶ 2 – 250 mL plastic (total and dissolved metals)
 - ▶ 1 – 50 mL vial for phytoplankton
 - ▶ 1 vial sulfuric acid
 - ▶ 2 vial hydrochloric acid
 - ▶ 2 vial nitric acid
 - ▶ 1 syringe & magnesium carbonate slurry
 - ▶ 1 syringe & Lugol's solution for phytoplankton
- Extra sample bottles in case of breakage or loss
- QA/QC field duplicate sampling containers & preservatives (same as above).
- Take one set of Travel Blank bottles into the field and transport and treat as other samples. Note that the Travel Blank bottles are not to be opened and no preservatives added.

In camp:

- Hand pump, filters, tweezers, tinfoil and magnesium carbonate for chlorophyll-a
- De-ionized water for rinsing equipment and collected field equipment blank
- Coolers (for storing and shipping samples)
- Ice packs (for shipping samples to laboratories)
- Address labels for coolers
- Chain-of-custody forms
- Large Ziploc bags (for sending chain-of-custody form in cooler)
- Packing tape (for affixing labels to sampling containers & sealing cooler)

The following table lists the specific bottles to be filled, parameters to be measured and preservatives required for each. Affix the labels to the sampling containers and then prior to shipping, wrap packing tape around the labels to ensure a waterproof seal.

(Note: this may be different for ALS Winnipeg)

Sampling Container	Parameters to be Measured	Preservatives to be Added
2 - 1 L plastic	Conventionals*	None
250 mL amber glass	TKN, Ammonia	1 vial of sulfuric acid
250 mL plastic	Total Metals	1 vial of nitric acid
250 mL plastic	Dissolved Metals	1 vial of nitric acid
125 mL amber glass	TOC	1 vial of hydrochloric acid
125 mL amber glass	DOC	1 vial of hydrochloric acid
50 mL vial	Phytoplankton	1/2 mL of Lugol's solution per 50 mL sample

* includes: hardness, conductivity, pH, TDS, TSS-low, turbidity, alkalinity (speciated), orthophosphate and total phosphate, chloride, fluoride, bromide, sulfate, nitrate-nitrogen, nitrite-nitrogen, silicate.

** **do not use filtrate water** for any sample. Use the in-line filters for collecting dissolved metals and dissolved organic carbon samples.

3. For **QAQC** purposes three kinds of samples are required; one set per 10 field samples as follows:

A: One field duplicate is collected for every 10 sample per event (i.e., 10%). All parameters measured in the original sample are measured in the field duplicate. The sampling station is selected randomly from one of the stations, and labeled as station CREMP [month] DUP-1, -2, -3, -4, etc. Prepare the QAQC labels and affix to the sampling containers, as described in step 2.

B: One set of travel blanks are to be carried into the field and treated like the other sampling vessels except that the bottles are not to be opened or anything added to them.

C: One equipment blank will be acquired per every 10 field samples. To collect an equipment blank set up the water sampling equipment as if a routine sample was to be collected except that the incurrent and excurrent hoses are placed into a 4L container of de-ionized water (**use DI water sent from ALS laboratories**). Pump for 2 minutes (just like in the field) to flush site water from the equipment (also attach the filter to flush for 30 seconds). Using a second 4L jug, flush another approx. 1L (this time excurrent hose is placed in sink or bucket) and then use the remaining 2L fill all bottles, except for TSS-low and chlorophyll parameters. Preserve and treat as other samples, including filtering where

necessary. Label as station CREMP [month] EB-1, -2, -3, -4, etc. Finally, fill a new suite of bottles (if available) directly from a third 4L jug to test for any problems with the DI water itself. Label these as CREMP [month] DI-1, -2, -3, -4.

4. Before and during sampling fill in the requested information on the field data form; complete one field data form in its entirety for each sampling station and sampling event. Forms are made of waterproof paper; print all information on the form using a lead pencil or a write-in-the-rain pen.
5. With the aid of a GPS unit, navigate the boat to the sampling station using the UTM coordinates (in NAD 83) provided. Approach the station from downstream of the wind direction. In windy conditions, anchor the boat upstream of the station and drift back; it is not necessary to anchor the boat in calm conditions providing the boat remains in the same position. Do not allow the anchor to drag through the sampling station. Record the **UTM coordinates** on the field data form.
6. Measure **water depth** at the sampling station using the 'Hawkeye' hand-held depth meter (or transom-mounted lowrance). Hold the meter in the water, facing the lake bottom, until the meter measures the depth. Record this information on the field data form. If you are in water that is too shallow (i.e., **must have at least 5 meters depth**), move to deeper water near the assigned station.
7. Measure the light attenuation at the sampling station using the **Secchi disk**. Lower the disk into the water, on the shady side of the boat, so that you can no longer see it. Slowly raise the disk to the point that you can see it and measure this depth using the markings on the disk rope.
8. Measure the pH of the water at the sampling station using the **pH meter** (unless the YSI includes this parameter). Hold the probe portion of the meter in the lake until the meter measures the pH. Record this information on the field data form.
9. Calibrate the YSI probe prior to going into the field; confirm elevation (m) of sampling environment. Check the DO calibration (adjust barometric pressure based on airport data) but also check the DO membrane (it may need to be replaced). At Meadowbank DO readings should be about 8 – 12mg/L; if meter is reading much lower/higher than this, membrane likely needs to be replaced. Keep a calibration log which includes date and time, type of calibration, results, and troubleshooting.
10. Lower the **YSI probe** into the lake to just below the water surface level. Measure the temperature (°C), specific conductance (i.e., temperature corrected; uS/cm) and dissolved oxygen concentration (mg/L) in the water and record on the field data form. Lower the meter to a depth of 1 m and record the field measurements. Allow the concentrations on the meter to stabilize for 10 to 15 seconds before recording the concentrations. Continue recording the field measurements at **1 m depth intervals** until you reach the whole metre mark above the lake bottom (i.e. if the lake depth is 9.3 meters, record field measurements up to a depth of 9 meters). Use the **turbidity probe** in a similar manner to record a turbidity profile.
11. Set up the **water pump** in the boat; attach the tubing to the pump using the C-clamps and attach the 12-V battery. Attach the 4 meter length of tubing to the intake valve, and the 1 meter length to the output valve. Attach the plastic coated ball weight to the end of the 4 meter length of tubing. Lower the 4 meter length of tubing into the water to **3 meters depth** and place the 1 meter length of tubing over the edge of the boat. Run the pump for **2 minutes** to flush the sampling device.

12. For each sampling station, **fill** the required **pre-labeled sampling containers** with water from the 1 meter length of tubing.
13. Dissolved metals and dissolved organic carbon samples are to be collected with an in-line high capacity filter with 0.45 um pore size. After all unfiltered samples have been collected, disconnect the battery from the pump and fix the filter onto the end of the discharge hose. Re-connect the pump and allow the water to discharge and flush through the filter for 15 – 20 seconds. Direct filtered water into the DOC and dissolved metals bottles. Flow from the hose can be controlled by pinching the incurrent end of the tube (**not the excurrent**). Once filtered samples have been collected remove the filter and place into a plastic or zip-loc bag for re-use. In the Meadowbank environment where the amount of suspended solids is typically low, filters can be re-used for up to 10 samples. Remember to use the same filter when collecting equipment blank samples, not a new filter.
14. **Add the specified preservatives** to the appropriate sampling containers (according to the information on the labels and table in step 2), seal and mix thoroughly by turning upside down and then upright a number of times.
15. To collect a phytoplankton sample, add site water from appropriate depth (i.e., 3 m) into the 50 mL vial provided. Make sure that site information is appropriately labeled on the jar. In the field or the lab, add a few drops of Lugol's solution to the sample so that it has the color of weak tea.
16. Rinse all sections of the water filter apparatus with site water.
17. Back in the office, to process the chlorophyll-a sample, use the hand-held pump apparatus and filters. Using the tweezers, place an ashless filter paper on the screen in the water filter apparatus, and, prior to filtering, add 1–2 drops of magnesium carbonate slurry directly onto the filter, then screw the two sections together and attach the hand-held vacuum pump. **Filter 1 L of water** through the water filter apparatus. Wrap the filter paper in a piece of tinfoil, then place the filter in the pre-labeled ziploc bag. Mark on the field collection data sheet the volume of water filtered. In some cases it is not possible to filter up to 1L. In such cases it is critical to note the actual volume filtered. After filtering remove the filter with clean tweezers, place on a piece of tin foil and double wrap. With a sharpie pen, write the appropriate sampling information on a label and stick to the tin foil. Place the folded tinfoil in a zip loc bag and put into the **freezer**.
18. Until ready for shipping, the water samples are stored **chilled** in a refrigerator in camp, if space is available. The filter for chlorophyll-a analysis must be **frozen**; store this bag in a deep freezer in the camp. Bottles should be put in plastic bubble bags prior to storage on ice to protect the labels from water damage. The phytoplankton samples are stored at **room temperature**.
19. If this sampling station is selected as the QAQC **field duplicate**, collect a second set of water samples (repeat step 10), fill the pre-labeled sampling containers, including the phytoplankton vial (repeat step 11-14) and collect a second filtered chlorophyll-a sample (step 16). Record which sampling station the QAQC samples are collected from on the appropriate field data form.
20. Fill out a **chain-of-custody** form for the water samples and filters being sent to **ALS Environmental**. The COC form must be completed carefully and in its entirety to ensure proper analysis. This includes listing all of the specific conventional parameters (see table in step 2), Azimuth and ALS

contact names, and checking off all of the specific boxes for requested analyses. The ALS laboratory quote number must be printed on the COC form to ensure proper billing.

A **digital COC** form is most commonly used; this form can be filled out in advance to ensure accuracy and efficiency and amended in the field as required. Note that using a digital copy of the COC requires printing 2 copies of the document in the field (one for the laboratory, one for Azimuth). Any questions regarding the COC form should be directed to the Azimuth project coordinator – Maggie McConnell. Put the completed COC form in a sealed ziploc plastic bag in a cooler with the water samples.

21. Fill out a **chain-of-custody** form for the phytoplankton samples being sent to **Plankton R Us Inc.**, Winnipeg, MB. Complete all of the required fields and then put the form in a sealed ziploc plastic bag in the cooler with the phytoplankton samples.

PACKAGING & SHIPPING SAMPLES:

1. Ensure all **water samples** are **sealed** securely. Prior to shipping, it is advisable to wrap the label of each sample bottle with clear tape to make sure that the label does not come off during shipping and handling. Dry the water bottle and wrap with tape. **Pack** water sampling containers upright in coolers with ice packs, and packing material, to ensure samples do not spill or break during transport. (Ideal storage and transport temperature is 4°C).
2. Ensure the COC form is enclosed and then seal the cooler(s). **Label the cooler(s)** with the following address:

ALS Environmental 101-8081 Lougheed Hwy. Burnaby, BC, Canada V5A 1W9 Tel: 604-253-4188 Attn: Brent Mack	OR	ALS Environmental 39 – Unit 12, 1329 Niakwa Road East Winnipeg, MB R2J 3T4 Tel: 204-255-9721
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3. Ensure **phytoplankton samples** are **sealed** securely and **pack** in a cooler with packing material to ensure samples do not break during transport. It is not necessary to keep samples cool.
4. Ensure the COC form is enclosed and then seal the cooler. **Label the cooler** with the following address:

Plankton R Us Inc. Dave Findlay 39 Alburg Drive Winnipeg, MB R2N 1M1 Tel: 204-254-7952

5. **Ship** the water **samples** to ALS Environmental as quickly as possible. Ship the phytoplankton samples to Dave Findlay at the end of each month or event.
6. Send completed **COC forms** and **field data forms** to **Azimuth Consulting Group**, attention the project coordinator – Maggie McConnell.

APPENDIX C

**WATER SAMPLING FIELD DATA SHEETS, AUGUST
2011**



Water Sampling - Meadowbank 2011

PROFILES
NOT USED
REVISITED

STATION INFORMATION

Lake Station: 1NUG
 Sample ID: LEEM
 Crew: Morgan & Maggie & Anne
 Date/ Time: Aug 18, 7:30
 Weather Observations: windy SE 25+km
 UTM Coordinates: Easting: 14W 0622830 097
 Northing: 7210816
 Photo #s: - Field DUP collected? x

FIELD MEASUREMENTS

Depth	Temperature	Conductivity	Dissolved Oxygen	pH	Turbidity
units:					
0	13.1	14.8	3.09	7.65	
1	13.1	14.6	3.99	7.69	
2	13.1	14.6	3.88	7.68	
3	13.1	14.6	3.80	7.81	
4	13.1	14.6	3.65	7.80	
5	13.1	14.7	3.53	7.81	
6	13.1	14.6	3.43	7.85	
7	13.1	14.7	3.43	7.43	
8					
9					
10					
11					
12					
13					
14					
15					
16					
17					
18					
19					
20					

Total Water Depth: 7.8
 Secchi Depth: -
 Phytoplankton collected? -
 Field Notes: _____

Chlorophyll Filter Volume Notes? -
 Equipment Blank Collection Notes? -

Water Sampling - Meadowbank 2011

STATION INFORMATION

Lake Station: 1NUG
 Sample ID: 1NUG - EEM
 Crew: Morgan + Myles
 Date/ Time: AUG. 25, 2011 935am
 Weather Observations: _____

UTM Coordinates: Easting: 14W 0622830
 Northing: 7216816

Photo #s: _____ Field DUP collected? _____

FIELD MEASUREMENTS

Depth	Temperature	Conductivity	Dissolved Oxygen	pH	Turbidity
	units: °C	$\mu S/cm^2$	mg/L		
0	12.0	16.22	8.48	7.68	-
1	12.0	16.4	8.53	7.70	-
2	12.0	16.3	8.53	7.76	-
3	12.0	16.3	8.47	7.82	-
4	12.0	16.3	8.56	7.88	-
5	12.0	16.3	8.50	8.00	-
6	12.0	16.3	8.41	8.10	-
7	12.0	16.4	8.33	8.05	-
8					
9					
10					
11					
12					
13					
14					
15					
16					
17					
18					
19					
20					

Total Water Depth: 8.0 m

Secchi Depth: _____

Phytoplankton collected? _____

Field Notes: _____

Chlorophyll Filter Volume Notes? _____

Equipment Blank Collection Notes? _____

TPX-ECM Aug 24

14W0636907 wp 176

7215067

Depth 14.2m

Morgan + maggie

Depth	Temp	Sec	DO	PH
0	10.5	21.9	9.05	7.52
1	10.5	22.0	8.09	7.56
2	10.5	22.0	9.09	7.56
3	10.4	22.3	9.20	7.59
4	10.4	22.5	9.17	7.60
5	10.4	23.0	9.13	7.61
6	10.4	22.7	9.16	7.60
7	10.4	22.5	9.16	7.63
8	10.4	23.3	9.05	7.64
9	10.4	23.2	9.19	7.64
10	10.4	23.2	9.09	7.71
11	10.4	23.1	9.19	7.69
12	10.4	23.7	9.15	7.76
13	10.4	24.0	9.04	7.65

wp 175
14W0637783
7208049 85m

TPS-ECM

DUP Field Blank Yes Yes

Morgan + maggie Aug 24

Temp Sec DO PH

7m	10.7	20.5	9.33	8.43
6	10.6	20.4	9.07	7.80
5	10.7	20.3	9.12	7.94
4	10.7	20.3	9.17	7.92
3	10.7	20.3	9.19	7.92
2	10.7	20.3	9.14	7.89
1	10.7	20.1	9.15	7.89
0	10.7	20.1	9.24	7.86

Note readings sk + at depth.

Water Sampling - Meadowbank 2011

PROFILES.
NOT USED
REVISITED

STATION INFORMATION

Lake Station: Pipedream
Sample ID: PDL - EEM
Crew: Mo'gan + Maggie + Anne.
Date/ Time: Aug 18th 8:20
Weather Observations: Aug 18th 8:20

UTM Coordinates: Easting: 14W0630636 Northing: 7223028 wp 098

Photo #s: — Field DUP collected? Yes *

FIELD MEASUREMENTS

Depth	Temperature	Conductivity	Dissolved Oxygen	pH	Turbidity
units:					
0	11.8	20.4	4.59	7.65	
1	11.8	20.4	4.51	7.59	
2	11.8	20.4	4.44	7.65	
3	11.9	20.4	4.34	7.68	
4	11.9	20.5	4.31	7.47	
5	11.9	20.4	4.30	7.45	
6					
7					
8					
9					
10					
11					
12					
13					
14					
15					
16					
17					
18					
19					
20					

Total Water Depth: 6.6
Secchi Depth: —
Phytoplankton collected? —
Field Notes: —

Chlorophyll Filter Volume Notes? —
Equipment Blank Collection Notes? —

Water Sampling - Meadowbank 2011

STATION INFORMATION

Lake Station: PDL
 Sample ID: PDL - EEm
 Crew: Morgan + Maggie
 Date/ Time: Aug. 25, 2011 9am
 Weather Observations: _____

UTM Coordinates: Easting: 14W 0630636 → Same as before
 Northing: 722 3028

Photo #s: X Field DUP collected? X

FIELD MEASUREMENTS

Depth	Temperature	Conductivity	Dissolved Oxygen	pH	Turbidity
	units: <u>'C</u>	<u>$\mu S/cm^2$</u>	<u>mg/L</u>		
0	<u>11.4</u>	<u>21.8</u>	<u>8.80</u>	<u>8.15</u>	<u>—</u>
1	<u>11.4</u>	<u>21.8</u>	<u>8.80</u>	<u>8.22</u>	<u>—</u>
2	<u>11.4</u>	<u>21.8</u>	<u>8.80</u>	<u>8.26</u>	<u>—</u>
3	<u>11.4</u>	<u>21.8</u>	<u>8.80</u>	<u>8.32</u>	<u>—</u>
4	<u>11.4</u>	<u>21.9</u>	<u>8.75</u>	<u>8.40</u>	<u>—</u>
5	<u>11.4</u>	<u>21.9</u>	<u>8.70</u>	<u>8.52</u>	<u>—</u>
6	<u>11.4</u>	<u>22.1</u>	<u>8.70</u>	<u>8.51</u>	<u>—</u>
7					
8					
9					
10					
11					
12					
13					
14					
15					
16					
17					
18					
19					
20					

Total Water Depth: 6.6
 Secchi Depth: —
 Phytoplankton collected? —
 Field Notes: _____

Chlorophyll Filter Volume Notes? —
 Equipment Blank Collection Notes? —

APPENDIX D

STANDARD OPERATION PROCEDURE (SOP) FOR CREMP SEDIMENT & BENTHOS SAMPLING



Standard Operating Procedure

Meadowbank Project Lakes & Baker Lake

CREMP Sediment & Benthos Sampling

GENERAL:

Project Coordinator:

Maggie McConnell
Azimuth Consulting Group
218-2902 West Broadway
Vancouver, BC, V6K 2G8
Telephone: 604-730-1220
Email: mmcconnell@azimuthgroup.ca

In case of **emergency**, contact Gary Mann (Azimuth telephone number 604-730-1220 or cell phone 604-908-0601).

LOCATION AND TIMING FOR FIELD ACTIVITIES:

Twelve (12) sampling stations have been chosen for benthos and sediment quality monitoring in the Meadowbank project lakes. These stations (with their corresponding abbreviation) are:

- Third Portage Lake – North Basin (TPN) [**EEM Sampling Area**]
- Third Portage Lake – East Basin (TPE)
- Second Portage Lake (SP)
- Tehek Lake (TE)
- Inuggugayualik Lake (INUG) [**EEM Sampling Area**]

- Tehek Lake – Far-field (TEFF)
- Third Portage Lake – South Basin (TPS)
- Wally Lake (WAL)
- Pipedream Lake (PDL) [**EEM Sampling Area**]

- Baker Lake – Barge Dock (BBD)
- Baker Lake – Proposed Jetty (BPJ)
- Baker Lake – Akilahaarjuk Point (BAP)

Field activities are scheduled for once per year, in **mid/late August**. The **target water depth** at each sampling station is approximately **8 meters +/- 1.5 m**; Wally Lake is the exception, with a total water depth of approximately 6 meters (target water depth is 5 to 6 meters).

BENTHOS & SEDIMENT CHEMISTRY SAMPLING:

1. Gather field collection materials:

In the boat:

- Field collection data forms, waterproof paper, pencils, waterproof markers & clipboard

- GPS unit, batteries
- Depth meter, batteries
- pH meter, batteries
- Rope
- Petite Ponar grab and rope
- 500 micron sieve bag
- 2 stainless steel bowls
- 2 stainless steel spoons
- Liquinox detergent and dish cleaning brush
- Plastic squirt bottle
- Sampling gloves
- Safety glasses
- Field sample jars & preservatives (per sampling station):
 - ▶ 8 – 125 mL glass jars (sediment samples : 2 jars for first 3 reps; 1 jar for last 2 reps, except for EEM where 2 jars for all 5 reps)
 - ▶ 5 – 500 mL plastic jars (benthos: 1 jar for each rep)
- QA/QC field duplicate sediment jars
- Ashless filter paper & tweezers; 1-125 mL glass jar

In camp:

- Formalin (10% of pure Formaldehyde)
- Labels for sampling containers
- Coolers, action packers (for storing and shipping samples)
- Ice packs (for shipping sediment samples to lab)
- Address labels for coolers
- Chain-of-custody forms
- Large Ziploc bags (for sending chain-of-custody form in coolers)
- Electrical tape (for sealing benthos jars)
- Packing tape (for affixing labels to sediment sample containers & sealing coolers)

2. Before going into the field, **label the lids** of all sampling containers using a permanent waterproof marker. After sampling, prepare appropriate labels for containers and affix them when bottles are dry enough to stick to. Use the following information:

- Azimuth company name
- Station abbreviation (e.g. TPE-1, INUG-3)
- Date of sample collection
- Parameters to be measured from individual jar (1 x 125 mL – total metals, pH, moisture, PAHs, LEPHs & HEPHs, Mineral Oil & Grease; 1 x 125 mL – grain size (PSA), TOC)

Affix the labels to the sediment jars and then wrap packing tape around the labels to ensure a waterproof seal.

For the **benthos containers**, print the following information directly onto both the jar and jar lid using a permanent waterproof marker:

- Azimuth company name
- Station abbreviation (e.g. TPE, INUG) and replicate number (e.g. TPE-1, TPE-2); there are a total of 5 replicates per sampling station
- Date of sample collection

Prepare **internal labels** for each of the benthos containers. On a small piece of waterproof paper, write, using a lead pencil, the station abbreviation and replicate number (e.g. TPE-1). If no waterproof paper is available, use regular paper. Store the labels in their corresponding sampling container.

3. For **QAQC** purposes, sediment samples are collected in duplicate from one station every sampling event. All parameters measured in the original sample are measured in the field duplicate. The sampling station is selected randomly from one of the ten stations, and labeled as station DUP. Prepare the QAQC labels and affix to the sediment jars, as described in step 2. Label one new 125 mL glass jar with the Azimuth company name, date, QAQC filter and total metals.
4. A 100% formalin solution is equivalent to a solution of 37% formaldehyde. The **target formalin concentration** in each of the sampling containers is 10%. A neutral buffered formalin solution is achieved by adding a sufficient amount of calcium carbonate powder or pellets to render the solution pH neutral (pH = 7.0). Borax powder may be substituted for calcium carbonate powder if necessary.

Transport Canada allows the free transport of formalin at concentrations less than 25% formaldehyde. Consequently, the formalin transported up to Meadowbank will be diluted in half (18.5% formaldehyde / 50% formalin solution).

To **prepare the neutral buffered formalin**, add a small amount of calcium carbonate powder or pellets to the 50% formalin solution, seal the container and shake until mixed. Check the pH of the solution using the pH pen. Continue adding the powder/pellets until the pH of the solution reaches approximately 7.0. Store at room temperature until ready to use. Only prepare the required volume of neutral buffered formalin for that sampling event. Buffered formalin will not store for long periods of time.

Follow all **safety precautions** when preparing the formalin solution. Formalin is a carcinogen and irritant. Wear sampling gloves and safety glasses when mixing the solution and prepare the solution in a well ventilated area.

5. Before and during the benthos and sediment sampling fill in the requested information on the **field data form**; complete one field data form in its entirety for each sampling station and sampling event. Forms are made of waterproof paper; **print** all information on the form using a **lead pencil** or write-in-the-rain pen.
6. With the aid of a GPS unit, **navigate the boat** to the sampling station using the UTM coordinates (in NAD 83) provided. Approach the station from downstream of the wind direction. In windy conditions, anchor the boat upstream of the station and drift back; it is not necessary to anchor the boat in calm conditions providing the boat remains within a 50 meter radius of the position. Do not allow the anchor to drag through the sampling station. Record the UTM coordinates on the field data form.
7. Measure the **water depth** at the sampling station using the 'Hawkeye' hand-held depth meter (or transom-mounted lowrance). Hold the meter in the water, facing the lake bottom, until the meter measures the depth. Record this information on the field data form Ensure sample depth is within the target (8 meters +/- 1.5 m, or 5-6 meters for Wally).

8. Begin collecting the benthos samples. Collecting the sediment first would disturb the benthic community.
9. Ensure the rope is securely attached to the **Ponar**. Rinse the Ponar grab, stainless steel bowl and spoon with lake water. **Wash** each of these items with liquinox soap by scrubbing with the dish cleaning brush and then thoroughly rinse with lake water. Put aside the stainless steel bowl and spoon until later (step 18).
10. Lower the **Ponar** to within 1 meter of the bottom of the lake. Lower the Ponar very slowly over the last meter and allow the rope to go slack. Raise the Ponar to the edge of the boat and check the grab for **acceptability**. The grab is acceptable if the sample:
 - does not contain large foreign objects;
 - has adequate penetration depth (i.e., 10-15 centimeters);
 - is not overfilled (sediment surface must not be touching the top of the Ponar);
 - did not leak (there is overlying water present in Ponar); and
 - is undisturbed (sediment surface relatively flat).Once the grab is deemed acceptable, open the Ponar jaws and drop the sample into a stainless steel bowl. Rinse the ponar with squirt bottles to make sure all of the material is in the bowl. Gently pour the contents of the bowl into the 500 micron sieve bag.
11. **Sieve the sample** in the lake water until only the benthic organisms and coarse materials remain. Care must be taken to ensure the benthic organisms are not damaged or crushed. Do not disturb the sample to the point that it is splashing out of the sieve. Do not forcibly push materials through the sieve; gently break apart any small clay balls. Rinse off any pieces of larger plant material or rocks in the sample and discard.
12. **Flush the remaining sample** in the bottom of the sieve into the pre-labeled plastic sampling container (i.e. station-1 jar). A plastic squirt bottle filled with lake water is useful for this purpose.
13. **Repeat (twice for EEM) steps 10-12**, flushing the sample into the same pre-labeled plastic sampling container (i.e., station-1 jar). Ensure the sample is collected in an area not previously disturbed by the Ponar. The two (three for EEM) independent grabs (per replicate) are composited to increase the surface area sampled.
14. **Rinse the sieve** bag to clear out any debris in the screen. To rinse, hold the sieve upside down and raise and lower the sieve into the water.
15. **Repeat steps 10-14** four more times; there must be a separation of **20 meters** or more from other replicate stations. Record the depth and GPS coordinates of each replicate station on the field data form. Put the samples from each replicate in pre-labeled station replicate jars 2 through 5. In total, 10 Ponar grabs will be collected for benthos collection, two grabs per replicate.
16. Ensure internal labels are in each sample container. Shake the formalin to ensure all of the calcium carbonate powder is in solution. **Add** a sufficient volume of **formalin** to each sampling container to make a corresponding formalin solution of approximately 10%. Volumes of formalin are added by 'eye' (for a 10% solution, a ratio of 4 parts water and 1 part 50% formalin solution). Overall, there must be enough liquid in the jar to cover the entire sample. Seal the sample container securely and

gently roll the container to mix the sample and formalin solution. Do not shake the sample container; this will crush the benthic organisms inside.

17. Prior to collecting the next REP, clean ponar and both bowls with liquinox and scrub brush, rinse with lake water. Begin collecting the sediment samples. Lower the **Ponar** to within 1 meter of the bottom of the lake, in an area not previously disturbed by the Ponar. Lower the Ponar very slowly over the last meter and allow the rope to go slack. Raise the Ponar to the edge of the boat and check the grab for **acceptability** (see step 10 for criteria).
18. Once the grab is deemed acceptable, open the top of the Ponar and remove any overlying water. Using the pre-cleaned stainless steel spoon, scoop out the **top 3-5 centimeters** of **sediment** and place in the pre-cleaned stainless steel bowl. Empty the remainder of the grab sample into a bucket in the boat, not directly into the lake, to ensure the area is not disturbed.
19. **Repeat steps 17 and 18** one more time, placing the sediment into the bowl with the other sediment sample(s).
20. **Homogenize** the sediment samples in the stainless steel bowl (by stirring with the spoon) until the sediment is thoroughly mixed. Scoop the sediment into pre-labeled sediment sampling containers. **Fill the jars** to the top and seal securely.
21. If this station is selected as the QAQC **field duplicate**, using the tweezers and a set of clean sampling gloves, **swipe** the stainless steel bowl and spoon with one piece of ashless **filter paper** and store in the pre-labeled 125 mL glass jar. Collect the duplicate sediment sample from the same sediment collected in steps 17-20. Fill the sampling containers labeled as station DUP. Record that the QAQC samples were collected from this sampling station on the field data form.
22. **Complete the field data form**, including a description of the sediment (grain size, consistency, colour, presence of biota, sheen, unusual appearance) and the sampling effort (equipment failure, control of vertical descent of sampler) required to collect the benthos and sediment samples.
23. **Rinse** out the Ponar, stainless steel bowl and spoon with lake water. Dump the sediment and water from the plastic bin into the lake.
24. Until ready for shipping, store the sediment samples and QAQC filter paper **chilled** (on ice) in a cooler or in a refrigerator in camp, if space is available. The sediment sampling containers may be put in plastic bags prior to storage on ice to further protect the labels from water damage. Benthos samples are stored in a cooler or action packer at **room temperature**.
25. Fill out a **chain-of-custody** form for the sediment samples being sent to **ALS Environmental**. The COC form must be completed carefully and in its entirety to ensure proper analysis. This includes listing all of the specific parameters to be analyzed (see step 2), Azimuth and ALS contact names, and checking off all of the specific boxes for requested analyses. The ALS laboratory quote number must be printed on the COC form to ensure proper billing.

A **digital COC** form is most commonly used; this form can be filled out in advance to ensure accuracy and efficiency and amended in the field as required. However, using a digital copy of the COC requires printing 2 copies of the document in the field (one for the laboratory, one for Azimuth). Any

questions regarding the COC form should be directed to the Azimuth project coordinator – Maggie McConnell. Put the completed COC form in a sealed ziploc plastic bag in the cooler with the samples.

26. Fill out a **chain-of-custody** form for the benthos samples being sent to **Zaranko Environmental Assessment Services (ZEAS)**. Complete all of the required fields and then put the form in a sealed ziploc plastic bag in the cooler with the benthos samples.

PACKAGING & SHIPPING SAMPLES:

1. Ensure all **sediment samples** are **sealed** securely. **Pack** sediment sampling containers upright in a cooler with ice packs, and packing material, to ensure containers do not break during transport. (Ideal storage and transport temperature is 4°C).
2. Ensure the COC form is enclosed and then seal the cooler(s). **Label the cooler(s)** with the following address:

ALS Environmental
101-8081 Lougheed Hwy.
Burnaby, BC, Canada
V5A 1W9
Tel: 604-253-4188
Attention: Brent Mack

3. Ensure **benthos samples** are **sealed** securely. Wrap electrical tape around the edge of the lids to ensure a tight seal. **Pack** benthos sampling containers upright in a cooler or action packer; ensure the cooler/action packer is well packed so the jars are not able to move around.
4. Ensure the COC form is enclosed and then seal the cooler(s). **Label the cooler(s)** with the following address:

Zaranko Environmental Assessment Services (ZEAS)
36 McCutcheon Avenue
P.O. Box 1045
Nobleton, ON
L0G 1N0
Tel: 905-859-7976

5. **Ship** the sediment **samples** to ALS Environmental as quickly as possible. Ship the benthos samples to ZEAS when convenient. Coordinate shipping with the camp manager.
6. Send completed **COC forms** and **field data forms** to **Azimuth** Consulting Group, attention the project coordinator – Maggie McConnell.

APPENDIX E

**SEDIMENT & BENTHOS SAMPLING FIELD DATA
SHEETS, AUGUST 2011**



Benthos and Sediment Sampling - Meadowbank 2011

STATION INFORMATION

Station ID: NAWA

Crew: Morgan, Maggie + Gary?

Date/ Time: Aug 14 09:45

Weather Observations (wind/waves): Wind S N 20 overcast

Locations for BENTHOS (2 grabs composited per rep; >20 m separation b/w reps)

Name	Depth (m)	Easting	Northing	Waypoint #
Rep1	6.9	14W0622816	7216850	030
Rep2	6.8	14W0622784	7216864	031
Rep3 may have three grabs	7.6	14W0622740	7216834	032
Rep4	8.5	14W0622711	7216786	033
Rep5	9.2	14W0622694	7216769	034

Locations for SED. CHEMISTRY (2-3 grabs composited per rep; collect top 3-5 cm, post benthos)

Name	Or same as above? Y / N	
Rep1 + A	Yes	Full Suite
Rep2 + A	Yes	Full Suite
Rep3 + A	Yes	Full Suite
Rep4 + A	Yes	TOC & PSA
Rep5 + A	Yes	TOC & PSA

Photo #s: _____

DUPS?: Rep 1

Filter Swipes?: Rep 1

FIELD NOTES

Sediment Characteristics:

(Grain size, consistency, colour, odour, biota, sheen or unusual appearance)

Sampling Effort (Equipment failure, control of vertical descent of sampler):

More than 500 mL container collected for any benthic sample?

BENTHOS / SEDIMENT SAMPLE CHECKLIST (per REP)

Jar Type	Parameters	Preservation / Storage
1 x 125 mL glass	Metals (incl. Al, Ph), pH, moisture, PAHs, LEPHs&HEPHs, MO&G	none / keep cool
1 x 125 mL glass	TOC & PSA	none / keep cool
1 x 500 mL plastic	Benthos Taxonomy	10% buffered formalin solution

Benthos and Sediment Sampling - Meadowbank 2011

STATION INFORMATION

Station ID: PDL
 Crew: MLM, MF
 Date/ Time: Aug. 13, 2011
 Weather Observations (wind/waves): sun + clouds 10 km N wind.

Locations for BENTHOS (2 grabs composited per rep; >20 m separation b/w reps)

Name	Depth (m)	Easting	Northing	Waypoint #
Rep1 + A	6.5	14W0630554	7222992	013
Rep2 + A	6.8	14W0630607	7223035	014
Rep3 + A	6.9	14W0630711	7222711	015
Rep4 + A	7.6	14W0630879	7223053	016
Rep5 + A	6.7	14W0630596	7223017	017

Locations for SED. CHEMISTRY (2-3 grabs composited per rep; collect top 3-5 cm, post benthos)

Name	Or same as above? Y/N	Full Suite
Rep1	YES	Full Suite
Rep2	YES	Full Suite
Rep3	YES	Full Suite
Rep4	YES	TOC & PSA Full
Rep5	YES	TOC & PSA Full

EEM station

Photo #s:

DUPS?: X

Filter Swipes?: X

FIELD NOTES

Sediment Characteristics:

(Grain size, consistency, colour, odour, biota, sheen or unusual appearance)

Rep 4 - 1st grab lots of clay

Sampling Effort (Equipment failure, control of vertical descent of sampler):

More than 500 mL container collected for any benthic sample?

BENTHOS / SEDIMENT SAMPLE CHECKLIST (per REP)

Jar Type	Parameters	Preservation / Storage
1 x 125 mL glass	Metals (incl. Al, Ph), pH, moisture, PAHs, LEPHs&HEPHs, MO&G	none / keep cool
1 x 125 mL glass	TOC & PSA	none / keep cool
1 x 500 mL plastic	Benthos Taxonomy	10% buffered formalin solution

Benthos and Sediment Sampling - Meadowbank 2011

STATION INFORMATION

Station ID: TPN
 Crew: Tripod
 Date/ Time: Aug. 19, 2011 10:00
 Weather Observations (wind/waves): Winds 20 SE
rainy

Locations for BENTHOS (2 grabs composited per rep; >20 m separation b/w reps)

Name	Depth (m)	Easting	Northing	Waypoint #
Rep1 +A	7.0m	14W0636289	7215547	102
Rep2 +A	9.0m	14W0636285	7215525	108
Rep3 +A	7.2m	14W0636417	7215511	109
Rep4 +A	8.6m	14W0636514	7215438	110
Rep5 +A	8.4m	14W0636607	7215409	111

Locations for SED. CHEMISTRY (2-3 grabs composited per rep; collect top 3-5 cm, post benthos)

Name	Or same as above? Y / N	
Rep1	yes.	Full Suite
Rep2	yes.	Full Suite
Rep3	yes.	Full Suite
Rep4	yes.	TOC & PSA
Rep5	yes.	TOC & PSA

EEM Stn.
Full
Full.

Photo #s: _____

DUPS?: _____

Filter Swipes?: _____

FIELD NOTES

Sediment Characteristics:

(Grain size, consistency, colour, odour, biota, sheen or unusual appearance)

GRABS were good at R1 + R2 but much more difficult after that

Sampling Effort (Equipment failure, control of vertical descent of sampler): _____

More than 500 mL container collected for any benthic sample? _____

BENTHOS / SEDIMENT SAMPLE CHECKLIST (per REP)

Jar Type	Parameters	Preservation / Storage
1 x 125 mL glass	Metals (incl. Al, Ph), pH, moisture, PAHs, LEPHs&HEPHs, MO&G	none / keep cool
1 x 125 mL glass	TOC & PSA	none / keep cool
1 x 500 mL plastic	Benthos Taxonomy	10% buffered formalin solution

APPENDIX F

**STANDARD OPERATION PROCEDURE (SOP) FOR
EEM FISH SAMPLING**



Standard Operating Procedure

Meadowbank Project Lakes

EEM Fish Sampling

GENERAL:

Project Coordinator:

Gary Mann/Randy Baker
Azimuth Consulting Group
218-2902 West Broadway
Vancouver, BC, V6K 2G8
Telephone: 604-730-1220; Fax: 604-739-8511
Email: gmann@azimuthgroup.ca; rbaker@azimuthgroup.ca

Crew Leader: Sue Hertam, North/South Consultants (on site)

In case of **emergency** contact Azimuth Office in Vancouver (604-730-1220)

LOCATION AND TIMING FOR FIELD ACTIVITIES:

Three lakes have been chosen for fish samplings at the Meadowbank Mine for the EEM cycle one study. These lakes (with their corresponding abbreviation) are:

- Third Portage Lake North Basin (TPN): Exposure Area
- Inuggugayualik Lake (INUG): Reference Area
- Pipedream Lake (PDL): Reference Area

Sampling areas within each lake can be viewed in **Figure 1**.

Field activities are scheduled for once per year, in **August to early September**, with a **minimum time allocation of 1 week sampling per lake** (or until sufficient fish numbers are captured). According to EEM guidance, if longer than a week is required to collect all fish samples at all lakes (this is anticipated at Meadowbank as 3 weeks have been allocated to this study), then samples at the reference areas should be collected before and after the exposure area.

FISH SAMPLING:

1. Gather field collection materials:

In the boat:

- Field collection data forms, waterproof paper, pencils, waterproof markers & clipboard
- Gill nets. "Standard" panels are (mm [in]): 126 [5], 102 [4], 76 [3], 51 [2], 38 [1.5], and 25 [1]. Typically deployed as indexed gang (1 of each panel). Panels are 22.7 m long and 1.8 m deep. Other smaller mesh sizes are also available. *Crew leader has flexibility to alter net deployment to meet project targets and minimize incidental catch.*
- Depth meter, batteries; GSP unit and batteries
- Thermometer
- Camera
- Two scales, one for big fish (10 kg +/- 25 g), one for smaller fish (4 kg max +/- 10 g).

- Fin clippers
- Scale collection envelopes for fin rays
- Floy tags/gun
- Cooler(s) and ice packs – one per boat
- Boat safety kit

In camp:

- Length board
- Scales (high range OHAUS Ranger RD60LS [± 2 g] for large fish [>200 g]; low range [± 0.01 g] for small fish [<200 g] and organs)
- Lab data forms
- Camera
- Collection envelopes for otoliths and fin rays
- Dissection utensils (scalpel, forceps, tweezers, etc.) and sampling gloves

Before going into the field, **double check that all gear is present, and that nets of various sizes are available. Make sure you have all field forms needed (i.e. partially filled out forms awaiting net-lifting and fish-catch data).**

General photographs of each location should be taken to document the areas.

Once in the Field

2. **Navigate the boat** to the desired sampling area within the allowable sampling area of the lake (see **Figure 1**). Record air and water temperature daily.
3. Once at location and the nets are organized and ready for deployment, fill out the **field data sheet**. Record on the field data sheet: the set ID, set date, set time, recorder, gang information (mesh sizes etc.), mesh sizes and any other comments. Record the “start” and “end” UTM location (Northing, Easting, Zone in NAD83) using a GPS unit, and water depth (m), using a depth meter. Take a picture of the sampling location. Forms are made of waterproof paper; **print** all information on the form using a **lead pencil** or write-in-the-rain pen.

Retrieving Nets

4. Initial day set durations should be approximately 2 – 3 hrs (duration may be lengthened by Crew Leader after discussion with Coordinator). Record all lift information and fish data on the same form that was filled out when the nets were set. Be sure to be clear on the “Lift Date” and “Lift Time”, as over-night sets would have a different “Set Date”.

Recording Fish Data

5. During net retrieval, record the number and size (FL mm) of each species per panel (mesh size) in the field data sheets. *Do not* inter-mingle fish from different net panels. *All* captured fish are retained for either non-lethal or lethal sampling, including non-target species. See **Table 1** for an overview of target numbers and required parameters for the EEM fish study.
6. **Scale:** Use the appropriate scale for the size of fish captured, i.e. a small fish should be weighed on a finer scale. **Scales must be calibrated before initial use and checked each day (record in log).** Check

that the scale is reading zero prior to measuring *each* fish. For more information on required precision of measurements see **Table 2**.

7. **Establish a person as “measurer” and a person as “recorder”.** This will maximize consistency throughout the study. The recorder should read back each recorded value to the measurer as a verification of accuracy. **Ensure all information is collected each set.**
8. **All fish:** ensure that **field data sheet** fields for species, sample ID and mesh size are filled in.
9. **Lake trout (non-lethal):** Lake trout are to be non-lethally sampled and should be handled quickly but carefully. Collect a wide range of sizes of fish (see historic range, **Figure 2**). Gender of fish is not a factor. Take photos of representative fish. For all lake trout the following parameters should be collected in the field:
 - Measure the length (fork [mm; accuracy to nearest mm]) and weight (g; accuracy to 1% of body weight) of each fish.
 - Record the external DELTs. This includes abnormalities of body form, body surface, fins, eyes, lesions, tumors, scars etc. Mark any of these on the fish diagram sheet (**Figure 3** for example). Take additional pictures as necessary.
 - Using fin-clippers, take the left pectoral fin clip (first 4 rays) and place in a paper scale envelope. Label the bag immediately with the Set ID, Date, and Fish ID.
 - Tag the fish with a Floy Tag, and record the Tag # in the field data sheet.
 - Carefully release the fish back to the water.

*Lake trout mortalities to be taken to the lab for further processing (see **Comment #14**).*

10. **Round whitefish:** Retain only sexually mature, current year spawners in specific weight categories until **lethal sampling** targets met¹ (see **Table 1**); once targets have been reached, cease sampling, *unless* the full quota of lake trout has not been met.

Field processing for **lethal sampling**:

- Sacrifice the fish by quick blow to the head.
- Bag the fish, and label it clearly with the Set ID and mesh size. Place the bag in a cooler with ice packs.
- Laboratory processing will be covered in **Comment 14**.

Field processing for **non-lethal sampling**:

- Check for fin clip. If present, then release immediately. If absent, then follow next steps.
- Measure the length (fork [mm; accuracy to nearest mm]) and weight (g; accuracy to 1% of body weight) of each fish.
- Record the external DELTs. This includes abnormalities of body form, body surface, fins, eyes, lesions, tumors, scars etc. Mark any of these on the fish diagram sheet (**Figure 3** for example). Take additional pictures as necessary.

¹ In the event that one or more targeted size categories of round whitefish are **not** filled and whitefish are captured within one or more of the 'filled' size categories, sample the additional fish as per the protocols to ensure that at least 20 fish of each gender are captured and processed. After 20 fish of each sex are sampled at an area (but all size categories still unfilled), discuss situation with project coordinator.

- Using fin-clippers, take a left pectoral fin clip (first 4 rays) and place in a paper scale envelope. Label the bag immediately with the Set ID, Mesh Size, Date, and Fish ID.
- Carefully release the fish back to the water.

11. Incidental catch (non-target species such as Arctic char):

Field processing for **non-lethal sampling**:

- Check for pectoral fin clip. If present, then release. If absent, then follow next steps.
- Measure the length (fork [mm; accuracy to nearest mm]) and weight (g; accuracy to 1% of body weight) of each fish.
- Using fin-clippers, take a left pectoral fin clip (first 4 rays) and place in a paper scale envelope.
- Carefully release the fish back to the water.

*Non-target species mortalities to be taken to the lab for further processing (**Comment #14**).*

- 12.** Once the gang of nets has been fully brought into the boat, and all fish processed as needed, the gang can be set again. Repeat process of setting nets.

Laboratory Fish Processing

- 13.** At the end of each field day, all dead fish are processed in the laboratory. Round whitefish are the only species routinely lethally sampled, but may also include incidental mortalities of lake trout and by-catch species (e.g., char).

- 14.** Detailed biological assessment requirements vary by fish species (**Table 1**). The following is presented for round whitefish, but applies to other species, but only for specific measurements.

Follow these procedures (recording details in the **detailed biological analysis lab sheet** [and **field data sheet** if certain measurements were not done in the field]; see **Table 2** for measurement precision):

- **All fish.** Weigh the fish to the nearest gram
- **All fish.** Place the fish on the length board with the right side down and the nose against the end of the board. Place the unique identifier on a piece of paper adjacent to the fish and photograph each whitefish prior to examining for external DELTS
- **All fish.** Examine the fish from both sides noting any irregularities in color, appearance, deformities, fin erosions, lesions, tumours, and scars. Photograph any abnormalities and record them on the fish drawing sheet.
- **All fish.** Cut the fish open from the anus to below the gill isthmus with a sharp knife or scissors taking care not to cut or damage any organs. Inspect the interior of the fish noting any lesions, growths, deformities, tumors and discoloration of major organs including the spleen, liver, kidney, gonad and stomach. Photograph any abnormalities.
- **All fish.** Record the presence and approximate load of any parasites (low [1-10], moderate [11-24], high [>25]) and location in the gut cavity or on major organs. Photograph representative examples of lake trout or char.
- **All fish.** Open the stomach and record relative percent fullness (10% increments) and rough taxonomic composition of stomach contents.
- **All fish, including lake trout, Arctic char, whitefish.** This new protocol was determined by the TAPP on August 16. Excise the liver using a scalpel or scissors – remove the gall bladder,

taking care to take only the liver and minimize taking connective tissue and blood vessels; weigh to the nearest 0.01 g in a tared weighing boat. Verify the weight.

- **All sexually mature, current year spawners only.** Excise the gonads taking care not to damage the testes/ovary and risk losing milt, eggs or fluid; minimize the collection of connective tissue and blood vessels; weigh to the nearest 0.01 g. Verify the weight.
- **All sexually mature, current year spawners.** For females, cut open the ovary and randomly remove and count a subset of 400 eggs. Weigh to the nearest 0.01 g.
- **All fish.** Remove otoliths and pectoral fin and place in a labeled scale envelope.

15. Once the detailed biological analysis has been completed, all biological waste should be gathered and disposed of appropriately. **All offal, small non-edible fish and very large lake trout shall be returned to the lake where captured. Only those fish deemed suitable for human consumption should be frozen for distribution to the community.**

16. Until ready for shipping, dry and store aging structures (fins and otoliths) in a secure and central location.

PACKAGING & SHIPPING SAMPLES:

1. Fill out a **chain-of-custody (COC)** form for the aging structure samples.
2. Ensure all **aging samples** are packaged together in a padded envelope or box, with the COC.
3. Ensure the COC form is enclosed and then seal the cooler(s). **Label the cooler** with the following address:

North/South Consultants
83 Scurfield Boulevard
Winnipeg, Manitoba R3Y 1G4
Tel: 204-284-3366
Attention: Sue Hertam

4. Send completed COC forms and data forms (field and lab) to Azimuth.

Table 1. Program summary for Meadowbank EEM Fish Study.

Target per water body <ul style="list-style-type: none">• Exposure: Third Portage N (TPN)• Reference: Inuggugayualik(INUG) Pipedream (PDL)	Round Whitefish		Lake Trout		Arctic char, Burbot		Notes
	lethal (5 of ripe M & F in each size class)	non-lethal	non-lethal	lethal	non-lethal	lethal	<ul style="list-style-type: none">• Weather (general)• Air & water temperature (when lifting)• UTM and depth (m) for each end of the net• Effort: panels & duration of set (i.e. time & date of set and pull/run)• General observations (depth, biota etc)• Photographs
	270-400 g 401-550 g 551-700 g >701 g	Over and above the quota	100+ (tag and return to water body)	As few as possible	As few as possible	As few as possible	
Parameters							
Species (& net & mesh in which it was captured)	*	*	*	*	*	*	
Fork length (mm)	*	*	*	*	*	*	If fish species with no forked tail, then measure total length.
Total weight (g)	*	*	*	*	*	*	
Sex, maturity, reproductive status	*			*		*	Mature, IMM, resting, unknown
Gonad weight, egg wt (400 eggs to 0.01 g)	*						Weight of 400 eggs (g), weight of gonads
DELTS (internal)	*			*		*	Parasites, deformities, lesions, tumors, general condition etc.
DELTS (external)	*	*	*	*			Parasites, deformities, lesions, tumors, general condition Use fish diagram if necessary
Age	Otoliths + Pectoral Fin	Pectoral Fin	Pectoral Fin	Otoliths + Pectoral Fin	Otoliths + Pectoral Fin	Otoliths + Pectoral Fin	Otoliths and fins (4 rays) in paper envelopes.
Liver weight (g)	*						
Floy Tag (Floy)			Floy (fin clip small ones)				
Stomach Contents	*			*		*	Note identifiable items and fullness (to nearest 25%)

Table 2. Required fish survey measurements, expected precision and summary statistics.

Measurement Requirement (MMER Schedule 5, s. 16 (a) and (b))	Expected Precision	Reporting of Summary Statistics(MMER Schedule 5 s. 16) and other general reporting
Length (fork or total or standard)	+/- 1 mm	Mean, median, SD, standard error, minimum and maximum values for sampling areas
Total body weight (fresh)	+/- 1.0%	Mean, median, SD, standard error, minimum and maximum values for sampling areas
Age	+/- 1 year (10% to be independently confirmed)	Mean, median, SD, standard error, minimum and maximum values for sampling areas
Gonad weight (mature fish only)	+/- 0.01 g for sexually mature species	Mean, median, SD, standard error, minimum and maximum values for sampling areas
Egg size (if fish are sexually mature)	+/- 0.001 g	Weight, (recommended minimum sub-sample sizes of 400 eggs), mean, median, standard error, minimum and maximum values for sampling areas
Fecundity (if fish are sexually mature)	+/- 1.0%	Total number of eggs per female, mean, median, standard error, minimum and maximum for sampling areas
Weight of liver	+/- 0.01 g for sexually mature fish	Mean, median, SD, standard error, minimum and maximum values for sampling areas
Abnormalities	N/A	Presence of any lesions, tumours, parasites, or other abnormalities
Sex	N/A	

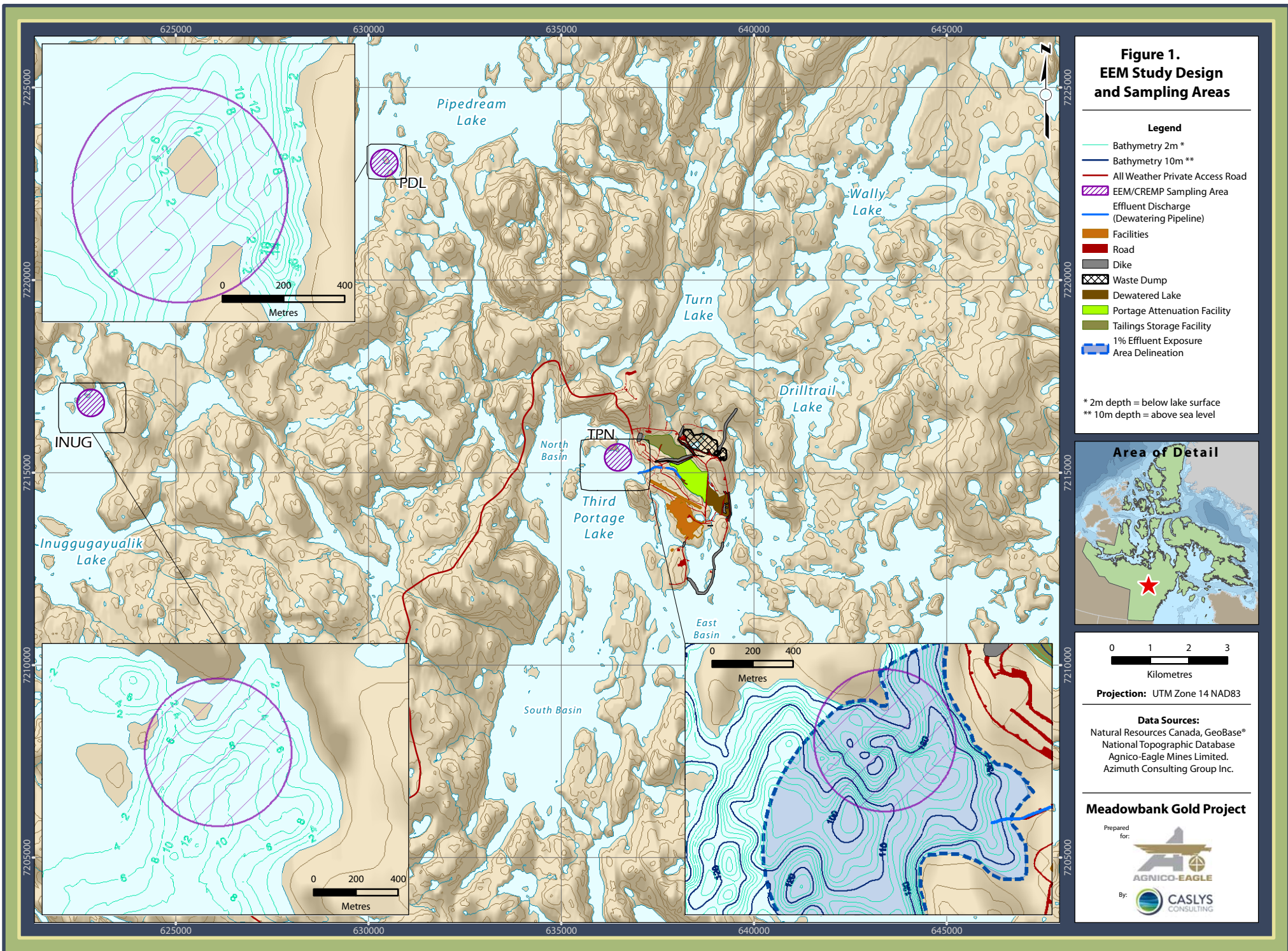
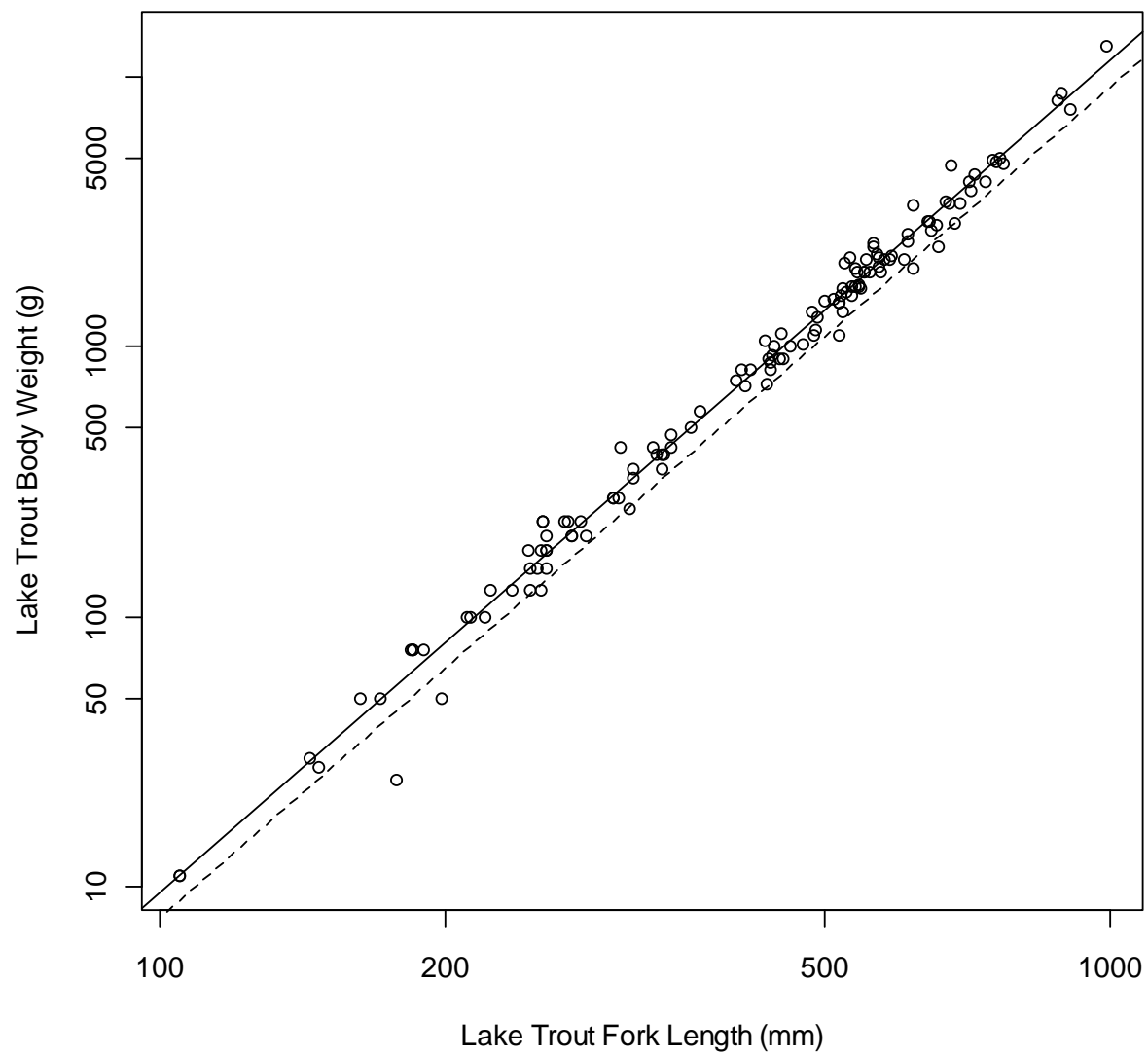


Figure 2. Relationship between body weight and fork length (solid line) and target effect size (dashed line) for Lake Trout.

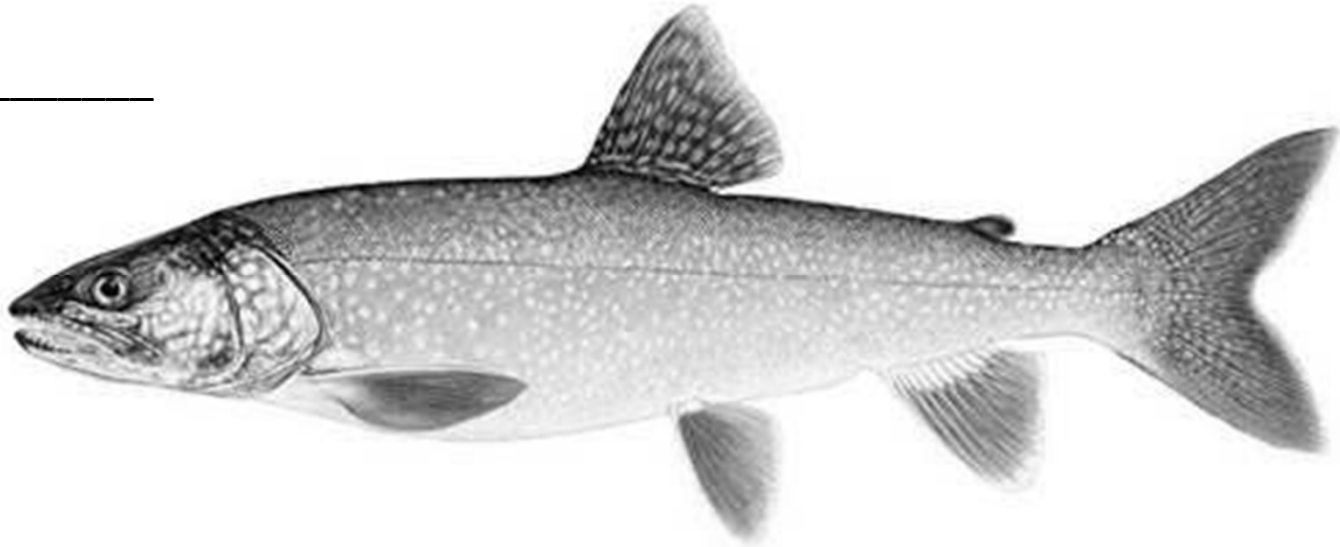


Note: axes shown on log scale.

External DELTs

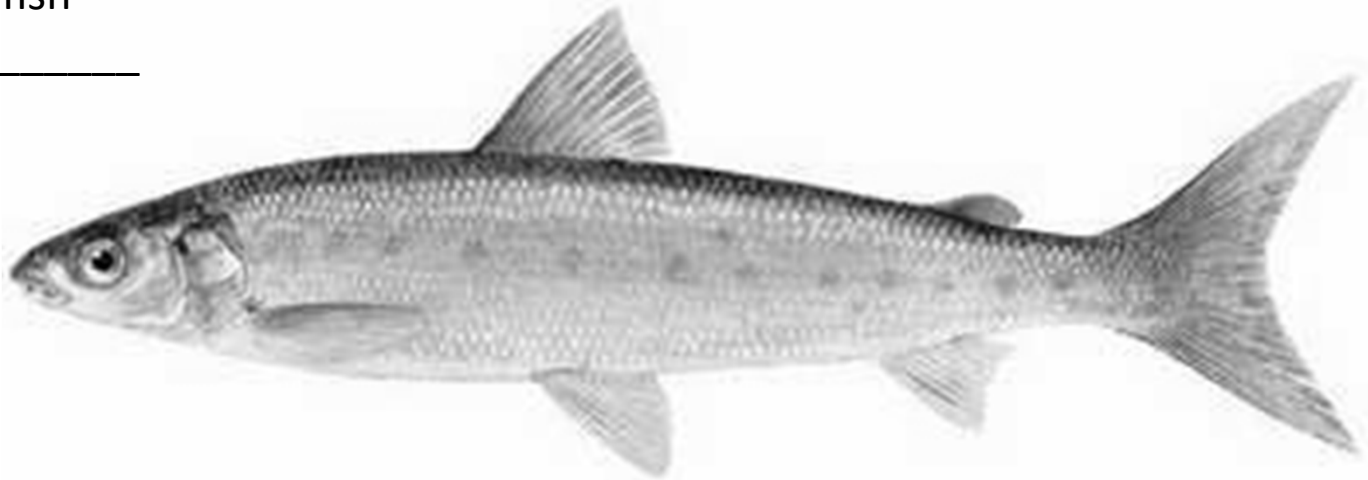
Lake Trout

Fish ID: _____



Round Whitefish

Fish ID: _____



APPENDIX G

FISH SAMPLING FIELD AND FIELD LABORATORY DATA SHEETS, AUGUST 2011



A- DAY

[illegible]

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

entered SM
QATOC RB

A-DAY

[illegible]

entered SH
QA/QC RB

A-DAY

[illegible]

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

QA/QC RB

A-DAY

[illegible]

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

entered SM
QA/QC RB

GILLNET SET & LIFT DATA

A - NIGHT B - DAY C - DAY

Set ID: <u>TPN-05 A, B, C</u>	Recorder: <u>Laura / Claire</u>	Water Body: <u>TPN</u>
Set date: <u>Aug 12/11</u>	Set time: <u>18:54</u>	Net Length (m): <u>22.7x6</u>
Start UTMS: <u>Wpt 412</u>	End UTMS: <u>Wpt 413</u>	Net Height (m): <u>1.8</u>
Easting: <u>0636932</u>	Easting: <u>0636993</u>	Net depth at each end (m): <u>2.2m / 8.9m</u>
Northing: <u>7214827</u>	Northing: <u>7214941</u>	Weather / Comments: <u>overcast</u>
Number of panels by mesh size (mm): 25: <u>S</u> 38: <u>✓</u> 51: <u>✓</u> 76: <u>✓</u> 102: <u>✓</u> 126: <u>E</u>		Water Temp (at lift) (°C): <u>10.5</u> / <u>10.5</u> / <u>10.9 (3rd)</u>
Lift date: <u>Aug 13/11</u>	Lift time: <u>08:20</u> ^(A)	Air Temp (at lift) (°C): <u>12.4</u> / <u>12.0</u>

Lift 2 date = Aug 13/11 8:20

Lift 2 time = 13:15

Lift 3 date = Aug 13/11 13:15

Lift 3 time = 19:00

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	TPN-05.01	126	518	1418	N	F	MA	RS	OT/PEC	N/A	NONE Dead
LKTR	TPN-05.02	126	503	1150	N	M	MA	RS	OT/PEC	N/A	NONE Dead
LKTR	TPN-05.03	126	392	605	N	F	MA	RS	OT/PEC	N/A	NONE Dead
LKTR	TPN-05.04	76	567	1425	Y	UN	UN	UN	PEC	87451	NONE Dead
LKTR	TPN-05.05	76	690	3100	Y	UN	UN	UN	PEC	8752	NONE
LKTR	TPN-05.06	76	345	400	Y	UN	UN	UN	PEC	8753	NONE
ARCH.	TPN-05.07	51	272	190	N	F	IM	UD	OT/PEC	N/A	lower lobe caudal missing
LKTR	TPN-05.08	51	190	50	N	IM	UN	UD	PEC	N/A	NONE Dead
ARCH.	TPN-05.09	51	275	216	N	M	IM	UD	OT/PEC	N/A	NONE Dead
ARCH.	TPN-05.10	51	294	225	Y	—	—	—	—	N/A	NONE
ARCH.	TPN-05.11	51	338	390	N	F	MA	RS	OT/PEC	N/A	NONE
ARCH.	TPN-05.12	51	250	144	N	F	IM	UD	OT/PEC	N/A	NONE Dead
ARCH.	TPN-05.13	51	301	248	N	F	IM	UD	OT/PEC	N/A	NONE Dead
LKTR	TPN-05.14	51	190	50	Y	IM	UN	UD	PEC	N/A	NONE Dead
ARCH.	TPN-05.15	51	229	148	N	M	IM	UD	OT/PEC	N/A	NONE Dead

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. UN = Unknown; PEC = Pectoral fin; DR = Dorsal ray

entered SM

ARCH

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84
85
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100

CODES
SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown
REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

A-NIGHT B-DAY C-DAY

Set ID: <u>TPN-06 A,B,C</u>	Recorder: <u>Laura / Claire</u>	Water Body: <u>TPN</u>
Set date: <u>Aug 12/11</u>	Set time: <u>19:03</u>	Net Length (m): <u>22.7 x 6</u>
Start UTMS <u>Wpt 4.14</u>	End UTMS <u>Wpt 4.5</u>	Net Height (m): <u>1.8</u>
Easting: <u>0636486</u>	Easting: <u>0636565</u>	Net depth at each end (m): <u>2.3 / 15.4</u>
Northing: <u>7215489</u>	Northing: <u>7215374</u>	Weather / Comments: <u>overcast</u>
Number of panels by mesh size (mm): 25: <u>8</u> 38: <u>1</u> 51: <u>1</u> 76: <u>1</u> 102: <u>1</u> 126: <u>E</u>		Water Temp (at lift) (°C): <u>10.5 / 10.5 / 10.9</u>
Lift date: <u>Aug 13/11</u>	Lift time: <u>09:06</u>	Air Temp (at lift) (°C): <u>12 / 12 /</u>

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
✓ LKTR	TPN-06.01	126	565	1525	Y	—	—	UN	PEC	87455	None
✓ LKTR	TPN-06.02	126	664	2964	N	M	MA	RS	OT/PEC	—	None Dead
✓ LKTR	TPN-06.03	102	510	1438	N	F	MA	RS	OT/PEC	—	None Dead
✓ ARCH	TPN-06.04	102	581	2025	Y	—	—	—	PEC	—	None
✓ LKTR	TPN-06.05	102	538	1686	N	F	MA	RS	OT/PEC	—	None Dead
✓ ARCH	TPN-06.06	102	576	2150	Y	—	—	OT	PEC	—	None
✓ LKTR	TPN-06.07	102	535	1600	Y	—	—	UN	PEC	87456	None
✓ ARCH	TPN-06.08	76	348	454	N	F	MA	RS	OT/PEC	—	None DEAD
✓ LKTR	TPN-06.09	51	551	1750	Y	—	—	OT	PEC	87457	NONE
✓ LKTR	TPN-06.10	51	425	700	Y	—	—	—	PEC	87458	NONE
✓ ARCH	TPN-06.11	51	488	1125	Y	F	MA	RS	PEC/OT	—	None Dead
✓ LKTR	TPN-06.12	51	606	1925	Y	—	—	—	PEC	87459	NONE
✓ LKTR	TPN-06.13	51	547	1775	Y	—	—	—	PEC	87460	NONE
✓ LKTR	TPN-06.14	51	266	178	N	M	IM	UN	PEC/OT	—	None Dead
✓ LKTR	TPN-06.15	38	231	126	N	M	IM	UN	OT/PEC	—	None Dead

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

entered SH
 QATAC SH

marked SH

(A)

⑧ entered MJ

©

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

A-DAY B-DAY

[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

A-DAY B-DAY

Set ID: <u>TPN-08</u>	Recorder: <u>Claire</u>	Water Body: <u>TPN</u>
Set date: <u>Aug 13/11</u>	Set time: <u>10:08</u>	Net Length (m): <u>22.7x6</u>
Start UTM's <u>WP 418</u>	End UTM's <u>WP 419</u>	Net Height (m): <u>1.8</u>
Easting: <u>14W 0636633</u>	Easting: <u>14W 0636473</u>	Net depth at each end (m): <u>start 1.8 / end 2.9</u>
Northing: <u>7214755</u>	Northing: <u>7214815</u>	Weather / Comments:
Number of panels by mesh size (mm): 25: <u>S</u> 38: <u>✓</u> 51: <u>✓</u> 76: <u>✓</u> 102: <u>✓</u> 126: <u>E</u>		Water Temp (at lift) (°C): <u>10.7°C</u>
Lift date: <u>Aug 13/11</u>	Lift time: <u>13:55</u>	Air Temp (at lift) (°C): <u>11.0°C</u>
* <u>ⓐ</u> Aug 13/11 1355		<u>ⓑ</u> 19:45

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	TPN-08.001	102	Escaped								
LKTR	TPN-08.002	76	503	1464	N	M	MA	RI	OT/PEC	—	NONE Dead
LKTR	TPN-08.003	51	548	1720	N	M	IM	UD	OT/PEC	—	NONE Dead
LKTR	TPN-08.004	102	569	1950	Y	—	—	—	PEC	87473	Gross NONE
"	TPN-08.005	76	575	2025	Y	—	—	—	PEC	87472	NONE
"	TPN-08.006	76	495	1194	N	M	IM	UD	OT/PEC	—	NONE DEAD
"	TPN-08.007	51	633	2470	N	F	MA	RS	OT/PEC	—	NONE DEAD
"	TPN-08.008	38	915	9050	Y	—	—	—	PEC	87470	NONE ALIVE
"	TPN-08.009	38	549	1875	Y	—	—	—	PEC	87471	NONE ALIVE

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

A-DAY B-DAY

(B) Aug 13 / 11

⑥ 20:20

[illegible]

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATAA- ~~RIGHT~~

Set ID: TPN-10	Recorder: Claire	Water Body: TPN
Set date: Aug 13/11	Set time: 20:15	Net Length (m): 22.7 x 7
Start UTM's WP 436	End UTM's WP 422	Net Height (m): 1.8
Easting: 0636769	Easting: 14W 0636783	Net depth at each end (m): ^{start} 1.8 / 15.4 ^{end}
Northing: 7214044	Northing: 7215014	Weather / Comments:
Number of panels by mesh size (mm):		Water Temp (at lift) (°C): 10.1°C
25: <u>S</u> 38: <u>✓</u> 51: <u>✓</u> 76: <u>2</u> 102: <u>✓</u> 126: <u>E</u> (7 panels)		Air Temp (at lift) (°C): 11.0°C
Lift date: Aug 14/11	Lift time: 10 ¹⁰	

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	TPN-10.01	25	579	1900	Y ^{1/25}	UN	UN	UN	PEC	NSC	None
ARCH	TPN-10.02	25	135	25	Y ^{1/25}	UN	UN	UN	—	—	None
ARCH	TPN-10.03	25	126	20	N	UN	IMM	UD	PEC/OT	—	None
ARCH	TPN-10.04	38/25	134	24	N	UN	IMM	UD	PEC/OT	—	None
ARCH	TPN-10.05	38/25	135	20	N	—	—	—	PEC/OT	—	None
ARCH	TPN-10.06	38/38	535	1500	Y ^{1/25}	UN	UN	UN	PEC	87450	Partially digested / Dumped in net
ARCH	TPN-10.07	38/38	580	2050	Y ^{1/25}	UN	UN	UN	PEC	—	None
ARCH	TPN-10.08	38	360	550	N	M	MA	RI	PEC/OT	—	Deformed 2 pectoral fins
LKTR	TPN-10.09	38	222	102	N	UN	Imm	UD	PEC/OT	—	None
LKTR	TPN-10.10	38	186	61	N	UN	Imm	UD	PEC/OT	—	None
LKTR	TPN-10.11	51	530	1750	Y ^{1/25}	UN	UN	UN	PEC	87449	None
ARCH	TPN-10.12	51	380	300	N	UN	Imm	UD	OTO	—	None
LKTR	TPN-10.13	51	385	574	N	UN	Imm	UD	PEC/OT	—	None
LKTR	TPN-10.14	51	252	162	N	UN	Imm	UD	PEC/OT	—	None
ARCH	TPN-10.15	51	302	294	N	UN	Imm	UD	OTO	—	Missing part of operculum

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
 MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
 AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

AUG 14

A

[illegible]

MAT. MA = Mature; IM = Immature; UN = Unknown

AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

A-NIGHT B-DAY C-NIGHT

Set ID: <u>TPN-11 A,B,C</u>	Recorder: <u>Claire</u>	Water Body: <u>TPN</u>
Set date: <u>Aug 13/11</u>	Set time: <u>20:50</u>	Net Length (m): <u>22.7 x 2</u>
Start UTM: <u>WP 424</u>	End UTM: <u>WP 423</u>	Net Height (m): <u>1.8</u>
Easting: <u>0637000</u>	Easting: <u>0637019</u>	Net depth at each end (m): <u>start 3.0 / end 9.0 m</u>
Northing: <u>7215219</u>	Northing: <u>7215269</u>	Weather / Comments: <u>Sunny</u>
Number of panels by mesh size (mm): 25: <u> </u> 38: <u> </u> 51: <u>1</u> 76: <u>1</u> 102: <u> </u> 126: <u> </u> (<u>2 panels only</u>)		Water Temp (at lift) (°C): <u>10.1°C</u>
Lift date: <u>August 14/11</u>	Lift time: <u>11:10</u> <u>REST 11:25 Aug 14/11</u>	Air Temp (at lift) (°C): <u>11°C</u>

Set (B) = Aug 14 @ 11:26

SET (B) = m + $\frac{11:20-10:10}{10:10-10:00}$

Set (C) = Aug 14/11 @ 18:02

pulled @ Aug 15/11 @ 9:25

 T_{H2O} : 10°C
 T_{AIR} : 9.5°CCATCH DATA

Pulled @ 17:55

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	TPN-11.01	51	255	160	N	UN	IMM	UD	PEC/OT	—	normal D
LKTR	TPN-11.02	51	513	1350	N	M	MA	R1	PEC/OT	—	upper lobe of caudal deformed D
LKTR	TPN-11.03	51	561	1734	N	M	MA	RS	PEC/OT	—	normal D
LKTR	TPN-11.04	51	224	150	Y $\frac{11:20}{10:00}$	UN	UN	UN	PEC	—	None D
LKTR	TPN-11.05	76	543	1670	N	M	MA	RS	PEC/OT	—	normal D
RNWH	TPN-11.06	76	383	618	N	F	MA	R1	PEC/OT	—	normal D
ARCM	TPN-11.07	76	378	650	Y $\frac{11:25}{10:00}$	UN	UN	UN	PEC	—	None A
LKTR	TPN-11.08	76	503	1408	N	F	MA	R1	PEC/OT	—	normal D
LKTR	TPN-11.09	76	432	806	N	F	MA	RS	PEC/OT	—	normal D
LKTR	TPN-11.10	51	268	153.1	N	UN	IMM	UD	OT/PEC	—	none D
↓	TPN-11.11	51	652	3225	Y				PEC	87437	none
	TPN-11.12	51	608	2400	Y				PEC	87438	adipose missing

CODESSEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = UnknownREPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

SET-B - EMPTY - NO CATCH

entered SA

87436 destroyed

entered SA

A-DAY

Set ID: TPN-12	Recorder: Claire	Water Body: TPN
Set date: Aug 14/11	Set time: 09:20	Net Length (m): 22.7 x 6
Start UTMs: WP 430	End UTMs: WP 431	Net Height (m): 1.8
Easting: 0637080	Easting: 0636944	Net depth at each end (m): start 5.2 / 13.1
Northing: 7214910	Northing: 7214945	Weather / Comments: sun/cloud, wind N 20
Number of panels by mesh size (mm): 25: 5 38: ✓ 51: ✓ 76: ✓ 102: ✓ 126: E (6 panels)		Water Temp (at lift) (°C): 10.8
Lift date: Aug 14/11	Lift time: 14:05	Air Temp (at lift) (°C): 12

[illegible]

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

entered SN

A-DACT DAN

[illegible]

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

entered SN

A-DAY

CATCH DATA											
Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	TPN-14.01	76	497	1260	Y		-	-	PEC	87426	None A

CODES
 SEX M = Male; F = Female; IM = Immature

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

entered 81

A-DAY

[illegible]

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

entered 8M

A-DAI

[illegible]

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

entured 8M

GILLNET SET & LIFT DATA

B-NIGHT

Set ID: TPN-16 (b)	Recorder: Laura	Water Body: TPN
Set date: Aug 14/11	Set time: 17:49	Net Length (m): 22.7
Start UTMs	End UTMs	Net Height (m): 1.8
Easting: Same	Easting: Same	Net depth at each end (m): 2.8 / 15.1
Northing:	Northing:	Weather / Comments: Sunny. Aug 14 = 2 pipes flowing.
Number of panels by mesh size (mm): 25: S 38: 1 51: 1 76: 2 102: 1 126: E 7 panels.		Water Temp (at lift) (°C): 10.0°C
Lift date: Aug 15/11	Lift time: 8:25	Air Temp (at lift) (°C): 9.5°C

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	TPN-16.05	38	175	60	Y (+100g)	-	-	-	-	-	None
LKTR	TPN-16.06	11	184	58	N	IM	IM	UD	PEC/OT	-	None
LKTR	TPN-16.07	51	275	190	Y (+100g)	-	-	-	PEC	-	None
LKTR	TPN-16.08	76	502	1450	Y	-	-	-	PEC	87474	None
LKTR	TPN-16.09	76	610	1625	Y	F	MA	RS	PEC/OT	87475	None
LKTR	TPN-16.10	76	562	1850	Y	-	-	-	PEC	87430	None
LKTR	TPN-16.11	76	536	1725	Y	-	-	-	PEC	87431	None
LKTR	TPN-16.12	76	492	1226	N	F	MA	RS	OT/PEC	-	None
LKTR	TPN-16.13	76	560	1732	N	F	MA	RS	OT/PEC	-	None
LKTR	TPN-16.14	76	484	1258	N	F	MA	RS	OT/PEC	-	None
LKTR	TPN-16.15	76	757	3825	Y	M	MA	RS	-	87361	recap none
LKTR	TPN-16.16	76	-	-	-	-	-	-	-	87428	recap none
LKTR	TPN-16.17	76	514	1438	N	M	MA	RS	OT/PEC	-	None
LKTR	TPN-16.18	76	732	4675	Y	-	-	-	PEC	87432	None
ARCH	TPN-16.19	102	558	1800	Y	-	-	-	PEC	-	redness (spots) around anus + peduncle

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

487429
destroyed

[illegible]

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

A-DAY

Set ID: TPN-17	Recorder: Claire	Water Body: TPN
Set date: Aug 14/11	Set time: 11:31	Net Length (m): 22.7 x 6
Start UTM's WP439	End UTM's WP440	Net Height (m): 1.8
Easting: 0636941	Easting: 0636843	Net depth at each end (m): 5.0 / 17
Northing: 7215326	Northing: 7215197	Weather / Comments: sun / clouds, wind W 20
Number of panels by mesh size (mm): 25: <u>E</u> 38: <input checked="" type="checkbox"/> 51: <input checked="" type="checkbox"/> 76: <input checked="" type="checkbox"/> 102: <input checked="" type="checkbox"/> 126: <u>S</u> 6 panels.		Water Temp (at lift) (°C): 10.8
Lift date: Aug 14/11	Lift time: 14:42	Air Temp (at lift) (°C): 12

CATCH DATA

[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

A-DAY

Set ID: TPN-18	Recorder: Claire	Water Body: TPN
Set date: Aug 14/11	Set time: 11:42	Net Length (m): 22.7 x 2
Start UTM's WP442	End UTM's WP443	Net Height (m): 1.8
Easting: 0636866	Easting: 0636834	Net depth at each end (m): 10.0 / 16
Northing: 7215416	Northing: 7215370	Weather / Comments: Sun / cloud, wind N 20
Number of panels by mesh size (mm): 25: 38 51: S 76: E 102: 126		Water Temp (at lift) (°C): 10.8
Lift date: Aug 14/11	Lift time: 15:00	Air Temp (at lift) (°C): 12

CATCH DATA

[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

A-DAY

[illegible]

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

A-DAY

Set ID: TPN - 20

Recorder:

Claire

Water Body: TPN

Set date: Aug 15/11

Set time:

07:55

Net Length (m): 22.7 x 2

Start UTM's WP 446

End UTMs

MP 447

Net Height (m): 1.8

Easting: 0636764

Easting:

0636718

Net depth at each end (m): 2.2 / 11.9

Northing: 7215647

Northing:

7215617

Weather / Comments:

Number of panels by mesh size (mm):

25: ~~6~~ 38: ~~1~~ 51: E 76: S 102: ~~1~~ 126: ~~1~~

Water Temp (at lift) (°C): 10.5

Lift date: Aug 15/11

Lift time: 15:50

Air Temp (at lift) (°C): 18

CATCH DATA

[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS: UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT: PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

A-DAY

[illegible]

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown.
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

Set ID: TPN-22	Recorder: Claire / Mike	Water Body: TPN
Set date: Aug 15 / 11	Set time: 08:13	Net Length (m): 22.7 x 6
Start UTMs WP 451	End UTMs WP 450	Net Height (m): 1.8
Easting: 0636479	Easting: 0636410	Net depth at each end (m): 29.7 / 20.9
Northing: 7214864	Northing: 7214985	Weather / Comments:
Number of panels by mesh size (mm): 25: S 38: ✓ 51: ✓ 76: ✓ 102: ✓ 126: E		Water Temp (at lift) (°C): 10.5°
Lift date: Aug 15 / 11	Lift time: 16:10	Air Temp (at lift) (°C): 08.0°

CATCH DATA

[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

A-DAY

7 panels

[illegible]

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

A-DAY B-NIGHT

Lift date: Aug 13/11
Lift date # 2 Aug 16/11

⑥ 1:ft 1734

A → No Fish

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

A - NIGHT

Set ID: <u>TPN-25</u>	Recorder:	Water Body: <u>TPN</u>
Set date: <u>Aug 15/11</u>	Set time: <u>17:00</u>	Net Length (m): <u>22.7 x 2</u>
Start UTM's <u>WP 457</u>	End UTM's <u>WP 458</u>	Net Height (m): <u>1.8</u>
Easting: <u>0637087</u>	Easting: <u>0637026</u>	Net depth at each end (m): <u>1.9 / 4.9</u>
Northing: <u>7215097</u>	Northing: <u>7215093</u>	Weather / Comments: <u>Aug 16/11 = 1 pipe spill</u> <u>Aug 15/11 = both pipes</u>
Number of panels by mesh size (mm): 25: <u>/</u> 38: <u>/</u> 51: <u>E</u> 76: <u>S</u> 102: <u>/</u> 126: <u>/</u>		Water Temp (at lift) (°C): <u>10°C</u>
Lift date: <u>Aug 16/11</u>	Lift time: <u>7:53</u>	Air Temp (at lift) (°C): <u>8°C</u>

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	TPN-25.01	51	385	630	<u>Y</u>	F	IM	UD	PEC, OT	—	none D
LKTR	TPN-25.02	51	346	392	<u>N</u>	F	IM	UD	PEC, OT	—	none D
ARCH	TPN-25.03	51	284	236	<u>N</u>	F	IM	UD	OT	—	none D
LKTR	TPN-25.04	51	593	1925	<u>Y</u>	—	—	—	PEC	87904	none
LKTR	TPN-25.05	51	574	2175	<u>Y</u>	—	—	—	PEC	87905	damaged dorsal fin pic
ARCH	TPN-25.06	51	278	230	<u>Y</u> ¹⁰	—	—	—	PEC	—	none
ARCH	TPN-25.07	51	233	122	<u>N</u>	F	IM	UN	OT	—	none D
LKTR	TPN-25.08	51	638	3025	<u>Y</u>	—	—	—	PEC	87906	none
ARCH	TPN-25.09	76	585	1675	<u>Y</u>	—	—	—	PEC	—	none
LKTR	TPN-25.10	76	572	1725	<u>Y</u>	—	—	—	PEC	87907	none
ARCH	TPN-25.11	76	416	825	<u>Y</u>	—	—	—	PEC	—	none

CODESSEX M = Male; F = Female; IM = Immature; UN = UnknownMAT. MA = Mature; IM = Immature; UN = UnknownREPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = UnknownAGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

A- NIGHT

Set ID: <u>INUG-01</u>	Recorder: <u>Mike J.</u>	Water Body: <u>INUG</u>
Set date: <u>Aug 16/11</u>	Set time: <u>18:13</u>	Net Length (m): <u>22.7x2</u>
Start UTM's	End UTM's	Net Height (m): <u>1.8m</u>
Easting: <u>622690</u>	Easting: <u>622657</u>	Net depth at each end (m): <u>2.3 16.3</u>
Northing: <u>7216943</u>	Northing: <u>7216894</u>	Weather / Comments: <u>overcast, windy rain</u>
Number of panels by mesh size (mm): 25: <u>✓</u> 38: <u>✓</u> 51: <u>107</u> 76: <u>16</u> 102: <u>✓</u> 125: <u>✓</u>		Water Temp (at lift) (°C): <u>10.</u>
Lift date: <u>Aug 17/11</u>	Lift time: <u>10:40</u>	Air Temp (at lift) (°C): <u>9°C</u>

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	INUG-01.01	51	558	1450	Y	—	—	—	—	87908	No tag - no fin ray - weak
LKTR	INUG-01.02	1	750	4550	Y	—	—	—	PEC	87908	none
LKTR	INUG-01.03	1	789	5600	Y	—	—	—	PEC	87909	none
LKTR	INUG-01.04	1	792	6750	Y	—	—	—	PEC	87910	none
LKTR	INUG-01.05	1	505	1300	Y	—	—	—	PEC	87911	none
LKTR	INUG-01.06	76	630	2575	Y	—	—	—	PEC	87912	none
LKTR	INUG-01.07	1	548	1600	Y	—	—	—	PEC	87913	none
LKTR	INUG-01.08	1	549	1556	N	F	MA	RS	OTO/PEC	—	none D
LKTR	INUG-01.09	51	278	208	N	F	IM	UD	OTO/PEC	—	none D
LKTR	INUG-01.10	51	267	204	N	F	IM	UD	OTO/PEC	—	none D
RNWH	INUG-01.11	51	413	656	N	M	MA	RI	OTO/PEC	—	none D
RNWH	INUG-01.12	51	408	664	N	F	MA	RI	OTO/PEC	—	none D
RNWH	INUG-01.13	51	400	642	N	F	MA	RI	OTO/PEC	—	none D
RNWH	INUG-01.14	51	360	488	N	F	MA	RI	OTO/PEC	—	none D
RNWH	INUG-01.15	51	310	292	N	M	MA	RI	OTO/PEC	—	none D

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

released
 1
 2
 3

CATCH DATA (continued)[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown

MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown

AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

A- NIGHT

Set ID: <u>INUG-02</u>	Recorder: <u>Mike / Laura</u>	Water Body: <u>INUG</u>
Set date: <u>Aug 16/11</u>	Set time: <u>18:25</u>	Net Length (m): <u>22.7 x 6</u>
Start UTMs <u>(14)</u>	End UTMs <u>(14)</u>	Net Height (m): <u>1.8m</u>
Easting: <u>623061</u>	Easting: <u>622923</u>	Net depth at each end (m): <u>6 16.1</u>
Northing: <u>7216287</u>	Northing: <u>7216335</u>	Weather / Comments: <u>overcast, windy & rain</u>
Number of panels by mesh size (mm): 25: <u>1</u> 38: <u>1</u> 51: <u>1</u> 76: <u>1</u> 102: <u>1</u> 126: <u>(5)</u>		Water Temp (at lift) (°C): <u>13.5°C</u>
Lift date: <u>Aug 17/11</u>		Air Temp (at lift) (°C): <u>8</u>
Lift time: <u>11:33</u>		

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	INUG-02.01	25	310	290	Y				PEC		none
LKTR	INUG-02.02	38			N						→ Escaped
LKTR	INUG-02.03	38			N						→ Escaped
RNWH	INUG-02.04	38	241	126	N	IM	IM	UN	PEC/OT		none D
LKTR	INUG-02.05	38	217	100	N	IM	IM	UN	PEC/OT		none D
LKTR	INUG-02.06	38	225	114	N	IM	IM	UN	PEC/OT		none D
LKTR	INUG-02.07	38	626	2400	Y				PEC	87942	none
RNWH	INUG-02.08	51	332	328	N	IF	IM	UD	PEC/OT		none D
LKTR	INUG-02.09	51	514	1560	Y				PEC	87915	none
LKTR	INUG-02.10	51	544	1508	Y				PEC	87916	none
LKTR	INUG-02.11	51	555	1550	Y				PEC	87917	Slight dorsal fin deformity + caudal fin
LKTR	INUG-02.12	51	598	2150	Y				PEC	87919	dorsal fin slightly deformed
LKTR	INUG-02.13	51	270	194	N	IF	IM	UD	OT/PEC		none D
RNWH	INUG-02.14	76	406	716	N	F	MA	RI	OT/PEC		none D
LKTR	INUG-02.15	76	406	654	N	M	IMM	UD	OT/PEC		none D

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

[illegible]

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

DAY

Set ID: INUG-03	Recorder: Laura	Water Body: INUG
Set date: Aug 17/11	Set time: 9:57	Net Length (m): 22.7 x 6
Start UTM's (Wpt 463)	End UTM's (14) (Wpt 464)	Net Height (m): 1.8
Easting: 0623060	Easting: 0622922	Net depth at each end (m): 2m / 4.4m
Northing: 7217055	Northing: 7217078	Weather / Comments:
Number of panels by mesh size (mm): 25: 5 38: 51: 76: 102: 126: E		Water Temp (at lift) (°C): 13° (on eagle)
Lift date: Aug 17/11	Lift time: 16:31	Air Temp (at lift) (°C): 7°C

[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

DAY

Set ID: INUG-04	Recorder: Laura	Water Body: INUG
Set date: Aug 17/11	Set time: 10:12	Net Length (m): 22.7 x 6
Start UTM's (14) (Wyp 465)	End UTM's (14) (Wyp 466)	Net Height (m): 1.8
Easting: 0623297	Easting: 0623163	Net depth at each end (m): 2.8 / 2.9
Northing: 7216702	Northing: 7216681	Weather / Comments:
Number of panels by mesh size (mm):		Water Temp (at lift) (°C): 13.2°
25: <input checked="" type="checkbox"/> E 38: <input checked="" type="checkbox"/> 51: <input checked="" type="checkbox"/> E 76: <input checked="" type="checkbox"/> S 102: <input checked="" type="checkbox"/> 126: <input checked="" type="checkbox"/> S		Air Temp (at lift) (°C):
Lift date: Aug 17/11	Lift time: 17:32	

CATCH DATA

[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

DAY

[illegible]

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray.

Set ID: INUG-06	Recorder: Laura	Water Body: INUG
Set date: Aug 17/11	Set time: 12:20	Net Length (m): 22.7 x 2
Start UTMs 844 (wypt 469)	End UTMs (wypt 470)	Net Height (m): 1.8
Easting: 0623367	Easting: 0623248	Net depth at each end (m): 1.7 / 1.8 m
Northing: 7216493	Northing: 7216519	Weather / Comments:
Number of panels by mesh size (mm): 25: <input checked="" type="checkbox"/> 38: <input type="checkbox"/> 51: <u>S</u> 76: <u>E</u> 102: <input type="checkbox"/> 126: <input type="checkbox"/>	(2 panels)	Water Temp (at lift) (°C): 13.2°C
Lift date: Aug 17/11	Lift time: 17:53	Air Temp (at lift) (°C):

[illegible]

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

NIGHT

Set ID: <u>INUG-07</u>	Recorder: <u>Laura</u>	Water Body: <u>INUG</u>
Set date: <u>Aug 17/11</u>	Set time: <u>16:51</u>	Net Length (m): <u>22.7+6</u>
Start UTM's <u>(14)</u> (<u>Wpt 471</u>)	End UTM's <u>(14)</u> (<u>Wpt 472</u>)	Net Height (m): <u>1.8</u>
Easting: <u>0623141</u>	Easting: <u>0623138</u>	Net depth at each end (m): <u>1.5m / 3.5</u>
Northing: <u>7215666</u>	Northing: <u>7215746</u>	Weather / Comments:
Number of panels by mesh size (mm): 25: <u>S</u> 38: <u>✓</u> 51: <u>✓</u> 76: <u>✓</u> 102: <u>✓</u> 126: <u>E</u>		Water Temp (at lift) (°C): <u>12.7°C</u>
Lift date: <u>Aug 18/11</u>	Lift time: <u>8:40</u>	Air Temp (at lift) (°C): <u>7.0°C</u>

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	INUG-07.01	102	504	1294	N	M	MA	RS	PEC/OT	-	NONE
ARCH	INUG-07.02	76	868	7650	Y	-	-	-	PEL	-	NONE
LKTR	INUG-07.03	76	519	1104	N	M	IM	UD	PEC/OT	-	NONE
LKTR	INUG-07.04	51	558	1750	Y	-	-	-	PEL	87925	NONE
LKTR	INUG-07.05	51	740	4600	Y	-	-	-	PEC	87438	NONE
LKTR	INUG-07.06	51	261	168	N	F	IM	UD	PEC/OT	-	NONE
LKTR	INUG-07.07	51	518	1400	Y	-	-	-	PEC	87439	NONE
LKTR	INUG-07.08	51	243	170	Y	-	-	-	PEL	-	NONE
LKTR	INUG-07.09	38	206	84	N	F	IM	UD	PEC/OT	-	NONE
LKTR	INUG-07.10	38	219	94	N	UN	IM	UD	PEC/OT	-	NORMAL
LKTR	INUG-07.11	38	554	1775	Y	-	-	-	PEC	87440	NONE
LKTR	INUG-07.12	38	223	112	N	F	IM	UD	PEC/OT	-	NONE
LKTR	INUG-07.13	38	232	170	Y	-	-	-	PEC	-	NONE
LKTR	INUG-07.14	25	ESCAPED	-	-	-	-	-	PEC	-	NONE
ARCH	INUG-07.15	25	150	40	N	UN	IM	UP	PEC/OT	-	NONE

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown

MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown

AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

NIGHT

Set ID: <u>INUG-08</u>	Recorder: <u>Laura</u>	Water Body: <u>INUG</u>
Set date: <u>Aug 17/11</u>	Set time: <u>17:34</u>	Net Length (m): <u>22.7 x 2</u>
Start UTMS <u>(Wyp 443)</u>	End UTMS <u>(14)</u> <u>(Wyp 424)</u>	Net Height (m): <u>1.8</u>
Easting: <u>0623193</u>	Easting: <u>0623150</u>	Net depth at each end (m): <u>1.8 / 5.4</u>
Northing: <u>7215863</u>	Northing: <u>7215868</u>	Weather / Comments:
Number of panels by mesh size (mm):		Water Temp (at lift) (°C): <u>12.7°C</u>
25: <u> </u> 38: <u> </u> 51: <u>E</u> 76: <u>S</u> 102: <u> </u> 126: <u> </u>		Air Temp (at lift) (°C): <u>7.5°C</u>
Lift date: <u>18 Aug 11</u>	Lift time: <u>9:50</u>	

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	INUG-08.01	51	514	1450	Y	-	-	-	PEC	87441	none
LKTR	INUG-08.02	51	584	1950	Y	-	-	-	PEC	87442	none
LKTR	INUG-08.03	51	564	1650	Y	-	-	-	PEC	87443	none
RNWA	INUG-08.04		370	520	N	F	MA	RI	PEC/OT	-	NONE D
RNWA	INUG-08.05		392	600	N	M	MA	RI	PEC/OT	-	none D
LKTR	INUG-08.06		258	172	N	M	IM	UD	PEC/OT	-	NONE D
LKTR	INUG-08.07		518	1500	Y	-	-	-	PEC	87444	scar on belly
RNWA	INUG-08.08		384	572	N	M	MA	RI	PEC/OT	-	none D
RNWA	INUG-08.09		376	536	N	M	MA	RI	PEC/OT	-	NONE D
ARCH	INUG-08.10		297	176	(Y (10))	-	-	-	PEC	-	mangled tail
RNWA	INUG-08.11		370	500	N	F	MA	RI	PEC/OT	-	NORMAL D
"	INUG-08.12		365	468	N	F	MA	RI	PEC/OT	-	normal D
"	INUG-08.13		281	204	N	F	IM	UD	PEC/OT	-	normal D
"	INUG-08.14		282	234	N	F	IM	UD	PEC/OT	-	none D
LKTR	INUG-08.15	✓	231	132	N	UN	IM	UD	PEC/OT	-	NONE D

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

CATCH DATA (continued)[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

Set ID: <u>1NUG-09</u>	Recorder: <u>MJLc</u>	Water Body: <u>1NUG</u>
Set date: <u>18 Aug 11</u>	Set time: <u>8:20</u>	Net Length (m): <u>22.7 x 6</u>
Start UTM's	End UTM's	Net Height (m): <u>1.8</u>
Easting: <u>622449 (475)</u>	Easting: <u>622318 (476)</u>	Net depth at each end (m): <u>2.0 / 0.3</u>
Northing: <u>7217011</u>	Northing: <u>7217101</u>	Weather / Comments:
Number of panels by mesh size (mm):		Water Temp (at lift) (°C): <u>11.6°C</u>
<u>25(2)</u> 38: <u>1</u> 51: <u>1</u> 76: <u>1</u> 102: <u>1</u> 126: <u>1 (5)</u>		Air Temp (at lift) (°C): <u>8°C</u>
Lift date: <u>18 Aug 11</u>	Lift time: <u>15:30</u>	

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	1NUG-09.01	51	332	450	Y	-	-	-	PEC	87445	none
LKTR	1NUG-09.02	76	875	5950	Y	-	-	-	PEC	87446	none
n	1NUG-09.03	76	616	2425	Y	-	-	-	PEC	48476	none
RNWH	1NUG-09.04	76	420	722	N	M	MA	RI	PEC/DR	-	none D
LKTR	1NUG-09.05	102	499	1550	Y	-	-	-	PEC	48477	none

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

DAY

CATCH DATA

CODES

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown

AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

DAY

[illegible]

SEX M = Male; F = Female; IM = Immature; UN = Unknown **REPRO STATUS** UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown **AGE STRUCT.** PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

Set ID: INUG-12	Recorder: Mike	Water Body: INUG
Set date: 18 Aug 11	Set time: 18:20	Net Length (m): 22.7+2
Start UTM's	End UTM's	Net Height (m): 1.8
Easting: 623056 (481)	Easting: 623005 (482)	Net depth at each end (m): 1.6 14.3
Northing: 7216294	Northing: 7216299	Weather / Comments:
Number of panels by mesh size (mm): 25: 38: 51: 51 76: 51 102: 126:		Water Temp (at lift) (°C): 12.4
Lift date: 18 Aug 11	Lift time: 16:30	Air Temp (at lift) (°C): 8

[illegible]

SEX. M = Male; F = Female; IM = Immature; UN = Unknown REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

NIGHT

Set ID: <u>INUG-13</u>	Recorder: <u>Nike</u>	Water Body: <u>INUG</u>
Set date: <u>Aug 19/11</u>	Set time: <u>15:50</u>	Net Length (m): <u>22.7x6</u>
Start UTMs <u>WP 486</u>	End UTMs	Net Height (m): <u>1.8</u>
Easting: <u>0622369</u>	Easting: <u>622349 (483)</u>	Net depth at each end (m): <u>1.6 17.6</u>
Northing: <u>7217026</u>	Northing: <u>7217156</u>	Weather-/ Comments:
Number of panels by mesh size (mm): 25: <u>1</u> 38: <u>1</u> 51: <u>1</u> 76: <u>1</u> 102: <u>1</u> 126: <u>1</u>		Water Temp (at lift) (°C): <u>11.6</u>
Lift date: <u>Aug 19/11</u>	Lift time: <u>8:25</u>	Air Temp (at lift) (°C): <u>7</u>

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	INUG-13.01	76	538	1500	Y	-	-	-	PEC	48479	none
RNWH	INUG-13.02	76	398	664	N	M	MA	RI	PEC/OT	-	NONE D.
LKTR	INUG-13.03	76	494	1400	Y	-	-	-	PEC	48480	none
RNWH	INUG-13.04	↓	420	748	N	M	MA	RI	PEC/OT	-	NONE D.
LKTR	INUG-13.05	↓	741	4032	N	F	MA	RS	PEC/OT	-	NONE D.
ARGR	INUG-13.06	↓	341	450	Y	-	-	-	PEC	-	none
LKTR	INUG-13.07	↓	515	1386	N	F	MA	RS	PEC/OT	-	NONE D
LKTR	INUG-13.08	51	253	160	Y(+10)	-	-	-	PEC	-	none
RNWH	INUG-13.09	51	385	596	N	M	MA	RI	PEC/OT	-	NONE D.
LKTR	INUG-13.10	↓	356	460	N	M	IM	VD	PEC/OT	-	NONE D
LKTR	INUG-13.11	↓	542	1550	Y	-	-	-	PEC	48481	none
RNWH	INUG-13.12	↓	385	602	N	F	MA	RI	PEC/OT	-	NONE D
LKTR	INUG-13.13	↓	205	100	Y(+10)	-	-	-	PEC	-	none
RNWH	INUG-13.14	↓	352	434	N	M	MA	RI	PEC/OT	-	NONE D
LKTR	INUG-13.15	38	175	-	N(+10)	-	-	-	PEC	-	none

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

CATCH DATA (continued)[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown

MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown

AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

NIGHT

Set ID: <u>INUG-14</u>	Recorder: <u>MJK</u>	Water Body: <u>INUG</u>
Set date: <u>Aug 18/11</u>	Set time: <u>16:15</u>	Net Length (m): <u>22.7 x 2</u>
Start UTM's	End UTM's	Net Height (m): <u>1.8</u>
Easting: <u>623105 (484)</u>	Easting: <u>623043</u>	Net depth at each end (m): <u>1.9 / 6.5</u>
Northing: <u>7216136</u>	Northing: <u>7216127</u>	Weather / Comments:
Number of panels by mesh size (mm):		Water Temp (at lift) (°C): <u>12.4</u>
25: _____ 38: _____ 51: <u>(5)</u> 76: <u>(5)</u> 102: _____ 126: _____		Air Temp (at lift) (°C): <u>7</u>
Lift date: <u>19 Aug 11</u>	Lift time: <u>9:05</u>	

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	INUG-14.01	51	283	238	N	F	IM	UD	PEC/OT	—	NONE D
RNWH	INUG-14.02		405	646	N	FE	MA	RI	PEC/OT	—	NONE D
LKTR	INUG-14.03		357	490	N	M	IM	UD	PEC/OT	—	NONE D
LKTR	INUG-14.04		—	—	—	—	—	—	—	87908	recap
LKTR	INUG-14.05		245	144	N	F	IM	UD	PEC/OT	—	NONE D
LKTR	INUG-14.06		375	625	Y	—	—	—	PEC	848482	none
ARCH	INUG-14.07		283	158	Y (+N)	—	—	—	PEC	—	none
RNWH	INUG-14.08		375	502	N	M	MA	RI	PEC/OT	—	NONE D
RNWH	INUG-14.09		383	602	N	M	MA	RI	PEC/OT	—	NONE D
RNWH	INUG-14.10	✓	343	436	N	M	MA	RI	PEC/OT	—	NONE D
RNWH	INUG-14.11	76	375	532	N	M	MA	RI	PEC/OT	—	NONE D
LKTR	INUG-14.12		577	1700	Y	—	—	—	PEC	48483	none
LKTR	INUG-14.13		625	2884	N	M	MA	RS	PEC/OT	—	NONE D
RNWH	INUG-14.14		385	570	N	M	MA	RI	PEC/OT	—	NONE D
LKTR	INUG-14.15	✓	548	1788	N	F	MA	RI	PEC/OT	—	NORMAL D

CODESSEX M = Male; F = Female; IM = Immature; UN = UnknownREPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = UnknownMAT. MA = Mature; IM = Immature; UN = UnknownAGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

A-NIGHT

Set ID: PD-01	Recorder: Susan Hartman/Claire	Water Body: Pipedream Lake
Set date: August 16, 2011	Set time: 1830	Net Length (m): 22.7 x 5
Start UTM's WP 108 14W	End UTM's WP 109 14W	Net Height (m): 1.8
Easting: 0630472	Easting: 0630350	Net depth at each end (m): 3.7 1 6.4
Northing: 7222914	Northing: 7223012	Weather / Comments: overcast, rain, wind E ~20 km/hr
Number of panels by mesh size (mm): 25: <u>S</u> 38: <u>✓</u> 51: <u>✓</u> 76: <u>✓</u> 102: <u>✓</u> 126: <u>E</u>		Water Temp (at lift) (°C): 11°C
Lift date: August 17, 2011	Lift time: 1030	Air Temp (at lift) (°C): 11.5°C

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	PD-01.01	126	621	2600	Y	-	-	-	PEC	77152	NORMAL A
LKTR	PD-01.02	102	819	7600	Y	-	-	-	PEC	77153	Dorsal fin form A
RNMH	PD-01.03	126 ¹²⁶	390	748	N	F	MA	RI	PEC/OT	-	NORMAL D
RNMH	PD-01.04	102	386	742	N	F	MA	RI	PEC/OT	-	NORMAL D
LKTR	PD-01.05	76	820	6000	Y	-	-	-	PEC	77154	lightly net damage small scratch dorsal side A
LKTR	PD-01.06	76	344	396	N	M	IM	VD	PEC/OT	-	NORMAL D
RNMH	PD-01.07	76	372	550	N	F	MA	RI	PEC/OT	-	NORMAL D
RNMH	PD-01.08	76	247	158	N	M	IM	UN	PEC/OT	-	NORMAL D
LKTR	PD-01.09	76	282	220	N	F	IM	VD	PEC/OT	-	NORMAL D
LKTR	PD-01.10	76	283	228	N	F	IM	VD	PEC/OT	-	NORMAL D
LKTR	PD-01.11	76	278	212	N	M	IM	VD	PEC/OT	-	NORMAL Net damage D
LKTR	PD-01.12	5138	804	6100	Y	-	-	-	PEC	77155	damage off dorsal (pic 77155) red spots below pectorals A
LKTR	PD-01.13	38	811	4450	Y	-	-	-	PEC	77156	unusually skinny flaccid belly A
ARCH	PD-01.14	38	243	152	N	M	IMH	VD	PEC/OT	-	None D
ARCH	PD-01.15	38	255	172	N	F	IM	VD	PEC/OT	-	NONE D

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

CATCH DATA (continued)[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

A-NIGHT

Set ID: PD-02.	Recorder: Susan Kortum.	Water Body: Pipedream
Set date: August 16, 2011	Set time: 1840	Net Length (m): 22.7 x 2
Start UTM's W 110 14W	End UTM's W 111 14W	Net Height (m): 1.8
Easting: 0630313	Easting: 0630251	Net depth at each end (m): 6.2 m / 8m.
Northing: 7223152	Northing: 7223190	Weather / Comments: overcast windy E ~20km/hr
Number of panels by mesh size (mm):		Water Temp (at lift) (°C): 11°C
25: 38: 51: 8 76: E 102: 126:		Air Temp (at lift) (°C): 8°C
Lift date: August 17, 2011	Lift time: 1135	

[illegible]

CODES

SEX. M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown
REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

A - DAY B - NIGHT

Set ID: PD-03	Recorder: Susan Newman	Water Body: Pipe dream
Set date: August 17, 2011	Set time: 1150	Net Length (m): 22.7 x 2
Start UTMs WP 112 14W	End UTMs WP 113 14W	Net Height (m): 1.8
Easting: 0630766	Easting: 0630709	Net depth at each end (m): 3.1 / 7.6
Northing: 7223440	Northing: 7223964	Weather / Comments: overcast, wind E 20-30 km/hr.
Number of panels by mesh size (mm): 25: ___ 38: ___ 51: <u>E</u> 76: <u>S</u> 102: ___ 126: ___		Water Temp (at lift) (°C): (A) 11°C / (B) 11°C
Lift date: August 17, 2011	Lift time: 1445	Air Temp (at lift) (°C): 9.5°C / 7.5°C

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	PD-03.01	51	403	640	Y	—	—	—	PEC	77157	none A
LKTR	PD-03.02	51	280	.465 lbs	N	M	IM	UD	PEC/OT	—	none D
LKTR	PD-03.03	51	281	.545 lbs	N	M	IM	UD	PEC/OT	—	none D
LKTR	PD-03.04	51	482	1090	N	F	MA	RS	PEC/OT	—	none D
LKTR	PD-03.05	51	496	1418	N	F	MA	RI	PEC/OT	—	none D
ARCM	PD-03.06	51	246	.355 lbs	N	M	IM	UD	PEC/OT	—	none D
LKTR	PD-03.07	51	—	—	N	—	—	—	—	—	—
RNWM	PD-03.08	51	547	564	N	M	MA	RI	PEC/OT	—	none D
LKTR	PD-03.09	51	522	1316	N	F	MA	RS	PEC/OT	—	none D
LKTR	PD-03.10	51	410	.4100	Y	—	—	—	PEC	77158	none D
LKTR	PD-03.11	51	269	230-14	Y ^{1/2 to 10}	—	—	—	PEC	—	none A
ARCM	PD-03.12	51	272	.470 lbs	N	U	IM	UD	PEC/OT	—	none D
LKTR	PD-03.13	51	253	.330 lbs	N	F	IM	UD	PEC/OT	—	none D
LKTR	PD-03.14	51	220	.265 lbs	N	U	IM	UD	PEC/OT	—	none D

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

died after release

GILLNET SET & LIFT DATA

A-DAY B-NIGHT

Set ID: <u>PD-04 AB</u>	Recorder: <u>Sue Hartman</u>	Water Body: <u>Pipedream</u>
Set date: <u>August 17, 2011</u>	Set time: <u>12:15</u>	Net Length (m): <u>22.7x6</u>
Start UTMs: <u>N6114 14W</u>	End UTMs: <u>N6115 14W</u>	Net Height (m): <u>1.8</u>
Easting: <u>0630601</u>	Easting: <u>0630458</u>	Net depth at each end (m): <u>9.5 / 9.9</u>
Northing: <u>7223313</u>	Northing: <u>7223363</u>	Weather / Comments: <u>overcast & windy ~20-30km/hr east</u>
Number of panels by mesh size (mm): 25: <u>5</u> 38: <u>✓</u> 51: <u>✓</u> 76: <u>✓</u> 102: <u>✓</u> 126: <u>E</u>		Water Temp (at lift) (°C): <u>(A) 11°C (B) 11°C</u>
Lift date: <u>August 17, 2011</u>	Lift time: <u>1702</u>	Air Temp (at lift) (°C): <u>9.5°C 7.5°C</u>

(B) set Aug 17, 2011 17:15 @ LPT 0935 Aug 18

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
no catch											
LKTR	PD-04.01	126	304	236	Y				PEC	77159	none A
LKTR	PD-04.02	102	534	1600	Y				PEC	77160	none A
LKTR	PD-04.03	102	818	6300	Y				PEC	77161	none A
LKTR	PD-04.04	102	584	2188	N	M	MA	RS	PEC/OTO		none D
LKTR	PD-04.05	76	521	1598	N	M	MA	RI	PEC/OTO		none D
LKTR	PD-04.06	76	566	1966	N	F	MA	RI	PEC/OTO		none D
LKTR	PD-04.07	76	532	1700	Y					77162	nick in adipose A
LKTR	PD-04.08	51	638	2552	N	M	IM	UD	PEC/OTO		none D
LKTR	PD-04.09	51	390	554	N	F	IM	UD	PEC/OTO		none D
LKTR	PD-04.10	51	697	3960	Y					77163	none A
LKTR	PD-04.11	38	184	61	N	F	IM	UD	PEC/OTO		none D
LKTR	PD-04.12	38	203	80	N	M	IM	UD	PEC/OTO		none D
LKTR	PD-04.13	38	193	78	N	F	IM	UD	PEC/OTO		none D

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

15 VLA

CATCH DATA (continued)[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown

MAT. MA = Mature; IM = Immature; UN = Unknown

AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

① - DAY

[illegible]

CODES

REPRO STATUS. UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

GILLNET SET & LIFT DATA

(A) - DAY

Set ID: PD-06	Recorder: <u>Sam / Laura</u>	Water Body: <u>Pipedream</u>
Set date: <u>August 18, 2011</u>	Set time: <u>1044</u>	Net Length (m): <u>22.7 x 6</u>
Start UTM _s <u>NP(119) 14W</u>	End UTM _s <u>NP(120) 14W</u>	Net Height (m): <u>1.8</u>
Easting: <u>0630452</u>	Easting: <u>0630305</u>	Net depth at each end (m): <u>2.7 / 4.3</u>
Northing: <u>7222898</u>	Northing: <u>7222869</u>	Weather / Comments: <u>overcast winds ESE ~20 km/hr</u>
Number of panels by mesh size (mm): 25: <u>5</u> 38: <u>✓</u> 51: <u>✓</u> 76: <u>✓</u> 102: <u>✓</u> 126: <u>E</u>		Water Temp (at lift) (°C): <u>10.5°C</u>
Lift date: <u>Aug 18/11</u>	Lift time: <u>16:00</u>	Air Temp (at lift) (°C): <u>8°C</u>

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	PD06.01	126	584	1850	Y	—	—	—	PEC	77165	none A
LKTR	PD06.02	102	548	1675	Y	—	—	—	PEC	77166	none A

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

(A) DAY. (B) NIGHT

⑧ Lift
Aug 19 lift time 0815

[illegible]

CODES

SEX M = Male; F = Female; IM = Immature; UN = Unknown
MAT. MA = Mature; IM = Immature; UN = Unknown

REPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = Unknown
AGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

own

③	LKTR	3	0
	ARCH	0	0
	BNWK	1	1

GILLNET SET & LIFT DATA

(A) - DAY (B) - NIGHT

Set ID: PD-08 A,B	Recorder: Sue/Laura	Water Body: Pipedream
Set date: August 18, 2011	Set time: 1110	Net Length (m): 22.7 x 6
Start UTMs: 123	End UTMs: 124	Net Height (m): 1.8
Easting: 0630412	Easting: 0630263	Net depth at each end (m): 3.1 / 8.5
Northing: 7222688	Northing: 7222712	Weather / Comments: overcast, wind ESE ~ 20km/hr
Number of panels by mesh size (mm):		Water Temp (at lift) (°C): (A) 10.5 (B) 10.5°C
25: S 38: S 51: ✓ 76: ✓ 102: ✓ 126: E		Air Temp (at lift) (°C): 8°C 7°C
Lift date: Aug 18/11	Lift time: 16:28	

(B) Set Aug 18/11 1640

(B) 0845 lift Aug 19, 2011

CATCH DATA

Species	Sample ID	Mesh Size (mm)	Fork Length (mm)	Weight (g)	Weighed in field? (Y/N)	Sex	Mat.	Repro Status	Age Struct.	Tag ID	Ext. DELT / Parasites?
LKTR	PD-08.01a	102	629	2746	N	F	MA	RI	PECTOR	—	none ID
ARUN	PD-08.01	102	255	174	N	U	IM	UD	PECTOR	—	none D
LKTR	PD-08.02	76	559	1675	Y	—	—	—	PEL	77167	none A
LKTR	PD-08.03	76	686	3400	Y	—	—	—	PEL	77168	none adipose dipan fin A
LKTR	PD-08.04	51	282	225	Y (600)	—	—	—	PEL	77169	none A
LKTR	PD-08.05	38	235	136	N	M	IM	UD	PEC/OTO	—	none D
LKTR	PD-08.06	38	202	80	N	F	IM	UD	PEC/OTO	—	none D
LKTR	PD-08.07	38	168	52	Y (600)	—	—	—	PEL	—	none A

CODESSEX M = Male; F = Female; IM = Immature; UN = UnknownMAT. MA = Mature; IM = Immature; UN = UnknownREPRO STATUS UD = Undeveloped; RI = Ripe; RU = Running; SP = Spent; RS = Resting; UN = UnknownAGE STRUCT. PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEET

Recorder: Laura Henderson

Date: Aug 13/11

Stomach (gut)
Parasites -
always on surface
of stomach & oesophagus
caecae

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs 400 (g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
✓ TPN-05.11	OT/PEC	—	normal	—	—	—	normal	normal	—	60% full; zooplankton Stomach.
✓ TPN-05.13	OT/PEC	—	normal	—	—	—	normal	normal	mod pic 3732/3733 inspired parasites around gut	Lower Intes. 100% full chironomids.
✓ TPN-05.09	OT/PEC	—	normal.	—	—	—	normal	normal	gut parasites mod	Lower intestine 100% Chironomids.
✓ TPN-05.07	OT/PEC	—	normal	—	—	—	normal	normal	gut parasites moderate	MT stomach; full intestine (100%).
✓ TPN-05.12.	OT/PEC	—	normal	—	—	—	normal	normal	gut parasites moderate	Stom. Empty, Intes. full.
✓ TPN-05.15.	OT/PEC	—	normal	—	—	—	normal	normal	gut parasites low	Stom. 50% full Intes. full. (100)
✓ TPN-05.01	OT/PEC	—	normal	—	—	—	normal	normal.	mod Stomach parasites.	Stomach MT.
✓ TPN-05.03.	OT/PEC	—	normal	—	—	—	normal	normal	gut parasites (moderate)	Stomach 10% full
✓ TPN-05.02	OT/PEC	—	pic 3735 normal:	—	—	—	normal	normal	gut parasites low (3)	Stomach 10% full
✓ TPN-06.08	OT/PEC	—	pic 3736 normal.	—	—	—	normal	normal	moderate gut parasites	Stomach 20% full.
✓ TPN-06.03	OT/PEC	—	normal	—	—	—	normal	normal	gut paras. moderate.	Stomach 50% full
✓ TPN-06-05	OT/PEC	—	normal	—	—	—	normal	normal	low gut	Stomach 50% full
TPN-06.14	OT/PEC	—	normal	—	—	—	normal	normal	mod para cysts on surface of stomach etc	Stomach 10% full
✓ TPN-06.11	OT/PEC	—	appears to have parasitic cyst pic 3737, 38	—	—	—	normal	normal	mod para on stomach + ovaries	empty

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEET

Recorder: Sue Nixtam / Claire Hrenchuk Date: August 13, 2011

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs 400 (g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
✓ TPN-06.02	OT/PEC	—	normal	—	—	—	slightly lightly colored pic 3739	normal	light (low) stomach	LKTR, chironomids 40% ↓ few
✓ TPN-06.20	OT/PEC	—	normal	—	—	—	normal	normal	low stomach	10%, 200 full
✓ TPN-06.15	OT/PEC	—	normal	—	—	—	normal	deformed (pinched) swim bladder	NONE	10% full, 200
✓ TPN-06.17	OT/PEC	—	normal	—	—	—	normal	none	none	empty
✓ TPN-06.19	OT/PEC	—	normal	—	—	—	normal	normal	light (one)	10% full, 200
✓ TPN-06.21	OT/PEC	—	normal	—	—	—	1 parasite cyst 3741, 3742	normal	light (one)	200 full Sculpin 1.5cm
✓ TPN-06.23	OT/PEC	—	normal	—	—	—	1 parasite of discoloration 3743, 3744	normal w parasites	mod stomach + tapeworm	10%, 200
✓ TPN-06.18	OT/PEC	—	normal	—	—	—	normal	normal	mod stomach cysts	10% + 200
✓ TPN-07.01	OT/PEC	—	normal	—	—	—	normal	normal w parasites	low light	10%, ephem. fish, chiron
✓ TPN-05.21	OT/PEC	—	normal ★ very large eggs - current year	—	—	—	normal	normal w parasites	low light	20%, fish remains
✓ TPN-08.002	OT/PEC	—	normal	—	—	—	normal	normal	none	5%, pupa, larval fish
✓ TPN-08.003	OT/PEC	—	normal	—	—	—	normal	normal	none	60%: chiron, trichop
✓ TPN-08.006	OT/PEC	—	normal	—	—	—	normal	normal	none	20%: chiron, sculpin x2
✓ TPN-08.007	OT/PEC	—	normal	—	—	—	normal	normal	none	5%: chiron

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEET

Recorder: Mike

Date: Aug 14/11

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs <u>400(g)</u>	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
TPN-11.01	pect oto	—	Normal	—	—	—	Normal	Normal	moderate gut para	10% fish remains
TPN-11.02	pect oto	—	Normal	—	—	—	Normal (cyst)	Normal	low gut para	25% fish remains
TPN-11.03	PECT oto	—	Normal	—	—	—	Normal Faint sm. white embedded cyst	Normal	low para in gut	410% (22%) ephem, chiron
TPN-11.05	pect oto	—	Normal	—	—	—	Normal	Normal	low gut para	60% fish + chiron
TPN-11.06	pect oto	29.6	Normal	1.02	—	8.20	Normal	Normal	No parasites	70% chiron, ephem, ostracod
TPN-11.08	pect oto	—	Normal	—	—	—	Normal	Normal	No parasites	20% sculpin
TPN-11.09	PECT oto	—	Normal	—	—	—	Normal 1 liver cyst	Normal	No parasites	90% chiron, trich., fish
TPN-10.03	PEL/OT	—	Normal	—	—	—	Normal	Normal none	low 1 cyst	50% Ephem
TPN-10.04	PEL/OT	—	"	—	—	—	"	None	None	5% chiron.
TPN-10.05	PEL/OT	—	—	—	—	—	—	—	—	—
TPN-10.08	pect oto	19.50 male	Normal	—	—	4.97	Normal	lots fat Normal	No parasite	10% Ephem + chiron.
TPN-10.09	pect oto	—	Normal	—	—	—	Normal	Normal	low para	10% zoop
TPN-10.10	pect oto	—	Normal	—	—	—	Normal	Normal	low para	MT
TPN-10.12	pect otolith	—	Normal	—	—	—	Normal	Normal	low gut para	80% zoop.

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEET

Recorder: Mike

Date: Aug 14/11

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs 900 (g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
TPN-10.13	pect oto	-	Normal	-	-	-	Normal	Normal	low parasite	10% chiron
TPN-10.14	pect oto	-	"	-	-	-	"	"	low parasite	90% fish remains
TPN-10.15	oto	-	Normal	-	-	-	Normal	Normal	None observed	80% zoopl.
TPN-10.16	oto	-	Normal	-	-	-	Normal	Normal	low parasite	50% zoopl.
TPN-10.19	pect oto	-	Normal	-	-	-	Normal	Normal	low gut parasite	40% zoopl.
TPN-10.20	pect oto	-	Normal	-	-	-	Normal	Normal	low parasite	MT
TPN-10.21	pect/oto	-	Normal	-	-	-	Normal	Normal	None	30% fish remain + chiron.
TPN-10.22	pect/oto	-	Testing normal	-	-	-	Normal	low gut parasite	low gut parasite	50% sculpin, trichop-
TPN-10.23	pect/oto	-	Normal testing	-	-	-	Normal	Normal	low gut parasite	10% chiron.
TPN-10.25	pect oto	-	Normal testing	-	-	-	Normal	Normal	low gut parasite	5% chiron
TPN-10.26	pect oto	-	Normal testing	-	-	-	Normal	Normal	low gut parasite	80% fish remains + chiron
<hr/>										
TPN-17.01	pect oto	-	Normal testing	-	-	-	Normal	Normal	low gut parasite	5% fish remains
TPN-16.04	pect/oto	-	"	-	-	-	"	"	low gut parasites	MT

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEET

Recorder: Laura

Date: Aug 15 2011

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs (g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
TPN-11.10	PEC/OT	—	normal	—	—	—	normal	normal	low	10% fish remains.
✓ TPN-16.06	PEC/OT	—	normal	—	—	—	normal	normal	low	20% fish
✓ TPN-16.21	PEC/OT	—	normal	—	—	—	normal	normal	Moderate	100% LKTR remains
✓ TPN-16.22	PEC/OT	—	normal	—	—	—	normal	normal	none	5% inverts.
✓ TPN-16.09	PEC/OT	—	normal	—	—	—	normal	normal	low	MT.
✓ TPN-16.12	PEC/OT	—	normal	—	—	—	normal	normal	low	20% chironomids/inverts.
✓ TPN-16.13	PEC/OT	—	normal	—	—	—	normal	normal	low	MT.
✓ TPN-16.14	PEC/OT	—	normal	—	—	—	normal	normal	low	30% fish/chironomids
✓ TPN-16.17	PEC/OT	—	normal	—	—	—	normal	normal	low	10% inverts (diptera)
✓ TPN-16.29	PEC/OT	—	normal	—	—	—	normal	normal	moderate	50% inverts. (chiron/diptera)
✓ TPN-16.25	PEC/OT	—	normal	—	—	—	normal	normal	low	MT.
✓ TPN-16.26	PEC/OT	—	normal	—	—	—	normal	normal	low	20% fish + invert remains.
✓ TPN-16.27	PEC/OT	—	normal	—	—	—	normal	normal	low	MT
✓ TPN-16.28	PEC/OT	—	normal	*wt of all eggs: 1664g	—	—	3759, 3760, 3761. (photos) A	normal	low	MT photo 3762

Age Structure Codes:

3754 - 3758

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

slightly
lightly colored

5/18

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEET

Recorder: Claire

Date: Aug 15/11

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs AOD (g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
✓ TPN-22.02	PEC/OT	—	Normal	—	—	—	Normal	Normal	None	20%; ^{DRIB} fish
✓ TPN-22.03	PEC/OT	—	NORMAL	—	—	—	NORMAL	NORMAL	1 cyst	20%; chiron
✓ TPN-23.01	PEC/OT	—	NORMAL	—	—	—	Parasite CYST	NORMAL	1 cyst cliver	80%; zoo

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEETRecorder: Susan HertamDate: August 16, 2011

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs 400 (g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
TPN-24.04	PEC ₁ OT	—	normal	—	—	—	normal	light parasites. stomach.	light cysts.	10% Trichopteran wings pupae.
TPN-25.07	OT	—	normal.	—	—	—	1 cyst/ tumor.	normal	none.	10% zoa
TPN-25.03	OT	—	normal	—	—	—	normal	normal	mod- heavy cysts.	empty.
TPN-25.02	PEC ₁ OT	—	normal	—	—	—	normal	normal	low	20% trichopterans
TPN-25.01	PEC ₁ OT	—	normal	—	—	—	normal	normal	low cysts	50% fish remains. + chironomids.
TPN-24.05	PEC ₁ OT	—	normal	—	—	—	light mottled	normal	low cysts.	MT

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OT = Otolith

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEETRecorder: LauraDate: Aug 17/11

entered MJ

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs (g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
✓ INUG-01.12	OTO/PEC	24.64	normal	0.89		7.25	normal	none	none	100% clam shrimp chironomid zooplankton; ostracods
✓ INUG-01.13	OTO/PEC	22.20	normal	0.94		8.73	normal	none	none	80% "
✓ INUG-01.14	OTO/PEC	11.13	normal	0.97		4.38	normal	none	none	100% "
✓ INUG-01.15	OTO/PEC	3.29	normal	—	—	1.60	normal	none	none	60% "
✓ INUG-01.18	OTO/PEC	—	normal	—	—	13.75	normal	none	low	MT
✓ INUG-01.16	OTO/PEC	—	normal	—	—	11.44	normal	none	high	MT
✓ INUG-01.17	OTO/PEC	31.24	normal	—	—	11.39	normal	none	low	70% clam shrimp
✓ INUG-01.18	OTO/PEC	30.91	normal	0.99		9.98	normal	none	none	MT
✓ INUG-01.19	OTO/PEC	32.28	normal	—	—	8.14	normal	none	none	10% clam shrimp
✓ INUG-01.20	OTO/PEC	22.03	normal	—	—	2.21	normal	none	none	50% clam shrimp ostracods
✓ INUG-01.21	OTO/PEC	25.02	normal	—	—	2.58	normal	none	none	MT
✓ INUG-01.22	OTO/PEC	16.86	normal	0.97		2.65	normal	none	none	50% clam shrimp

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEET

Recorder: Laura

Date: Aug 17/11

entered SH

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs ADD (g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
✓ INUG-02.15	OTO/PEC	0.42	normal	—	—	5.06	normal	none	moderate	10% fish remains (salmonids)
✓ INUG-02.14	OTO/PEC	33.69	normal	1.25	—	10.06	normal	none	no.	100% full clam shrimp chironomids/clams
✓ INUG-02.06	OTO/PEC	too small	normal	—	—	1.45	normal	none	low	50% full fish remains / clams
✓ INUG-02.05	OTO/PEC	too small	normal	—	—	0.86	a couple parasitic cysts	none.	low	MT
✓ INUG-02.04	OTO/PEC	0.07	normal	—	—	0.79	normal	none	no.	80% clam shrimp.
✓ INUG-02.13	OTO/PEC	0.36	normal	—	—	2.15	normal	none	low	40% zooplankton
✓ INUG-02.08	OTO/PEC	1.15	normal	—	—	2.26	normal	none.	none	80% clams, chironomids. clam shrimp
✓ INUG-02.21	OTO/PEC	too small	normal	—	—	0.45	normal	none	moderate	70% chironomids + sculpin
✓ INUG-02.16	OTO/PEC	35.53	normal	—	—	59.68	normal	none	low	90% LKTR. (1 large) 1 other fish (small)
✓ INUG-02.17	OTO/PEC	1.69	normal	—	—	14.49	normal	none	moderate	40% fish remains
✓ INUG-02.18	OTO/PEC	12.10	normal	—	—	20.44	normal	none	low	100% clam shrimp fish, ephemeroptera ostracods
✓ INUG-01.09	OTO/PEC	—	normal	—	—	12.0	normal	none	moderate	MT
✓ INUG-01.10	OTO/PEC	—	normal	—	—	3.26	normal	none	low	80% clam shrimp
✓ INUG-01.11	OTO/PEC	23.72	normal	—	—	5.13	normal	none	none	70% clam shrimp chironom, zooplankton, ostracods.

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

9/18

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEETRecorder: Claire HrenchukDate: Aug 17/11

entered 8/11/11

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs 400 (g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
✓ INUG-05.01	PEC/OT	23.09	NORMAL	0.92	—	8.23	NORMAL	NORMAL	NONE	50% ; clam shrimp
✓ INUG-03.01	PEC/OT	19.23	NORMAL	0.91	—	6.85	NORMAL	NORMAL	NONE	30% ; cladocera clam shrimp
✓ INUG-04.03	PEC/OT	—	NORMAL	—	—	4.15	NORMAL	NORMAL	MOD cysts	20% fish remains
✓ INUG-05.05	PEC/OT	—	NORMAL	—	—	14.84	NORMAL	NORMAL	MODERATE	EMPTY
✓ INUG-05.06	PEC/OT	—	NORMAL	—	—	20.73	slightly light coloured	NORMAL	NONE	30% ; Fish remains wasp (Chrysomelidae)
✓ INUG-05.07	PEC/OT	—	NORMAL	—	—	68.46	Discoloured + blotchy	NORMAL	MODERATE	80% ; Fish remains
✓ INUG-05.08	PEC/OT	—	NORMAL	—	—	13.00	NORMAL	NORMAL	HEAVY	20% ; Clam shrimp

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

Aug 18 / 11										
Fish Sample ID	Age Struct	Gonad Weight	Gonad App	Weight ⁴⁰⁰ eggs	total	Liver Weight	Liv app	Int Delt	Parasites	Stomach
✓ INUG-07.09	PEC/OT	-	NORMAL	-	-	2.18	Normal	NORMAL	LOW	EMPTY
✓ INUG-07.10	PEC/OT	-	NORMAL	-	-	1.24	NORMAL	NORMAL	LOW	EMPTY
✓ INUG-07.12	PEC/OT	-	NORMAL	-	-	1.32	NORMAL	NONE	Moderate	EMPTY
✓ INUG-07.01	PEC/OT	-	NORMAL	-	-	14.24	NORMAL	NONE	LOW	50%; clam shrimp
✓ INUG-07.03	PEC/OT	-	NORMAL	-	-	8.63	NORMAL	NONE	LOW	100% (pic=3769); mammal
✓ INUG-07.06	PEC/OT	-	NORMAL	-	-	1.56	NORMAL	NONE	Moderate	90%; clam shrimp
✓ INUG-07.15	(one only) PEC/OT	-	NORMAL	-	-	0.46	NORMAL	NONE	HIGH	20%; zoo
✓ INUG-08.06	PEC/OT	-	NORMAL	-	-	1.93	NORMAL	NONE	Moderate	EMPTY
✓ INUG-08.15	PEC/OT	-	NORMAL	-	-	1.49	NORMAL	NONE	LOW	30%; fish
✓ INUG-08.04	(one only) PEC/OT	20.21	NORMAL	1.06	-	6.54	NORMAL	NONE	NONE NONE	40%; clam shrimp chiron
✓ INUG-08.05	PEC/OT	21.40	normal	-	-	3.39	normal	none	none	60%; clam shrimp ostracods chir
✓ INUG-08.08	PEC/OT	15.71	NORMAL	-	-	4.07	NORMAL	NONE	NONE	60%; clam shrimp ostracods

Fish Sample	Age Struct	Gonad Weight	Gonad App	Weight of 400 eggs	totaleff	Liver Weight	Liv App	Int DELT	Parasites	Stomach
✓ INUG-08.09	PEC/OT	14.39	NORMAL	—	—	4.36	NORMAL	NONE	NONE	30%; clam shrimp, ostracod, chiron
✓ INUG-08.11	PEC/OT	15.85	NORMAL	0.79	—	4.19	NORMAL	NONE	NONE	30%; clam shrimp, ostracod, chiron
✓ INUG-08.12	PEC/OT	14.11	normal	6.73	—	6.88	normal	none	none	18%; clam shrimp
✓ INUG-08.13	PEC/OT	14.11	normal	—	—	1.10	normal	none	none	18%; chiron
✓ INUG-08.14	PEC/OT	14.11	normal	—	—	1.81	normal	none	none	20%; clam shrimp, ostracod
✓ INUG-08.17	PEC/OT	42.79	NORMAL	—	—	11.20	NORMAL	NONE	MODERATE	50%; clam shrimp, ostracods
✓ INUG-08.18	PEC/OT	—	NORMAL	—	—	19.25	NORMAL	NONE	MODERATE	40%; fish (trout)
✓ INUG-08.19	PEC/OT	—	NORMAL	—	—	18.81	NORMAL	NONE	MODERATE	30%; clam shrimp
✓ INUG-08.20	PEC/OT	194.43	NORMAL	35.08	—	34.39	NORMAL	NONE	LOW	EMPTY
✓ INUG-08.16	PEC/OT	4.06	normal	—	—	3.65	normal	none	none	10% clam shrimp
✓ INUG-10.04	PEC/OT	35.08	NORMAL	—	—	16.49	NORMAL	NONE	LOW	EMPTY
✓ INUG-10.02	PEC/OT	191.21	NORMAL	29.07	—	40.93	NORMAL	NONE	LOW	20%; clam shrimp
INUG-09.04	PEC/OT	27.82	normal	—	—	4.16	normal	none	none	20%; clam shrimp, ostracods, chiron

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEETRecorder: Claire HrenchukDate: Aug 19/11

entered SH, MJ

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs 400 (g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
✓ INUG-14.13	PEC/OT	-	NORMAL	-	-	44.09	NORMAL	NONE	LOW HIGH	EMPTY
✓ INUG-14.15	PEC/OT	2260	NORMAL	29.57	-	42.75	NORMAL	NONE	LOW	50%; clam shrimp
✓ INUG-14.16	PEC/OT	-	NORMAL	-	-	16.11	normal	none	low	60%; clam shrimp
✓ INUG-14.18	PEC/OT	-	NORMAL	-	-	17.06	NORMAL	NONE	LOW	40%; fish (burbot)
✓ INUG-14.19	PEC/OT	-	NORMAL	-	-	8.68	NORMAL	NONE	LOW	50%; clam shrimp
✓ INUG-14.11	PEC/OT	15.11	NORMAL	-	-	3.34	NORMAL	NONE	NONE	70%; chiron
✓ INUG-14.14	PEC/OT	14.75	NORMAL	-	-	3.32	NORMAL	NONE	NONE	10%; clam shrimp, chiron
✓ INUG-14.01	PEC/OT	-	NORMAL	-	-	2.54	NORMAL	NONE	MODERATE	20%; fish, tape worm, 200
✓ INUG-14.03	PEC/OT	-	NORMAL	-	-	5.37	NORMAL	NONE	LOW	20%; fish (Sculpin)
✓ INUG-14.05	PEC/OT	-	NORMAL	-	-	1.34	NORMAL	NONE	LOW	10%; fish
✓ INUG-14.02	PEC/OT	27.85	NORMAL	* 1.81 PIC 3782, 378	-	10.01	NORMAL	NONE	LOW	30%; clam shr, ostracods
✓ INUG-14.08	PEC/OT	12.54	NORMAL	-	-	3.96	NORMAL	NONE	NONE	20%; trichop, 200 → photo #3780, 378
✓ INUG-14.09	PEC/OT	16.42	NORMAL	-	-	5.46	NORMAL	NONE	NONE	30%; clam shr, ostracods
✓ INUG-14.10	PEC/OT	7.99	NORMAL	-	-	3.25	NORMAL	NONE	NONE	40%; clam shr, ostra trichop

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

13/18

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEET

Recorder: Claire

Date: Aug 19/11

entered SN+M1

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs 400(g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
✓ INUG-13.10	PEC/OT	—	NORMAL	—	—	4.54	NORMAL	NORMAL NONE	HIGH	30% clam shrimp
✓ INUG-13.09	PEC/OT	20.66	NORMAL	—	—	4.70	NORMAL	NORMAL NONE	NONE	EMPTY
✓ INUG-13.12	PEC/OT	22.18	NORMAL	0.47	—	5.53	NORMAL	NORMAL NONE	NONE	20% ; clam shrimp ostracods
✓ INUG-13.14	^{LONG} PEC/OT	12.68	NORMAL	—	—	4.27	NORMAL	NONE	NONE	EMPTY
✓ INUG-13.05	PEC/OT	—	NORMAL	—	—	41.68	NORMAL	NONE	LOW	EMPTY
✓ INUG-13.07	PEC/OT	—	NORMAL	—	—	16.56	NORMAL	NONE	MODERATE	30% ; clam shrimp
✓ INUG-13.02	PEC/OT	20.68	NORMAL	—	—	4.81	NORMAL	NONE	NONE	20% ; clam shrimp 200 inverts
✓ INUG-13.04	PEC/OT	28.80	NORMAL	—	—	3.0	NORMAL	NONE	NONE	10% ; trichop clam shrimp
✓ INUG-13.18	PEC/OT	23.62	NORMAL	—	—	4.62	NORMAL	NONE	NONE	20% ; clam shrimp ostracods
✓ INUG-13.17	PEC/OT	117.88	NORMAL	22.98	—	21.07	NORMAL	NONE	MODERATE	40% ; clam shrimp
✓ INUG-13.19	PEC/OT	17.00	NORMAL	0.85	—	5.37	NORMAL	NONE	LOW	40% ; clam shrimp 200

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

14/18

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEET

Recorder: Claire

Date: Aug 17/11
entered SM, MJ

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 400 eggs (g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
✓ PD-01.03	PEC/OT	28.77	NORMAL	0.84	—	11.82	NORMAL	Lots of fat in body cavity NORMAL	NONE	80% ; Chiron, Zoo
✓ PD-01.04	PEC/OT	29.98	NORMAL	0.91	—	8.39	NORMAL	LOTS of fat NORMAL	NONE	90% ; Chiron, Trichop
✓ PD-01.14	PEC/OT	—	normal	—	—	1.89	normal	none	none	80% ; Zoo plankton
✓ PD-01.15	PEC/OT	—	NORMAL	—	—	2.15	CYST	CYST IN LIVER	very low	70% ; Zoo
✓ PD-01.16	PEC/OT	—	NORMAL	—	—	0.93	NORMAL	NORMAL	LOW	90% ; Fish remains
✓ PD-01.17	PEC/OT	—	NORMAL	—	—	1.14	NORMAL	NORMAL NORMAL	MODERATE	90% ; Fish remains, chiron
✓ PD-02.01	PEC/OT	—	NORMAL	—	—	2.79	NORMAL	NONE	Low	EMPTY
✓ PD-02.02	PEC/OT	—	NORMAL	—	—	1.62	NORMAL	NONE	LOW	EMPTY
✓ PD-01.07	PEC/OT	17.76	NORMAL	0.76	—	8.84	NORMAL	Lots of fat NORMAL	NONE	90% ; trichop, chiron
✓ PD-01.08	PEC/OT	—	NORMAL	—	—	1.19	NORMAL	lots of fat NORMAL	NONE	EMPTY
✓ PD-01.06	PEC/OT	—	NORMAL	—	—	4.52	NORMAL	NORMAL	LOW	EMPTY
✓ PD-01.09	PEC/OT	—	NORMAL	—	—	1.70	NORMAL	NORMAL	MODERATE (CESTODES)	5% ; cestode
✓ PD-01.10	PEC/OT	—	NORMAL	—	—	1.93	Parasite cyst NORMAL	NORMAL	Low	20% ; unknown
✓ PD-01.11	PEC/OT	—	NORMAL	—	—	1.79	NORMAL	NORMAL	LOW	EMPTY

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

2011 EEM for Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEET

entered 8H, MJ

Recorder: Laura

Date: Aug 18/11

PIPEDREAM

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 400 eggs (g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
✓ PD-04.11	PEC/OTO	—	normal	—	—	0.50	normal	none	low	20% chironomids/ other inverts. UN
✓ PD-04.12	PEC/OTO	—	normal	—	—	0.68	pic. 3771 cyst	cyst on liver.	low	MT
✓ PD-04.13	PEC/OTO	—	normal	—	—	0.83	normal	none.	moderate	MT
✓ PD-04.08	PEC/OTO	—	normal	—	—	3.22	normal	none	moderate	100% fish LKTR. Remains.
✓ PD-04.09	PEC/OTO	—	normal	—	—	5.84	normal	none	low	80% fish remains (sculpin)
PD-04.14	PEC/OTO	—	normal	—	—	0.22	normal	none	none	MT
✓ PD-04.04	PEC/OTO	—	normal	—	—	20.51	normal	none	low	10% Sculpin. fish remains
✓ PD-04.05	PEC/OTO	75.14	pic 3773 normal	—	—	16.11	normal	none	low	20% inverts & fish remains (diptera) UN
✓ PD-04.06	PEC/OTO	236	normal	30.52	—	45.07	normal	none	low	10% inverts (diptera)
✓ PD-03.06	PEC/OTO	246	normal	—	—	1.79	normal	none.	low	10% zooplankton
✓ PD-03.12	PEC/OTO	2.22	normal	—	—	2.22	normal	none	low	30% zooplankton.
✓ PD-03.13	PEC/OTO	—	normal	—	—	1.73	normal	none.	low	empty.
✓ PD-03.14	PEC/OTO	—	normal	—	—	1.05	normal	none	mod	10% inverts (diptera)
✓ PD-03.02	PEC/OTO	—	normal	—	—	1.79	normal	none	low	60% fish remains.

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

16/18

2011 EEM Meadowbank Gold Mine: DETAILED BIOLOGICAL ANALYSIS LAB SHEET

Recorder: Susan

Date: August 18, 2011

inferred SK, MJ.

PIPEDREAM

Fish Sample ID (from field data sheet)	Age Structure	Gonad weight (g)	Gonad appearance	Weight of 100 eggs 400(g)	Total # of eggs	Liver weight (g)	Liver appearance	Internal DELT	Internal parasites?	Stomach contents (identifiable items; fullness)
✓ PD-03.03	PEC/OT	—	normal	—	—	2.33	normal	none	*cestodes low	50% ostracods, tapeworms.
✓ PD-03.08	PEC/OT	24.87	normal	—	—	4.62	normal	none	none	20% trichoptera, diptera.
✓ PD-03.04	PEC/OT	—	normal	—	—	9.91	1 parasite cyst	none	low	empty.
✓ PD-03.05	PEC/OT	71.31	normal	11.26	—	28.85	normal	none	low	empty
✓ PD-03.09	PEC/OT	—	normal	—	—	14.21	normal	none	low	empty.
✓ PD-03.18	PEC/OT	48.78	normal	—	—	5.93	normal	none	none	10% trichoptera.
✓ PD-03.15	PEC/OT	230	normal	27.33	—	49.74	normal	none	mod.	empty.
✓ PD-03.16	PEC/OT	—	normal	—	—	16.05	1 parasite cyst	none	low	5% diptera.
✓ PD-03.17	PEC/OT	76.63	normal	—	—	15.97	1 parasite cyst	none	none	empty.
✓ PD-03.20	PEC/OT	62.71	normal	—	—	19.23	normal	none	low	10% fish remains
✓ PD-08.01a	PEC/OT	390	normal	35.19	—	68.74	normal	none	mod.	empty.

Age Structure Codes:

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

17/18

Date: August 19, 2011

pic
3779
gonads

Clarke's
p/c 116, 117, 118
Sue's 3777
3778

PEL = Pelvic fin; PEC = Pectoral fin; DR = Dorsal ray; OTO = Otolith

18/18

APPENDIX H

FISH DATABASE



Lake	Set ID	Date	Mesh Size (mm)	Day/Night	Species	Sample ID	Mortality	Length.mm	Weight.g	K
TPN	TPN-03	11-Aug-11	38	Day	ARCH	TPN-03.03	N	214	100	1.020372346
TPN	TPN-05a	13-Aug-11	51	Night	ARCH	TPN-05.07	Y	272	190	0.944162808
TPN	TPN-05a	13-Aug-11	51	Night	ARCH	TPN-05.09	Y	275	216	1.038617581
TPN	TPN-05a	13-Aug-11	51	Night	ARCH	TPN-05.11	Y	338	390	1.009984029
TPN	TPN-06a	13-Aug-11	102	Night	ARCH	TPN-06.04	N	581	2025	1.03251562
TPN	TPN-06a	13-Aug-11	102	Night	ARCH	TPN-06.06	N	576	2150	1.12504789
TPN	TPN-06a	13-Aug-11	76	Night	ARCH	TPN-06.08	Y	345	454	1.105601369
TPN	TPN-06a	13-Aug-11	51	Night	ARCH	TPN-06.11	Y	488	1125	0.96803945
TPN	TPN-06a	13-Aug-11	38	Night	ARCH	TPN-06.17	Y	201	78	0.96052004
TPN	TPN-06a	13-Aug-11	38	Night	ARCH	TPN-06.19	Y	218	90	0.868706415
TPN	TPN-06a	13-Aug-11	38	Night	ARCH	TPN-06.20	Y	257	146	0.860108988
TPN	TPN-06a	13-Aug-11	25	Night	ARCH	TPN-06.24	N	115		
TPN	TPN-06a	13-Aug-11	25	Night	ARCH	TPN-06.25	N	136		
TPN	TPN-06b	13-Aug-11	102	Day	ARCH	TPN-06.26	N	577	2125	1.1061945
TPN	TPN-10	14-Aug-11	25	Night	ARCH	TPN-10.02	N	135	25	1.016105269
TPN	TPN-10	14-Aug-11	25	Night	ARCH	TPN-10.03	Y	126	20	0.999812035
TPN	TPN-10	14-Aug-11	25	Night	ARCH	TPN-10.04	Y	134	24	0.997463119
TPN	TPN-10	14-Aug-11	25	Night	ARCH	TPN-10.05	Y	135	20	0.812884215
TPN	TPN-10	14-Aug-11	38	Night	ARCH	TPN-10.07	N	580	2050	1.050678585
TPN	TPN-10	14-Aug-11	51	Night	ARCH	TPN-10.12	Y	310	300	1.007015542
TPN	TPN-10	14-Aug-11	51	Night	ARCH	TPN-10.15	Y	302	294	1.067398287
TPN	TPN-10	14-Aug-11	51	Night	ARCH	TPN-10.16	Y	256	168	1.001358032
TPN	TPN-10	14-Aug-11	51	Night	ARCH	TPN-10.17	N	340	450	1.144921636
TPN	TPN-11a	14-Aug-11	76	Night	ARCH	TPN-11.07	N	378	650	1.20347745
TPN	TPN-16a	14-Aug-11	126	Day	ARCH	TPN-16.01	N	604	2425	1.100527571
TPN	TPN-16b	15-Aug-11	102	Night	ARCH	TPN-16.19	N	558	1800	1.03602422
TPN	TPN-23	15-Aug-11	76	Day	ARCH	TPN-23.01	Y	596	1926	0.909740446
TPN	TPN-24b	16-Aug-11	51	Night	ARCH	TPN-24.03	N	284	225	0.98226235
TPN	TPN-25	16-Aug-11	51	Night	ARCH	TPN-25.03	Y	284	236	1.030284065
TPN	TPN-25	16-Aug-11	51	Night	ARCH	TPN-25.06	N	278	230	1.070516704
TPN	TPN-25	16-Aug-11	51	Night	ARCH	TPN-25.07	Y	233	122	0.964477427
TPN	TPN-25	16-Aug-11	76	Night	ARCH	TPN-25.09	N	585	1675	0.836656546
TPN	TPN-25	16-Aug-11	76	Night	ARCH	TPN-25.11	N	416	825	1.145971869
TPN	TPN-01	11-Aug-11	102	Day	LKTR	TPN-01.01	Y	567	1800	0.987468677
TPN	TPN-01	11-Aug-11	76	Day	LKTR	TPN-01.02	N	560	1700	0.968021137
TPN	TPN-01	11-Aug-11	51	Day	LKTR	TPN-01.03	N	517	1400	1.013109543
TPN	TPN-02	11-Aug-11	51	Day	LKTR	TPN-02.02	N	498	1225	0.991854711
TPN	TPN-02	11-Aug-11	25	Day	LKTR	TPN-02.04	N	123	50	2.686919592
TPN	TPN-03	11-Aug-11	76	Day	LKTR	TPN-03.01	N	527	1700	1.161494281
TPN	TPN-03	11-Aug-11	51	Day	LKTR	TPN-03.02	N	575	2050	1.078326621
TPN	TPN-05a	13-Aug-11	126	Night	LKTR	TPN-05.01	Y	518	1418	1.020203835
TPN	TPN-05a	13-Aug-11	126	Night	LKTR	TPN-05.02	Y	503	1150	0.903636751
TPN	TPN-05a	13-Aug-11	126	Night	LKTR	TPN-05.03	Y	392	605	1.004377959
TPN	TPN-05a	13-Aug-11	76	Night	LKTR	TPN-05.04	N	567	1425	0.781746036
TPN	TPN-05a	13-Aug-11	76	Night	LKTR	TPN-05.05	N	690	3100	0.943657556
TPN	TPN-05a	13-Aug-11	76	Night	LKTR	TPN-05.06	N	345	400	0.974098122
TPN	TPN-05a	13-Aug-11	51	Night	LKTR	TPN-05.08	Y	190	50	0.728969237
TPN	TPN-05a	13-Aug-11	51	Night	LKTR	TPN-05.10	N	294	225	0.885402058
TPN	TPN-05a	13-Aug-11	51	Night	LKTR	TPN-05.12	Y	250	144	0.9216
TPN	TPN-05a	13-Aug-11	51	Night	LKTR	TPN-05.13	Y	301	278	1.019401596
TPN	TPN-05a	13-Aug-11	51	Night	LKTR	TPN-05.14	N	190	50	0.728969237
TPN	TPN-05a	13-Aug-11	51	Night	LKTR	TPN-05.15	Y	229	118	0.982597286
TPN	TPN-05a	13-Aug-11	25	Night	LKTR	TPN-05.16	Y	120		
TPN	TPN-05a	13-Aug-11	25	Night	LKTR	TPN-05.17	N	126	50	2.499530088
TPN	TPN-05a	13-Aug-11	25	Night	LKTR	TPN-05.18	Y	130		
TPN	TPN-05a	13-Aug-11	25	Night	LKTR	TPN-05.19	N	771	5000	1.090955083
TPN	TPN-05b	13-Aug-11	38	Day	LKTR	TPN-05.20	N	748	4450	1.06329854
TPN	TPN-05b	13-Aug-11	102	Day	LKTR	TPN-05.21	Y	506	1620	1.250441746
TPN	TPN-05c	13-Aug-11	51	Day	LKTR	TPN-05.23	N	539	1625	1.037736446
TPN	TPN-06a	13-Aug-11	126	Night	LKTR	TPN-06.01	N	565	1525	0.845521198
TPN	TPN-06a	13-Aug-11	126	Night	LKTR	TPN-06.02	Y	664	2944	1.005619225
TPN	TPN-06a	13-Aug-11	102	Night	LKTR	TPN-06.03	Y	510	1438	1.084047614
TPN	TPN-06a	13-Aug-11	102	Night	LKTR	TPN-06.05	Y	538	1686	1.082706498
TPN	TPN-06a	13-Aug-11	102	Night	LKTR	TPN-06.07	N	535	1600	1.044861282
TPN	TPN-06a	13-Aug-11	51	Night	LKTR	TPN-06.09	N	551	1750	1.046124208
TPN	TPN-06a	13-Aug-11	51	Night	LKTR	TPN-06.10	N	425	700	0.911866477
TPN	TPN-06a	13-Aug-11	51	Night	LKTR	TPN-06.12	N	606	1925	0.864993535
TPN	TPN-06a	13-Aug-11	51	Night	LKTR	TPN-06.13	N	547	1775	1.084517036
TPN	TPN-06a	13-Aug-11	51	Night	LKTR	TPN-06.14	Y	266	178	0.945747261
TPN	TPN-06a	13-Aug-11	38	Night	LKTR	TPN-06.15	Y	231	126	1.022197008
TPN	TPN-06a	13-Aug-11	38	Night	LKTR	TPN-06.16	N	609	2450	1.084711405
TPN	TPN-06a	13-Aug-11	38	Night	LKTR	TPN-06.18	Y	176	54	0.990502911
TPN	TPN-06a	13-Aug-11	38	Night	LKTR	TPN-06.21	Y	169	50	1.035881055
TPN	TPN-06a	13-Aug-11	38	Night	LKTR	TPN-06.22	N	536	1525	0.990319828
TPN	TPN-06a	13-Aug-11	38	Night	LKTR	TPN-06.23	Y	203	74	0.884593219
TPN	TPN-06b	13-Aug-11	51	Day	LKTR	TPN-06.27	N	565	1875	1.039575243
TPN	TPN-06c	13-Aug-11	76	Day	LKTR	TPN-06.28	N	525	1475	1.019328366
TPN	TPN-06c	13-Aug-11	51	Day	LKTR	TPN-06.29	N	562	1725	0.97180729
TPN	TPN-06c	13-Aug-11	38	Day	LKTR	TPN-06.30	N	668	2900	0.972900912

Lake	Set ID	Date	Mesh Size (mm)	Day/Night	Species	Sample ID	Mortality	Length.mm	Weight.g	K
TPN	TPN-07a	13-Aug-11	76	Day	LKTR	TPN-07.01	Y	534	1474	0.967996334
TPN	TPN-08a	13-Aug-11	76	Day	LKTR	TPN-08.02	Y	503	1464	1.150368872
TPN	TPN-08a	13-Aug-11	51	Day	LKTR	TPN-08.03	Y	548	1720	1.045169605
TPN	TPN-08b	13-Aug-11	102	Day	LKTR	TPN-08.04	N	569	1950	1.058516939
TPN	TPN-08b	13-Aug-11	76	Day	LKTR	TPN-08.05	N	575	2025	1.065176297
TPN	TPN-08b	13-Aug-11	76	Day	LKTR	TPN-08.06	Y	495	1194	0.984438817
TPN	TPN-08b	13-Aug-11	51	Day	LKTR	TPN-08.07	Y	633	2470	0.973835996
TPN	TPN-08b	13-Aug-11	38	Day	LKTR	TPN-08.08	N	915	9050	1.181368256
TPN	TPN-08b	13-Aug-11	38	Day	LKTR	TPN-08.09	N	549	1875	1.133141744
TPN	TPN-10	14-Aug-11	25	Night	LKTR	TPN-10.01	N	579	1900	0.978853977
TPN	TPN-10	14-Aug-11	38	Night	LKTR	TPN-10.06	N	535	1500	0.979557452
TPN	TPN-10	14-Aug-11	38	Night	LKTR	TPN-10.09	Y	222	102	0.932269011
TPN	TPN-10	14-Aug-11	38	Night	LKTR	TPN-10.10	Y	186	61	0.947962161
TPN	TPN-10	14-Aug-11	51	Night	LKTR	TPN-10.11	N	530	1750	1.175466996
TPN	TPN-10	14-Aug-11	51	Night	LKTR	TPN-10.13	Y	385	574	1.005841856
TPN	TPN-10	14-Aug-11	51	Night	LKTR	TPN-10.14	Y	252	162	1.012309686
TPN	TPN-10	14-Aug-11	51	Night	LKTR	TPN-10.18	N	268	170	0.88317047
TPN	TPN-10	14-Aug-11	51	Night	LKTR	TPN-10.19	Y	252	162	1.012309686
TPN	TPN-10	14-Aug-11	51	Night	LKTR	TPN-10.20	Y	179	50	0.871788049
TPN	TPN-10	14-Aug-11	76	Night	LKTR	TPN-10.21	Y	203	80	0.956316994
TPN	TPN-10	14-Aug-11	76	Night	LKTR	TPN-10.22	Y	502	1406	1.111409665
TPN	TPN-10	14-Aug-11	76	Night	LKTR	TPN-10.23	Y	569	1944	1.055259964
TPN	TPN-10	14-Aug-11	76	Night	LKTR	TPN-10.24	N	467	1200	1.178231432
TPN	TPN-10	14-Aug-11	102	Night	LKTR	TPN-10.25	Y	601	2214	1.019892036
TPN	TPN-10	14-Aug-11	102	Night	LKTR	TPN-10.26	Y	535	1424	0.929926541
TPN	TPN-11a	14-Aug-11	51	Night	LKTR	TPN-11.01	Y	255	160	0.964938071
TPN	TPN-11a	14-Aug-11	51	Night	LKTR	TPN-11.02	Y	513	1350	0.999957802
TPN	TPN-11a	14-Aug-11	51	Night	LKTR	TPN-11.03	Y	561	1735	0.982677235
TPN	TPN-11a	14-Aug-11	51	Night	LKTR	TPN-11.04	N	224	150	1.334587965
TPN	TPN-11a	14-Aug-11	76	Night	LKTR	TPN-11.05	Y	543	1670	1.043078473
TPN	TPN-11a	14-Aug-11	76	Night	LKTR	TPN-11.08	Y	503	1408	1.106365691
TPN	TPN-11a	14-Aug-11	76	Night	LKTR	TPN-11.09	Y	432	806	0.999732479
TPN	TPN-11c	14-Aug-11	51	Night	LKTR	TPN-11.10	Y	268	153	0.794853423
TPN	TPN-11c	14-Aug-11	51	Night	LKTR	TPN-11.11	N	652	3225	1.16355504
TPN	TPN-11c	14-Aug-11	51	Night	LKTR	TPN-11.12	N	608	2400	1.067826031
TPN	TPN-14	14-Aug-11	76	Day	LKTR	TPN-14.01	N	497	1260	1.026363925
TPN	TPN-16a	14-Aug-11	102	Day	LKTR	TPN-16.02	N	629	2125	0.85389997
TPN	TPN-16a	14-Aug-11	76	Day	LKTR	TPN-16.03	N	590	1750	0.852083222
TPN	TPN-16a	14-Aug-11	76	Day	LKTR	TPN-16.04	Y	585	2000	0.99899289
TPN	TPN-16b	15-Aug-11	38	Night	LKTR	TPN-16.05	N	175	60	1.119533528
TPN	TPN-16b	15-Aug-11	38	Night	LKTR	TPN-16.06	Y	184	58	0.931053259
TPN	TPN-16b	15-Aug-11	51	Night	LKTR	TPN-16.07	N	275	190	0.913598798
TPN	TPN-16b	15-Aug-11	76	Night	LKTR	TPN-16.08	N	502	1450	1.146190622
TPN	TPN-16b	15-Aug-11	76	Night	LKTR	TPN-16.09	Y	610	1625	0.715918954
TPN	TPN-16b	15-Aug-11	76	Night	LKTR	TPN-16.10	N	562	1850	1.042228108
TPN	TPN-16b	15-Aug-11	76	Night	LKTR	TPN-16.11	N	536	1725	1.120197838
TPN	TPN-16b	15-Aug-11	76	Night	LKTR	TPN-16.12	Y	492	1226	1.029426069
TPN	TPN-16b	15-Aug-11	76	Night	LKTR	TPN-16.13	Y	560	1732	0.986242711
TPN	TPN-16b	15-Aug-11	76	Night	LKTR	TPN-16.14	Y	484	1258	1.109544069
TPN	TPN-16b	15-Aug-11	76	Night	LKTR	TPN-16.15	N	757	3825	0.881746615
TPN	TPN-16b	15-Aug-11	76	Night	LKTR	TPN-16.17	Y	514	1438	1.058935552
TPN	TPN-16b	15-Aug-11	76	Night	LKTR	TPN-16.18	N	752	4675	1.099330035
TPN	TPN-16b	15-Aug-11	102	Night	LKTR	TPN-16.20	N	515	1575	1.153078491
TPN	TPN-16b	15-Aug-11	102	Night	LKTR	TPN-16.21	Y	591	2100	1.017318289
TPN	TPN-16b	15-Aug-11	102	Night	LKTR	TPN-16.22	Y	496	1538	1.26040845
TPN	TPN-16b	15-Aug-11	126	Night	LKTR	TPN-16.23	N	624	2975	1.224427855
TPN	TPN-16b	15-Aug-11	126	Night	LKTR	TPN-16.24	N	544	1800	1.118087536
TPN	TPN-16b	15-Aug-11	126	Night	LKTR	TPN-16.25	Y	670	3122	1.038026619
TPN	TPN-16b	15-Aug-11	126	Night	LKTR	TPN-16.26	Y	548	1722	1.046384919
TPN	TPN-16b	15-Aug-11	126	Night	LKTR	TPN-16.27	Y	786	4384	0.902823609
TPN	TPN-16b	15-Aug-11	126	Night	LKTR	TPN-16.28	Y	961	10294	1.159882513
TPN	TPN-16b	15-Aug-11	76	Night	LKTR	TPN-16.29	Y	573	1782	0.947204677
TPN	TPN-17	14-Aug-11	76	Day	LKTR	TPN-17.01	Y	506	1394	1.075997403
TPN	TPN-20	15-Aug-11	51	Day	LKTR	TPN-20.01	N	241	150	1.071618324
TPN	TPN-22	15-Aug-11	76	Day	LKTR	TPN-22.01	N	643	2525	0.949791905
TPN	TPN-22	15-Aug-11	76	Day	LKTR	TPN-22.02	Y	598	2260	1.056829402
TPN	TPN-22	15-Aug-11	76	Day	LKTR	TPN-22.03	Y	494	1498	1.242598905
TPN	TPN-22	15-Aug-11	51	Day	LKTR	TPN-22.04	N	616	1600	0.684506925
TPN	TPN-22	15-Aug-11	38	Day	LKTR	TPN-22.05	N	600	2450	1.134259259
TPN	TPN-24b	16-Aug-11	51	Night	LKTR	TPN-24.01	N	267	200	1.050742289
TPN	TPN-24b	16-Aug-11	51	Night	LKTR	TPN-24.02	N	410	700	1.015655606
TPN	TPN-24b	16-Aug-11	51	Night	LKTR	TPN-24.04	Y	313	318	1.037036655
TPN	TPN-24b	16-Aug-11	76	Night	LKTR	TPN-24.05	Y	727	3806	0.990525251
TPN	TPN-25	16-Aug-11	51	Night	LKTR	TPN-25.01	Y	385	630	1.103972769
TPN	TPN-25	16-Aug-11	51	Night	LKTR	TPN-25.02	Y	346	392	0.94636304
TPN	TPN-25	16-Aug-11	51	Night	LKTR	TPN-25.04	N	593	1925	0.923138053
TPN	TPN-25	16-Aug-11	51	Night	LKTR	TPN-25.05	N	574	2175	1.150068171
TPN	TPN-25	16-Aug-11	51	Night	LKTR	TPN-25.08	N	638	3025	1.164832134
TPN	TPN-25	16-Aug-11	76	Night	LKTR	TPN-25.10	N	572	1725	0.921724249
TPN	TPN-10	14-Aug-11	38	Night	RNWH	TPN-10.08	Y	360	550	1.178840878
TPN	TPN-11a	14-Aug-11	76	Night	RNWH	TPN-11.06	Y	383	618	1.099998653

Lake	Set ID	Date	Weighed with 25 g scale	Sex	Maturity	Repro Status	Age Structure	Age.OTO.yr	Age.PEC.yr
TPN	TPN-03	11-Aug-11	Y						
TPN	TPN-05a	13-Aug-11	N	F	IM	UD	PEC, OTO	5	5
TPN	TPN-05a	13-Aug-11	N	M	IM	UD	PEC, OTO	5	4
TPN	TPN-05a	13-Aug-11	N	F	MA	RS	PEC, OTO	2	7
TPN	TPN-06a	13-Aug-11	Y				PEC		10
TPN	TPN-06a	13-Aug-11	Y				PEC		9
TPN	TPN-06a	13-Aug-11	N	F	MA	RS	PEC, OTO	8	7
TPN	TPN-06a	13-Aug-11	Y	F	MA	RS	PEC, OTO	9	8
TPN	TPN-06a	13-Aug-11	N	M	IM	UD	PEC, OTO	3	2
TPN	TPN-06a	13-Aug-11	N	F	IM	UD	PEC, OTO	4	2
TPN	TPN-06a	13-Aug-11	N	M	IM	UD	PEC, OTO	5	3
TPN	TPN-06a	13-Aug-11	N						
TPN	TPN-06b	13-Aug-11	Y						
TPN	TPN-10	14-Aug-11	N (± 1)						
TPN	TPN-10	14-Aug-11	N	UN	IM	UD	PEC, OTO	3	1
TPN	TPN-10	14-Aug-11	N	UN	IM	UD	PEC, OTO	4	2
TPN	TPN-10	14-Aug-11	N				PEC, OTO	3	2
TPN	TPN-10	14-Aug-11	Y				PEC		10
TPN	TPN-10	14-Aug-11	N	UN	IM	UD	OTO	6	
TPN	TPN-10	14-Aug-11	N	UN	IM	UD	OTO	5	
TPN	TPN-10	14-Aug-11	N	UN	IM	UD	OTO	4	
TPN	TPN-10	14-Aug-11	Y				PEC		6
TPN	TPN-11a	14-Aug-11	Y				PEC		5
TPN	TPN-16a	14-Aug-11	Y				PEC		9
TPN	TPN-16b	15-Aug-11	Y				PEC		7
TPN	TPN-23	15-Aug-11	N	F	MA	RS	OTO	11	
TPN	TPN-24b	16-Aug-11	Y				PEC		4
TPN	TPN-25	16-Aug-11	N	F	IM	UD	OTO	5	
TPN	TPN-25	16-Aug-11	N (± 10)				PEC		5
TPN	TPN-25	16-Aug-11	N	F	IM	UD	OTO	4	
TPN	TPN-25	16-Aug-11	Y				PEC		10
TPN	TPN-25	16-Aug-11	Y				PEC		8
TPN	TPN-01	11-Aug-11	Y	F	MA	RS	PEC, OTO	15	17
TPN	TPN-01	11-Aug-11	Y				PEC		13
TPN	TPN-01	11-Aug-11	Y				PEC		10
TPN	TPN-02	11-Aug-11	Y				PEC		11
TPN	TPN-02	11-Aug-11	Y						
TPN	TPN-03	11-Aug-11	Y				PEC		14
TPN	TPN-03	11-Aug-11	Y				PEC		13
TPN	TPN-05a	13-Aug-11	N	F	MA	RS	PEC, OTO	21	10
TPN	TPN-05a	13-Aug-11	N	M	MA	RS	PEC, OTO	19	15
TPN	TPN-05a	13-Aug-11	N	F	MA	RS	PEC, OTO	14	11
TPN	TPN-05a	13-Aug-11	Y				PEC		16
TPN	TPN-05a	13-Aug-11	Y				PEC		17
TPN	TPN-05a	13-Aug-11	Y				PEC		6
TPN	TPN-05a	13-Aug-11	N	M	IM	UD	PEC		3
TPN	TPN-05a	13-Aug-11	Y						
TPN	TPN-05a	13-Aug-11	N	F	IM	UD	PEC, OTO	6	4
TPN	TPN-05a	13-Aug-11	N	F	IM	UD	PEC, OTO	5	5
TPN	TPN-05a	13-Aug-11	Y				PEC		3
TPN	TPN-05a	13-Aug-11	N	M	IM	UD	PEC, OTO	3	4
TPN	TPN-05a	13-Aug-11	N	UN	IM	UD			
TPN	TPN-05a	13-Aug-11	Y				PEC		2
TPN	TPN-05a	13-Aug-11	N						
TPN	TPN-05a	13-Aug-11	Y				PEC		25
TPN	TPN-05b	13-Aug-11	Y				PEC		21
TPN	TPN-05b	13-Aug-11	N	F	MA	RI	PEC, OTO	15	15
TPN	TPN-05c	13-Aug-11	Y				PEC		17
TPN	TPN-06a	13-Aug-11	Y				PEC		17
TPN	TPN-06a	13-Aug-11	N	M	MA	RS	PEC, OTO	26	15
TPN	TPN-06a	13-Aug-11	N	F	MA	RS	PEC, OTO	18	14
TPN	TPN-06a	13-Aug-11	N	F	MA	RS	PEC, OTO	19	14
TPN	TPN-06a	13-Aug-11	Y				PEC		17
TPN	TPN-06a	13-Aug-11	Y				PEC		12
TPN	TPN-06a	13-Aug-11	Y				PEC		10
TPN	TPN-06a	13-Aug-11	Y				PEC		18
TPN	TPN-06a	13-Aug-11	Y				PEC		15
TPN	TPN-06a	13-Aug-11	N	M	IM	UD	PEC, OTO	12	8
TPN	TPN-06a	13-Aug-11	N	M	IM	UD	PEC, OTO	5	4
TPN	TPN-06a	13-Aug-11	Y				PEC		18
TPN	TPN-06a	13-Aug-11	N	M	IM	UD	PEC, OTO	6	ua
TPN	TPN-06a	13-Aug-11	N	F	IM	UD	PEC, OTO	6	3
TPN	TPN-06a	13-Aug-11	Y				PEC		15
TPN	TPN-06a	13-Aug-11	N	M	IM	UD	PEC, OTO	6	4
TPN	TPN-06b	13-Aug-11	Y						
TPN	TPN-06c	13-Aug-11	Y				Adipose		
TPN	TPN-06c	13-Aug-11	Y				PEC		14
TPN	TPN-06c	13-Aug-11	Y				PEC		17

Lake	Set ID	Date	Weighed with 25 g scale	Sex	Maturity	Repro Status	Age Structure	Age.OTO.yr	Age.PEC.yr
TPN	TPN-07a	13-Aug-11	N	UN	IM	UD	PEC, OTO	13	13
TPN	TPN-08a	13-Aug-11	N	M	MA	RI	PEC, OTO	19	15
TPN	TPN-08a	13-Aug-11	N	M	IM	UD	PEC, OTO	17	14
TPN	TPN-08b	13-Aug-11	Y				PEC		12
TPN	TPN-08b	13-Aug-11	Y				PEC		15
TPN	TPN-08b	13-Aug-11	N	M	IM	UD	PEC, OTO	22	11
TPN	TPN-08b	13-Aug-11	N	F	MA	RS	PEC, OTO	22	14
TPN	TPN-08b	13-Aug-11	Y				PEC		26
TPN	TPN-08b	13-Aug-11	Y				PEC		14
TPN	TPN-10	14-Aug-11	Y				PEC		22
TPN	TPN-10	14-Aug-11	Y				PEC		14
TPN	TPN-10	14-Aug-11	N	UN	IM	UD	PEC, OTO	7	4
TPN	TPN-10	14-Aug-11	N	UN	IM	UD	PEC, OTO	5	4
TPN	TPN-10	14-Aug-11	Y				PEC		11
TPN	TPN-10	14-Aug-11	N	UN	IM	UD	PEC, OTO	16	9
TPN	TPN-10	14-Aug-11	N	UN	IM	UD	PEC, OTO	13	6
TPN	TPN-10	14-Aug-11	N (± 10)				PEC		8
TPN	TPN-10	14-Aug-11	N	UN	IM	UD	PEC, OTO	6	4
TPN	TPN-10	14-Aug-11	N	UN	IM	UD	PEC, OTO	5	4
TPN	TPN-10	14-Aug-11	N	UN	IM	UD	PEC, OTO	4	3
TPN	TPN-10	14-Aug-11	N	F	MA	RS	PEC, OTO	18	9
TPN	TPN-10	14-Aug-11	N	M	MA	RS	PEC, OTO	23	17
TPN	TPN-10	14-Aug-11	Y				PEC		14
TPN	TPN-10	14-Aug-11	N	M	MA	RS	PEC, OTO	17	16
TPN	TPN-10	14-Aug-11	N	F	MA	RS	PEC, OTO	40	15
TPN	TPN-11a	14-Aug-11	N	UN	IM	UD	PEC, OTO	8	5
TPN	TPN-11a	14-Aug-11	N	M	MA	RI	PEC, OTO	21	14
TPN	TPN-11a	14-Aug-11	N	M	MA	RS	PEC, OTO	19	14
TPN	TPN-11a	14-Aug-11	N (± 10)				PEC		5
TPN	TPN-11a	14-Aug-11	N	M	MA	RS	PEC, OTO	26	13
TPN	TPN-11a	14-Aug-11	N	F	MA	RI	PEC, OTO	20	13
TPN	TPN-11a	14-Aug-11	N	F	MA	RS	PEC, OTO	15	11
TPN	TPN-11c	14-Aug-11	N	UN	IM	UD	PEC, OTO	7	7
TPN	TPN-11c	14-Aug-11	Y				PEC		20
TPN	TPN-11c	14-Aug-11	Y				PEC		17
TPN	TPN-14	14-Aug-11	Y				PEC		14
TPN	TPN-16a	14-Aug-11	Y				PEC		19
TPN	TPN-16a	14-Aug-11	Y				PEC		17
TPN	TPN-16a	14-Aug-11	N	M	IM	UD	PEC, OTO	25	21
TPN	TPN-16b	15-Aug-11	N (± 1)						
TPN	TPN-16b	15-Aug-11	N	UN	IM	UD	PEC, OTO	8	3
TPN	TPN-16b	15-Aug-11	N (± 10)				PEC		6
TPN	TPN-16b	15-Aug-11	Y				PEC		18
TPN	TPN-16b	15-Aug-11	Y	F	MA	RS	PEC, OTO	18	14
TPN	TPN-16b	15-Aug-11	Y				PEC		16
TPN	TPN-16b	15-Aug-11	Y				PEC		15
TPN	TPN-16b	15-Aug-11	N	F	MA	RS	PEC, OTO	22	14
TPN	TPN-16b	15-Aug-11	N	F	MA	RS	PEC, OTO	20	11
TPN	TPN-16b	15-Aug-11	N	F	MA	RS	PEC, OTO	15	11
TPN	TPN-16b	15-Aug-11	Y				PEC		18
TPN	TPN-16b	15-Aug-11	N	M	MA	RS	PEC, OTO	21	19
TPN	TPN-16b	15-Aug-11	Y				PEC		26
TPN	TPN-16b	15-Aug-11	Y				PEC		14
TPN	TPN-16b	15-Aug-11	N	UN	IM	UD	PEC, OTO	19	17
TPN	TPN-16b	15-Aug-11	N	M	MA	RI	PEC, OTO	15	15
TPN	TPN-16b	15-Aug-11	Y				PEC		18
TPN	TPN-16b	15-Aug-11	Y				PEC		15
TPN	TPN-16b	15-Aug-11	N	F	MA	RS	PEC, OTO	28	26
TPN	TPN-16b	15-Aug-11	N	F	MA	RS	PEC, OTO	17	17
TPN	TPN-16b	15-Aug-11	N	F	MA	RS	PEC, OTO	34	25
TPN	TPN-16b	15-Aug-11	N	F	MA	RI	PEC, OTO	37	ua
TPN	TPN-16b	15-Aug-11	N	M	MA	RS	PEC, OTO	34	20
TPN	TPN-17	14-Aug-11	N	F	MA	RS	PEC, OTO	16	12
TPN	TPN-20	15-Aug-11	N (± 10)				PEC		6
TPN	TPN-22	15-Aug-11	Y				PEC		18
TPN	TPN-22	15-Aug-11	N	F	MA	RS	PEC, OTO	20	15
TPN	TPN-22	15-Aug-11	N	F	MA	RI	PEC, OTO	18	13
TPN	TPN-22	15-Aug-11	Y				PEC		16
TPN	TPN-22	15-Aug-11	Y				PEC		25
TPN	TPN-24b	16-Aug-11	Y				PEC		6
TPN	TPN-24b	16-Aug-11	Y				PEC		10
TPN	TPN-24b	16-Aug-11	N	M	IM	UD	PEC, OTO	9	5
TPN	TPN-24b	16-Aug-11	N	M	MA	RI	PEC, OTO	29	22
TPN	TPN-25	16-Aug-11	N	F	IM	UD	PEC, OTO	13	9
TPN	TPN-25	16-Aug-11	N	F	IM	UD	PEC, OTO	14	7
TPN	TPN-25	16-Aug-11	Y				PEC		22
TPN	TPN-25	16-Aug-11	Y				PEC		18
TPN	TPN-25	16-Aug-11	Y				PEC		19
TPN	TPN-25	16-Aug-11	Y				PEC		21
TPN	TPN-10	14-Aug-11	N	M	MA	RI	PEC, OTO	14	14
TPN	TPN-11a	14-Aug-11	N	F	MA	RI	PEC, OTO	14	14

Lake	Set ID	Date	Ext. DELT	Ext. Parasites	Tagged?	Tag ID	RECAP?	Gonad weight (g)
TPN	TPN-03	11-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06b	13-Aug-11	NONE	NONE	Y	87465	N	
TPN	TPN-10	14-Aug-11	NONE	NONE	N		N	
TPN	TPN-10	14-Aug-11	NONE	NONE	N		N	
TPN	TPN-10	14-Aug-11	NONE	NONE	N		N	
TPN	TPN-10	14-Aug-11	NONE	NONE	N		N	
TPN	TPN-10	14-Aug-11	NONE	NONE	N		N	
TPN	TPN-10	14-Aug-11	NONE	NONE	N		N	
TPN	TPN-10	14-Aug-11	missing part of left operculum	none	N		N	
TPN	TPN-10	14-Aug-11	NONE	NONE	N		N	
TPN	TPN-10	14-Aug-11	NONE	NONE	N		N	
TPN	TPN-11a	14-Aug-11	NONE	NONE	N		N	
TPN	TPN-16a	14-Aug-11	NONE	NONE	N		N	
TPN	TPN-16b	15-Aug-11	redness (spots) around anus and peduncle	NONE			N	
TPN	TPN-23	15-Aug-11	NONE	NONE	N		N	
TPN	TPN-24b	16-Aug-11	NONE	NONE	N		N	
TPN	TPN-25	16-Aug-11	NONE	NONE	N		N	
TPN	TPN-25	16-Aug-11	NONE	NONE	N		N	
TPN	TPN-25	16-Aug-11	NONE	NONE	N		N	
TPN	TPN-25	16-Aug-11	NONE	NONE	N		N	
TPN	TPN-25	16-Aug-11	NONE	NONE	N		N	
TPN	TPN-01	11-Aug-11	NONE	NONE	N		N	
TPN	TPN-01	11-Aug-11	NONE	NONE	N		N	
TPN	TPN-01	11-Aug-11	NONE	NONE	N		N	
TPN	TPN-02	11-Aug-11	NONE	NONE	N		N	
TPN	TPN-02	11-Aug-11	NONE	NONE	N		N	
TPN	TPN-03	11-Aug-11	NONE	NONE	N		N	
TPN	TPN-03	11-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	Y	87451	N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	Y	87452	N	
TPN	TPN-05a	13-Aug-11	Lower lobe of caudal missing	none	Y	87453	N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05a	13-Aug-11	NONE	NONE	Y	87454	N	
TPN	TPN-05b	13-Aug-11	NONE	NONE	Y	87464	N	
TPN	TPN-05b	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-05c	13-Aug-11			Y	87467	N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N	87455	N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	Y	87456	N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	Y	87457	N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	Y	87458	N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	Y	87459	N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	Y	87460	N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	Y	87461	N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	Y	87462	N	
TPN	TPN-06a	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06b	13-Aug-11	NONE	NONE	Y	87466	N	
TPN	TPN-06c	13-Aug-11	NONE	NONE	N		N	
TPN	TPN-06c	13-Aug-11	NONE	NONE	Y	87468	N	
TPN	TPN-06c	13-Aug-11	NONE	NONE	Y	87469	N	

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Lake	Set ID	Date	Gonad appearance	Weight of 400 eggs (g)	Liver weight (g)	Liver appearance
TPN	TPN-03	11-Aug-11				
TPN	TPN-05a	13-Aug-11	Normal			NORMAL
TPN	TPN-05a	13-Aug-11	Normal			NORMAL
TPN	TPN-05a	13-Aug-11	Normal			NORMAL
TPN	TPN-06a	13-Aug-11				
TPN	TPN-06a	13-Aug-11				
TPN	TPN-06a	13-Aug-11	normal			NORMAL
TPN	TPN-06a	13-Aug-11	Normal			NORMAL
TPN	TPN-06a	13-Aug-11	Normal			NORMAL
TPN	TPN-06a	13-Aug-11	Normal			NORMAL
TPN	TPN-06a	13-Aug-11	Normal			NORMAL
TPN	TPN-06a	13-Aug-11				
TPN	TPN-06a	13-Aug-11				
TPN	TPN-06b	13-Aug-11				
TPN	TPN-10	14-Aug-11				
TPN	TPN-10	14-Aug-11	Normal			NORMAL
TPN	TPN-10	14-Aug-11	Normal			NORMAL
TPN	TPN-10	14-Aug-11				
TPN	TPN-10	14-Aug-11				
TPN	TPN-10	14-Aug-11	Normal			NORMAL
TPN	TPN-10	14-Aug-11	Normal			NORMAL
TPN	TPN-10	14-Aug-11	Normal			NORMAL
TPN	TPN-10	14-Aug-11				
TPN	TPN-11a	14-Aug-11				
TPN	TPN-16a	14-Aug-11				
TPN	TPN-16b	15-Aug-11				
TPN	TPN-23	15-Aug-11	normal			ONE PARASITIC CYST
TPN	TPN-24b	16-Aug-11				
TPN	TPN-25	16-Aug-11	normal			NORMAL
TPN	TPN-25	16-Aug-11				
TPN	TPN-25	16-Aug-11	normal			ONE PARASITIC CYST/TUMOR
TPN	TPN-25	16-Aug-11				
TPN	TPN-25	16-Aug-11				
TPN	TPN-01	11-Aug-11				
TPN	TPN-01	11-Aug-11				
TPN	TPN-01	11-Aug-11				
TPN	TPN-02	11-Aug-11				
TPN	TPN-02	11-Aug-11				
TPN	TPN-03	11-Aug-11				
TPN	TPN-03	11-Aug-11				
TPN	TPN-05a	13-Aug-11	Normal			NORMAL
TPN	TPN-05a	13-Aug-11	Normal			NORMAL
TPN	TPN-05a	13-Aug-11	Normal			NORMAL
TPN	TPN-05a	13-Aug-11				
TPN	TPN-05a	13-Aug-11				
TPN	TPN-05a	13-Aug-11				
TPN	TPN-05a	13-Aug-11				
TPN	TPN-05a	13-Aug-11	Normal			NORMAL
TPN	TPN-05a	13-Aug-11	Normal			NORMAL
TPN	TPN-05a	13-Aug-11				
TPN	TPN-05a	13-Aug-11	Normal			NORMAL
TPN	TPN-05a	13-Aug-11				
TPN	TPN-05a	13-Aug-11				
TPN	TPN-05a	13-Aug-11				
TPN	TPN-05b	13-Aug-11				
TPN	TPN-05b	13-Aug-11	Normal, very large eggs			NORMAL
TPN	TPN-05c	13-Aug-11				
TPN	TPN-06a	13-Aug-11				
TPN	TPN-06a	13-Aug-11	Normal			slightly, lightly coloured
TPN	TPN-06a	13-Aug-11	Normal			NORMAL
TPN	TPN-06a	13-Aug-11	Normal			NORMAL
TPN	TPN-06a	13-Aug-11				
TPN	TPN-06a	13-Aug-11				
TPN	TPN-06a	13-Aug-11				
TPN	TPN-06a	13-Aug-11				
TPN	TPN-06a	13-Aug-11	Normal			NORMAL
TPN	TPN-06a	13-Aug-11	Normal			NORMAL
TPN	TPN-06a	13-Aug-11				
TPN	TPN-06a	13-Aug-11	Normal			NORMAL
TPN	TPN-06a	13-Aug-11	Normal			ONE PARASITIC CYST
TPN	TPN-06a	13-Aug-11				
TPN	TPN-06a	13-Aug-11	Normal			ONE PARASITIC CYST OR DISCOLORATION
TPN	TPN-06b	13-Aug-11				
TPN	TPN-06c	13-Aug-11				
TPN	TPN-06c	13-Aug-11				
TPN	TPN-06c	13-Aug-11				

Lake	Set ID	Date	Int. DELT	Int. Parasites
TPN	TPN-03	11-Aug-11		
TPN	TPN-05a	13-Aug-11	NONE	MODERATE
TPN	TPN-05a	13-Aug-11	NONE	HIGH
TPN	TPN-05a	13-Aug-11	NONE	NONE
TPN	TPN-06a	13-Aug-11		
TPN	TPN-06a	13-Aug-11		
TPN	TPN-06a	13-Aug-11	NONE	MODERATE
TPN	TPN-06a	13-Aug-11	NONE	MODERATE
TPN	TPN-06a	13-Aug-11	NONE	NONE
TPN	TPN-06a	13-Aug-11	NONE	LOW
TPN	TPN-06a	13-Aug-11	NONE	LOW
TPN	TPN-06a	13-Aug-11		
TPN	TPN-06a	13-Aug-11		
TPN	TPN-06b	13-Aug-11		
TPN	TPN-10	14-Aug-11		
TPN	TPN-10	14-Aug-11	NONE	LOW (1 cyst)
TPN	TPN-10	14-Aug-11	NONE	NONE
TPN	TPN-10	14-Aug-11		
TPN	TPN-10	14-Aug-11		
TPN	TPN-10	14-Aug-11	NONE	LOW
TPN	TPN-10	14-Aug-11	NONE	NONE
TPN	TPN-10	14-Aug-11	NONE	LOW
TPN	TPN-10	14-Aug-11		
TPN	TPN-11a	14-Aug-11		
TPN	TPN-16a	14-Aug-11		
TPN	TPN-16b	15-Aug-11		
TPN	TPN-23	15-Aug-11	NONE	LOW (parasitic cyst)
TPN	TPN-24b	16-Aug-11		
TPN	TPN-25	16-Aug-11	NONE	MODERATE
TPN	TPN-25	16-Aug-11		
TPN	TPN-25	16-Aug-11	NONE	NONE
TPN	TPN-25	16-Aug-11		
TPN	TPN-25	16-Aug-11		
TPN	TPN-01	11-Aug-11		
TPN	TPN-01	11-Aug-11		
TPN	TPN-01	11-Aug-11		
TPN	TPN-02	11-Aug-11		
TPN	TPN-02	11-Aug-11		
TPN	TPN-03	11-Aug-11		
TPN	TPN-03	11-Aug-11		
TPN	TPN-05a	13-Aug-11	NONE	MODERATE
TPN	TPN-05a	13-Aug-11	NONE	LOW
TPN	TPN-05a	13-Aug-11	NONE	MODERATE
TPN	TPN-05a	13-Aug-11		
TPN	TPN-05a	13-Aug-11		
TPN	TPN-05a	13-Aug-11		
TPN	TPN-05a	13-Aug-11		
TPN	TPN-05a	13-Aug-11	NONE	MODERATE
TPN	TPN-05a	13-Aug-11	NONE	MODERATE
TPN	TPN-05a	13-Aug-11		
TPN	TPN-05a	13-Aug-11	NONE	LOW
TPN	TPN-05a	13-Aug-11		
TPN	TPN-05a	13-Aug-11		
TPN	TPN-05a	13-Aug-11		
TPN	TPN-05b	13-Aug-11		
TPN	TPN-05b	13-Aug-11	NONE	LOW
TPN	TPN-05c	13-Aug-11		
TPN	TPN-06a	13-Aug-11		
TPN	TPN-06a	13-Aug-11	NONE	LOW
TPN	TPN-06a	13-Aug-11	NONE	MODERATE
TPN	TPN-06a	13-Aug-11	NONE	LOW
TPN	TPN-06a	13-Aug-11		
TPN	TPN-06a	13-Aug-11		
TPN	TPN-06a	13-Aug-11		
TPN	TPN-06a	13-Aug-11		
TPN	TPN-06a	13-Aug-11		
TPN	TPN-06a	13-Aug-11	NONE	MODERATE
TPN	TPN-06a	13-Aug-11	Pinched swim bladder	NONE
TPN	TPN-06a	13-Aug-11		
TPN	TPN-06a	13-Aug-11	NONE	MODERATE
TPN	TPN-06a	13-Aug-11	one parasite cyst	LOW
TPN	TPN-06a	13-Aug-11		
TPN	TPN-06a	13-Aug-11	one parasite or discolouration to liver	moderate gut parasites; tapeworm
TPN	TPN-06b	13-Aug-11		
TPN	TPN-06c	13-Aug-11		
TPN	TPN-06c	13-Aug-11		
TPN	TPN-06c	13-Aug-11		

Lake	Set ID	Date	Int. DELT	Int. Parasites
TPN	TPN-07a	13-Aug-11	NONE	LOW
TPN	TPN-08a	13-Aug-11	NONE	NONE
TPN	TPN-08a	13-Aug-11	NONE	NONE
TPN	TPN-08b	13-Aug-11		
TPN	TPN-08b	13-Aug-11		
TPN	TPN-08b	13-Aug-11	NONE	NONE
TPN	TPN-08b	13-Aug-11	NONE	NONE
TPN	TPN-08b	13-Aug-11		
TPN	TPN-08b	13-Aug-11		
TPN	TPN-10	14-Aug-11		
TPN	TPN-10	14-Aug-11		
TPN	TPN-10	14-Aug-11	NONE	LOW
TPN	TPN-10	14-Aug-11	NONE	LOW
TPN	TPN-10	14-Aug-11		
TPN	TPN-10	14-Aug-11	NONE	LOW
TPN	TPN-10	14-Aug-11	NONE	LOW
TPN	TPN-10	14-Aug-11		
TPN	TPN-10	14-Aug-11	NONE	LOW
TPN	TPN-10	14-Aug-11	NONE	LOW
TPN	TPN-10	14-Aug-11	NONE	NONE
TPN	TPN-10	14-Aug-11	NONE	LOW
TPN	TPN-10	14-Aug-11	NONE	LOW
TPN	TPN-10	14-Aug-11		
TPN	TPN-10	14-Aug-11	NONE	LOW
TPN	TPN-10	14-Aug-11	NONE	LOW
TPN	TPN-11a	14-Aug-11	NONE	MODERATE
TPN	TPN-11a	14-Aug-11	NONE	LOW
TPN	TPN-11a	14-Aug-11	NONE	LOW
TPN	TPN-11a	14-Aug-11		
TPN	TPN-11a	14-Aug-11	NONE	LOW
TPN	TPN-11a	14-Aug-11	NONE	NONE
TPN	TPN-11a	14-Aug-11	NONE	NONE
TPN	TPN-11c	14-Aug-11	NONE	LOW
TPN	TPN-11c	14-Aug-11		
TPN	TPN-11c	14-Aug-11		
TPN	TPN-14	14-Aug-11		
TPN	TPN-16a	14-Aug-11		
TPN	TPN-16a	14-Aug-11		
TPN	TPN-16a	14-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11		
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11		
TPN	TPN-16b	15-Aug-11		
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11		
TPN	TPN-16b	15-Aug-11		
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11		
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11	NONE	MODERATE
TPN	TPN-16b	15-Aug-11	NONE	NONE
TPN	TPN-16b	15-Aug-11		
TPN	TPN-16b	15-Aug-11		
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11	NONE	LOW
TPN	TPN-16b	15-Aug-11	NONE	MODERATE
TPN	TPN-17	14-Aug-11	NONE	LOW
TPN	TPN-20	15-Aug-11		
TPN	TPN-22	15-Aug-11		
TPN	TPN-22	15-Aug-11	NONE	NONE
TPN	TPN-22	15-Aug-11	NONE	LOW (1 CYST)
TPN	TPN-22	15-Aug-11		
TPN	TPN-22	15-Aug-11		
TPN	TPN-24b	16-Aug-11		
TPN	TPN-24b	16-Aug-11		
TPN	TPN-24b	16-Aug-11	NONE	LOW
TPN	TPN-24b	16-Aug-11	NONE	LOW
TPN	TPN-25	16-Aug-11	NONE	LOW
TPN	TPN-25	16-Aug-11	NONE	LOW
TPN	TPN-25	16-Aug-11		
TPN	TPN-25	16-Aug-11		
TPN	TPN-25	16-Aug-11		
TPN	TPN-25	16-Aug-11		
TPN	TPN-10	14-Aug-11	NONE	NONE
TPN	TPN-11a	14-Aug-11	NONE	NONE

Lake	Set ID	Date	Stomach contents
TPN	TPN-03	11-Aug-11	
TPN	TPN-05a	13-Aug-11	MT stomach full intestine
TPN	TPN-05a	13-Aug-11	MT stomach full intestine - Chir
TPN	TPN-05a	13-Aug-11	60% full of zoops
TPN	TPN-06a	13-Aug-11	
TPN	TPN-06a	13-Aug-11	
TPN	TPN-06a	13-Aug-11	20% full
TPN	TPN-06a	13-Aug-11	MT
TPN	TPN-06a	13-Aug-11	MT
TPN	TPN-06a	13-Aug-11	10% ZOOPLANKTON
TPN	TPN-06a	13-Aug-11	10% ZOOPLANKTON
TPN	TPN-06a	13-Aug-11	
TPN	TPN-06a	13-Aug-11	
TPN	TPN-06b	13-Aug-11	
TPN	TPN-10	14-Aug-11	
TPN	TPN-10	14-Aug-11	50% - EPHEMEROPTERA
TPN	TPN-10	14-Aug-11	5% - CHIRONOMIDS
TPN	TPN-10	14-Aug-11	
TPN	TPN-10	14-Aug-11	
TPN	TPN-10	14-Aug-11	80% ZOOPLANKTON
TPN	TPN-10	14-Aug-11	80% ZOOPLANKTON
TPN	TPN-10	14-Aug-11	50% ZOOPLANKTON
TPN	TPN-10	14-Aug-11	
TPN	TPN-11a	14-Aug-11	
TPN	TPN-16a	14-Aug-11	
TPN	TPN-16b	15-Aug-11	
TPN	TPN-23	15-Aug-11	80% ZOOPLANKTON
TPN	TPN-24b	16-Aug-11	
TPN	TPN-25	16-Aug-11	MT
TPN	TPN-25	16-Aug-11	
TPN	TPN-25	16-Aug-11	10% ZOOPLANKTON
TPN	TPN-25	16-Aug-11	
TPN	TPN-25	16-Aug-11	
TPN	TPN-01	11-Aug-11	
TPN	TPN-01	11-Aug-11	
TPN	TPN-01	11-Aug-11	
TPN	TPN-02	11-Aug-11	
TPN	TPN-02	11-Aug-11	
TPN	TPN-03	11-Aug-11	
TPN	TPN-03	11-Aug-11	
TPN	TPN-05a	13-Aug-11	MT
TPN	TPN-05a	13-Aug-11	10%
TPN	TPN-05a	13-Aug-11	10%
TPN	TPN-05a	13-Aug-11	
TPN	TPN-05a	13-Aug-11	
TPN	TPN-05a	13-Aug-11	
TPN	TPN-05a	13-Aug-11	
TPN	TPN-05a	13-Aug-11	MT stomach full intestine
TPN	TPN-05a	13-Aug-11	MT, full intestine
TPN	TPN-05a	13-Aug-11	
TPN	TPN-05a	13-Aug-11	50% stomach, full intestine
TPN	TPN-05a	13-Aug-11	
TPN	TPN-05a	13-Aug-11	
TPN	TPN-05a	13-Aug-11	
TPN	TPN-05a	13-Aug-11	
TPN	TPN-05b	13-Aug-11	
TPN	TPN-05b	13-Aug-11	20% - fish remains
TPN	TPN-05c	13-Aug-11	
TPN	TPN-06a	13-Aug-11	
TPN	TPN-06a	13-Aug-11	40% -LKTR remains, Chir
TPN	TPN-06a	13-Aug-11	50% full
TPN	TPN-06a	13-Aug-11	50% full
TPN	TPN-06a	13-Aug-11	
TPN	TPN-06a	13-Aug-11	
TPN	TPN-06a	13-Aug-11	
TPN	TPN-06a	13-Aug-11	
TPN	TPN-06a	13-Aug-11	10% full
TPN	TPN-06a	13-Aug-11	10% - zoops
TPN	TPN-06a	13-Aug-11	
TPN	TPN-06a	13-Aug-11	10% ZOOPLANKTON
TPN	TPN-06a	13-Aug-11	20% FISH REMAINS (SCULPIN)
TPN	TPN-06a	13-Aug-11	
TPN	TPN-06a	13-Aug-11	10% ZOOPLANKTON
TPN	TPN-06b	13-Aug-11	
TPN	TPN-06c	13-Aug-11	
TPN	TPN-06c	13-Aug-11	
TPN	TPN-06c	13-Aug-11	

Lake	Set ID	Date	Stomach contents
TPN	TPN-07a	13-Aug-11	10% - EPHEMEROPTERA, FISH REMAINS, CHIRONOMIDS
TPN	TPN-08a	13-Aug-11	5% LARVAL FISH, PUPA UNID
TPN	TPN-08a	13-Aug-11	60% - CHIRONOMIDS, TRICHOPTERANS
TPN	TPN-08b	13-Aug-11	
TPN	TPN-08b	13-Aug-11	
TPN	TPN-08b	13-Aug-11	20% CHIRONOMIDS, FISH REMAINS (SCULPIN)
TPN	TPN-08b	13-Aug-11	5% - CHIRONOMIDS
TPN	TPN-08b	13-Aug-11	
TPN	TPN-10	14-Aug-11	
TPN	TPN-10	14-Aug-11	
TPN	TPN-10	14-Aug-11	10% ZOOPLANKTON
TPN	TPN-10	14-Aug-11	MT
TPN	TPN-10	14-Aug-11	
TPN	TPN-10	14-Aug-11	10% CHIRONOMIDS
TPN	TPN-10	14-Aug-11	90% FISH REMAINS (LKTR)
TPN	TPN-10	14-Aug-11	
TPN	TPN-10	14-Aug-11	40% ZOOPLANKTON
TPN	TPN-10	14-Aug-11	MT
TPN	TPN-10	14-Aug-11	30% CHIRONOMIDS, FISH REMAINS UNID
TPN	TPN-10	14-Aug-11	50% TRICHOPTERA, FISH REMAINS (SCULPIN)
TPN	TPN-10	14-Aug-11	10% CHIRONOMIDS
TPN	TPN-10	14-Aug-11	
TPN	TPN-10	14-Aug-11	5% CHIRONOMIDS
TPN	TPN-10	14-Aug-11	80% CHIRONOMIDS, FISH REMAINS UNID
TPN	TPN-11a	14-Aug-11	10% FISH REMAINS UNID
TPN	TPN-11a	14-Aug-11	25% FISH REMAINS UNID
TPN	TPN-11a	14-Aug-11	< 10% EPHEMEROPTERA, CHIRONOMIDS
TPN	TPN-11a	14-Aug-11	
TPN	TPN-11a	14-Aug-11	60% FISH REMAINS, CHIRONOMIDS
TPN	TPN-11a	14-Aug-11	20% FISH REMAINS (SCULPIN)
TPN	TPN-11a	14-Aug-11	90% CHIRONOMIDS, TRICHOPTERANS, FISH REMAINS UNID
TPN	TPN-11c	14-Aug-11	10% FISH REMAINS UNID
TPN	TPN-11c	14-Aug-11	
TPN	TPN-11c	14-Aug-11	
TPN	TPN-14	14-Aug-11	
TPN	TPN-16a	14-Aug-11	
TPN	TPN-16a	14-Aug-11	
TPN	TPN-16a	14-Aug-11	MT
TPN	TPN-16b	15-Aug-11	
TPN	TPN-16b	15-Aug-11	20% FISH REMAINS UNID
TPN	TPN-16b	15-Aug-11	
TPN	TPN-16b	15-Aug-11	
TPN	TPN-16b	15-Aug-11	MT
TPN	TPN-16b	15-Aug-11	
TPN	TPN-16b	15-Aug-11	
TPN	TPN-16b	15-Aug-11	20% CHIRONOMIDS, UNID INVERTS
TPN	TPN-16b	15-Aug-11	MT
TPN	TPN-16b	15-Aug-11	30% CHIRONOMIDS, FISH REMAINS UNID
TPN	TPN-16b	15-Aug-11	
TPN	TPN-16b	15-Aug-11	10% DIPTERA
TPN	TPN-16b	15-Aug-11	
TPN	TPN-16b	15-Aug-11	
TPN	TPN-16b	15-Aug-11	100% FISH REMAINS (LKTR)
TPN	TPN-16b	15-Aug-11	5% INVERTS UNID
TPN	TPN-16b	15-Aug-11	
TPN	TPN-16b	15-Aug-11	
TPN	TPN-16b	15-Aug-11	MT
TPN	TPN-16b	15-Aug-11	20% FISH REMAINS UNID, INVERT REMAINS UNID
TPN	TPN-16b	15-Aug-11	MT
TPN	TPN-16b	15-Aug-11	MT
TPN	TPN-16b	15-Aug-11	50% CHIRONOMIDS, DIPTERANS
TPN	TPN-17	14-Aug-11	5% FISH REMAINS UNID
TPN	TPN-20	15-Aug-11	
TPN	TPN-22	15-Aug-11	
TPN	TPN-22	15-Aug-11	20% FISH REMAINS (UNID)
TPN	TPN-22	15-Aug-11	20% CHIRONOMIDS
TPN	TPN-22	15-Aug-11	
TPN	TPN-22	15-Aug-11	
TPN	TPN-24b	16-Aug-11	
TPN	TPN-24b	16-Aug-11	
TPN	TPN-24b	16-Aug-11	10% TRICHOPTERAN PUPAE
TPN	TPN-24b	16-Aug-11	MT
TPN	TPN-25	16-Aug-11	50% CHIRONOMIDS, FISH REMAINS UNID
TPN	TPN-25	16-Aug-11	20% TRICHOPTERANS
TPN	TPN-25	16-Aug-11	
TPN	TPN-25	16-Aug-11	
TPN	TPN-25	16-Aug-11	
TPN	TPN-10	14-Aug-11	10% EPHEMEROPTERA, CHIRONOMIDS
TPN	TPN-11a	14-Aug-11	70% CHIRONOMIDS, EPHEMEROPTERA, OSTRACODA

Lake	Set ID	Date	Mesh Size (mm)	Day/Night	Species	Sample ID	Mortality	Length.mm	Weight.g
INUG	INUG-02	17-Aug-11	51	NIGHT	ARCH	INUG-02.13	Y	270	194
INUG	INUG-07	18-Aug-11	76	NIGHT	ARCH	INUG-07.02	N	868	7650
INUG	INUG-07	18-Aug-11	25	NIGHT	ARCH	INUG-07.15	Y	150	40
INUG	INUG-08	18-Aug-11	51	NIGHT	ARCH	INUG-08.10	N	297	170
INUG	INUG-14	19-Aug-11	51	NIGHT	ARCH	INUG-14.07	N	253	150
INUG	INUG-01	17-Aug-11	51	DAY	LKTR	INUG-01.01	N	558	1450
INUG	INUG-01	17-Aug-11	51	DAY	LKTR	INUG-01.02	N	750	4550
INUG	INUG-01	17-Aug-11	51	DAY	LKTR	INUG-01.03	N	789	5600
INUG	INUG-01	17-Aug-11	51	DAY	LKTR	INUG-01.04	N	792	6750
INUG	INUG-01	17-Aug-11	51	DAY	LKTR	INUG-01.05	N	505	1300
INUG	INUG-01	17-Aug-11	76	DAY	LKTR	INUG-01.06	N	630	2575
INUG	INUG-01	17-Aug-11	76	DAY	LKTR	INUG-01.07	N	548	1600
INUG	INUG-01	17-Aug-11	76	DAY	LKTR	INUG-01.08	Y	549	1556
INUG	INUG-01	17-Aug-11	51	DAY	LKTR	INUG-01.09	Y	278	208
INUG	INUG-01	17-Aug-11	51	DAY	LKTR	INUG-01.10	Y	267	204
INUG	INUG-01	17-Aug-11	76	DAY	LKTR	INUG-01.16	Y	506	1142
INUG	INUG-01	17-Aug-11	76	DAY	LKTR	INUG-01.17	Y	467	1150
INUG	INUG-02	17-Aug-11	25	NIGHT	LKTR	INUG-02.01	N	310	290
INUG	INUG-02	17-Aug-11	38	NIGHT	LKTR	INUG-02.05	Y	217	100
INUG	INUG-02	17-Aug-11	38	NIGHT	LKTR	INUG-02.06	Y	225	114
INUG	INUG-02	17-Aug-11	38	NIGHT	LKTR	INUG-02.07	N	626	2400
INUG	INUG-02	17-Aug-11	51	NIGHT	LKTR	INUG-02.09	N	514	1500
INUG	INUG-02	17-Aug-11	51	NIGHT	LKTR	INUG-02.10	N	544	1508
INUG	INUG-02	17-Aug-11	51	NIGHT	LKTR	INUG-02.11	N	555	1550
INUG	INUG-02	17-Aug-11	51	NIGHT	LKTR	INUG-02.12	N	598	2150
INUG	INUG-02	17-Aug-11	76	NIGHT	LKTR	INUG-02.15	Y	406	654
INUG	INUG-02	17-Aug-11	102	NIGHT	LKTR	INUG-02.16	Y	705	3712
INUG	INUG-02	17-Aug-11	102	NIGHT	LKTR	INUG-02.17	Y	560	1650
INUG	INUG-02	17-Aug-11	102	NIGHT	LKTR	INUG-02.18	Y	496	1192
INUG	INUG-02	17-Aug-11	126	NIGHT	LKTR	INUG-02.19	N	903	6350
INUG	INUG-02	17-Aug-11	126	NIGHT	LKTR	INUG-02.20	N	517	1500
INUG	INUG-02	17-Aug-11	25	NIGHT	LKTR	INUG-02.21	Y	152	36
INUG	INUG-04	17-Aug-11	51	DAY	LKTR	INUG-04.01	N	322	425
INUG	INUG-04	17-Aug-11	51	DAY	LKTR	INUG-04.02	N	688	3255
INUG	INUG-04	17-Aug-11	51	DAY	LKTR	INUG-04.03	Y	365	458
INUG	INUG-05	17-Aug-11	51	DAY	LKTR	INUG-05.03	N	885	10300
INUG	INUG-05	17-Aug-11	76	DAY	LKTR	INUG-05.04	N	394	575
INUG	INUG-05	17-Aug-11	76	DAY	LKTR	INUG-05.05	Y	522	1420
INUG	INUG-05	17-Aug-11	76	DAY	LKTR	INUG-05.06	Y	524	1638
INUG	INUG-05	17-Aug-11	76	DAY	LKTR	INUG-05.07	Y	665	3302
INUG	INUG-05	17-Aug-11	76	DAY	LKTR	INUG-05.08	Y	508	1360
INUG	INUG-06	17-Aug-11	76	DAY	LKTR	INUG-06.01	N	300	270
INUG	INUG-07	18-Aug-11	102	NIGHT	LKTR	INUG-07.01	Y	504	1214
INUG	INUG-07	18-Aug-11	76	NIGHT	LKTR	INUG-07.03	Y	519	1104
INUG	INUG-07	18-Aug-11	51	NIGHT	LKTR	INUG-07.04	N	558	1750
INUG	INUG-07	18-Aug-11	51	NIGHT	LKTR	INUG-07.05	N	740	4600
INUG	INUG-07	18-Aug-11	51	NIGHT	LKTR	INUG-07.06	Y	261	168
INUG	INUG-07	18-Aug-11	51	NIGHT	LKTR	INUG-07.07	N	518	1400
INUG	INUG-07	18-Aug-11	51	NIGHT	LKTR	INUG-07.08	N	243	170
INUG	INUG-07	18-Aug-11	38	NIGHT	LKTR	INUG-07.09	Y	206	84
INUG	INUG-07	18-Aug-11	38	NIGHT	LKTR	INUG-07.10	Y	219	94
INUG	INUG-07	18-Aug-11	38	NIGHT	LKTR	INUG-07.11	N	554	1775
INUG	INUG-07	18-Aug-11	38	NIGHT	LKTR	INUG-07.12	Y	223	112
INUG	INUG-07	18-Aug-11	38	NIGHT	LKTR	INUG-07.13	N	232	170
INUG	INUG-08	18-Aug-11	51	NIGHT	LKTR	INUG-08.01	N	514	1450
INUG	INUG-08	18-Aug-11	51	NIGHT	LKTR	INUG-08.02	N	584	1950
INUG	INUG-08	18-Aug-11	51	NIGHT	LKTR	INUG-08.03	N	564	1650
INUG	INUG-08	18-Aug-11	51	NIGHT	LKTR	INUG-08.06	Y	258	172
INUG	INUG-08	18-Aug-11	51	NIGHT	LKTR	INUG-08.07	N	518	1500
INUG	INUG-08	18-Aug-11	51	NIGHT	LKTR	INUG-08.15	Y	231	132
INUG	INUG-08	18-Aug-11	76	NIGHT	LKTR	INUG-08.17	Y	515	1293
INUG	INUG-08	18-Aug-11	76	NIGHT	LKTR	INUG-08.18	Y	598	2046
INUG	INUG-08	18-Aug-11	76	NIGHT	LKTR	INUG-08.19	Y	525	1501
INUG	INUG-08	18-Aug-11	76	NIGHT	LKTR	INUG-08.20	Y	543	1613
INUG	INUG-09	18-Aug-11	51	DAY	LKTR	INUG-09.01	N	332	450
INUG	INUG-09	18-Aug-11	76	DAY	LKTR	INUG-09.02	N	875	5900
INUG	INUG-09	18-Aug-11	76	DAY	LKTR	INUG-09.03	N	616	2425
INUG	INUG-09	18-Aug-11	102	DAY	LKTR	INUG-09.05	N	499	1550
INUG	INUG-10	18-Aug-11	51	DAY	LKTR	INUG-10.01	N	705	3900

Lake	Set ID	Date	Mesh Size (mm)	Day/Night	Species	Sample ID	Mortality	Length.mm	Weight.g
INUG	INUG-10	18-Aug-11	51	DAY	LKTR	INUG-10.02	Y	524	1700
INUG	INUG-10	18-Aug-11	76	DAY	LKTR	INUG-10.04	Y	562	1602
INUG	INUG-13	19-Aug-11	76	NIGHT	LKTR	INUG-13.01	N	538	1500
INUG	INUG-13	19-Aug-11	76	NIGHT	LKTR	INUG-13.03	N	494	1400
INUG	INUG-13	19-Aug-11	76	NIGHT	LKTR	INUG-13.05	Y	741	4032
INUG	INUG-13	19-Aug-11	76	NIGHT	LKTR	INUG-13.07	Y	515	1386
INUG	INUG-13	19-Aug-11	51	NIGHT	LKTR	INUG-13.08	N	253	160
INUG	INUG-13	19-Aug-11	51	NIGHT	LKTR	INUG-13.10	Y	356	460
INUG	INUG-13	19-Aug-11	51	NIGHT	LKTR	INUG-13.11	N	542	1550
INUG	INUG-13	19-Aug-11	51	NIGHT	LKTR	INUG-13.13	N	205	100
INUG	INUG-13	19-Aug-11	38	NIGHT	LKTR	INUG-13.15	N	175	
INUG	INUG-13	19-Aug-11	38	NIGHT	LKTR	INUG-13.16	N	217	110
INUG	INUG-13	19-Aug-11	76	NIGHT	LKTR	INUG-13.17	Y	449	1008
INUG	INUG-14	19-Aug-11	51	NIGHT	LKTR	INUG-14.01	Y	283	238
INUG	INUG-14	19-Aug-11	51	NIGHT	LKTR	INUG-14.03	Y	357	490
INUG	INUG-14	19-Aug-11	51	NIGHT	LKTR	INUG-14.05	Y	245	144
INUG	INUG-14	19-Aug-11	51	NIGHT	LKTR	INUG-14.06	N	375	625
INUG	INUG-14	19-Aug-11	76	NIGHT	LKTR	INUG-14.12	N	577	1700
INUG	INUG-14	19-Aug-11	76	NIGHT	LKTR	INUG-14.13	Y	625	2884
INUG	INUG-14	19-Aug-11	76	NIGHT	LKTR	INUG-14.15	Y	548	1788
INUG	INUG-14	19-Aug-11	76	NIGHT	LKTR	INUG-14.16	Y	487	1142
INUG	INUG-14	19-Aug-11	76	NIGHT	LKTR	INUG-14.17	N	499	1525
INUG	INUG-14	19-Aug-11	76	NIGHT	LKTR	INUG-14.18	Y	542	1506
INUG	INUG-14	19-Aug-11	76	NIGHT	LKTR	INUG-14.19	Y	411	720
INUG	INUG-01	17-Aug-11	51	DAY	RNWH	INUG-01.11	Y	413	656
INUG	INUG-01	17-Aug-11	51	DAY	RNWH	INUG-01.12	Y	408	664
INUG	INUG-01	17-Aug-11	51	DAY	RNWH	INUG-01.13	Y	400	642
INUG	INUG-01	17-Aug-11	51	DAY	RNWH	INUG-01.14	Y	360	488
INUG	INUG-01	17-Aug-11	51	DAY	RNWH	INUG-01.15	Y	310	292
INUG	INUG-01	17-Aug-11	76	DAY	RNWH	INUG-01.18	Y	430	826
INUG	INUG-01	17-Aug-11	76	DAY	RNWH	INUG-01.19	Y	430	884
INUG	INUG-01	17-Aug-11	76	DAY	RNWH	INUG-01.20	Y	370	578
INUG	INUG-01	17-Aug-11	76	DAY	RNWH	INUG-01.21	Y	385	652
INUG	INUG-01	17-Aug-11	76	DAY	RNWH	INUG-01.22	Y	388	530
INUG	INUG-02	17-Aug-11	38	NIGHT	RNWH	INUG-02.04	Y	241	126
INUG	INUG-02	17-Aug-11	51	NIGHT	RNWH	INUG-02.08	Y	332	328
INUG	INUG-02	17-Aug-11	76	NIGHT	RNWH	INUG-02.14	Y	406	716
INUG	INUG-03	17-Aug-11	51	DAY	RNWH	INUG-03.01	Y	362	542
INUG	INUG-05	17-Aug-11	51	DAY	RNWH	INUG-05.01	Y	382	600
INUG	INUG-05	17-Aug-11	51	DAY	RNWH	INUG-05.02	N	365	425
INUG	INUG-08	18-Aug-11	51	NIGHT	RNWH	INUG-08.04	Y	370	528
INUG	INUG-08	18-Aug-11	51	NIGHT	RNWH	INUG-08.05	Y	392	600
INUG	INUG-08	18-Aug-11	51	NIGHT	RNWH	INUG-08.08	Y	384	572
INUG	INUG-08	18-Aug-11	51	NIGHT	RNWH	INUG-08.09	Y	376	536
INUG	INUG-08	18-Aug-11	51	NIGHT	RNWH	INUG-08.11	Y	370	500
INUG	INUG-08	18-Aug-11	51	NIGHT	RNWH	INUG-08.12	Y	365	468
INUG	INUG-08	18-Aug-11	51	NIGHT	RNWH	INUG-08.13	Y	281	204
INUG	INUG-08	18-Aug-11	51	NIGHT	RNWH	INUG-08.14	Y	282	234
INUG	INUG-08	18-Aug-11	76	NIGHT	RNWH	INUG-08.16	Y	364	469
INUG	INUG-09	18-Aug-11	76	DAY	RNWH	INUG-09.04	Y	420	722
INUG	INUG-13	19-Aug-11	76	NIGHT	RNWH	INUG-13.02	Y	398	664
INUG	INUG-13	19-Aug-11	76	NIGHT	RNWH	INUG-13.04	Y	420	748
INUG	INUG-13	19-Aug-11	51	NIGHT	RNWH	INUG-13.09	Y	385	596
INUG	INUG-13	19-Aug-11	51	NIGHT	RNWH	INUG-13.12	Y	385	602
INUG	INUG-13	19-Aug-11	51	NIGHT	RNWH	INUG-13.14	Y	352	434
INUG	INUG-13	19-Aug-11	76	NIGHT	RNWH	INUG-13.18	Y	400	634
INUG	INUG-13	19-Aug-11	76	NIGHT	RNWH	INUG-13.19	Y	375	512
INUG	INUG-14	19-Aug-11	51	NIGHT	RNWH	INUG-14.02	Y	405	646
INUG	INUG-14	19-Aug-11	51	NIGHT	RNWH	INUG-14.08	Y	375	502
INUG	INUG-14	19-Aug-11	51	NIGHT	RNWH	INUG-14.09	Y	383	602
INUG	INUG-14	19-Aug-11	51	NIGHT	RNWH	INUG-14.10	Y	343	436
INUG	INUG-14	19-Aug-11	76	NIGHT	RNWH	INUG-14.11	Y	375	532
INUG	INUG-14	19-Aug-11	76	NIGHT	RNWH	INUG-14.14	Y	385	570

Lake	Set ID	Date	K	Weighed with 25 g scale	Sex	Maturity	Repro Status	Age Structure
INUG	INUG-02	17-Aug-11	0.98562211	N	F	IM	UD	PEC/OTO
INUG	INUG-07	18-Aug-11	1.169774796	Y				PEC
INUG	INUG-07	18-Aug-11	1.185185185	N	UN	IM	UD	PEC/OTO
INUG	INUG-08	18-Aug-11	0.648902688	N (± 10)				PEC
INUG	INUG-14	19-Aug-11	0.926253145	Y				PEC
INUG	INUG-01	17-Aug-11	0.834575066	Y				
INUG	INUG-01	17-Aug-11	1.078518519	Y				PEC
INUG	INUG-01	17-Aug-11	1.140136941	Y				PEC
INUG	INUG-01	17-Aug-11	1.358714556	Y				PEC
INUG	INUG-01	17-Aug-11	1.009413754	Y				PEC
INUG	INUG-01	17-Aug-11	1.029806396	Y				PEC
INUG	INUG-01	17-Aug-11	0.972250796	Y				PEC
INUG	INUG-01	17-Aug-11	0.940356562	N	F	MA	RS	PEC/OTO
INUG	INUG-01	17-Aug-11	0.968119454	N	F	IM	UD	PEC/OTO
INUG	INUG-01	17-Aug-11	1.071757135	N	F	IM	UD	PEC/OTO
INUG	INUG-01	17-Aug-11	0.881484243	N	M	IM	UD	PEC/OTO
INUG	INUG-01	17-Aug-11	1.129138456	N	M	MA	RI	PEC/OTO
INUG	INUG-02	17-Aug-11	0.973448357	Y				PEC
INUG	INUG-02	17-Aug-11	0.978635123	N	UN	IM	UD	PEC/OTO
INUG	INUG-02	17-Aug-11	1.000823045	N	UN	IM	UD	PEC/OTO
INUG	INUG-02	17-Aug-11	0.978336467	Y				PEC
INUG	INUG-02	17-Aug-11	1.104592022	Y				PEC
INUG	INUG-02	17-Aug-11	0.936708891	Y				PEC
INUG	INUG-02	17-Aug-11	0.906677313	Y				PEC
INUG	INUG-02	17-Aug-11	1.005390803	Y				PEC
INUG	INUG-02	17-Aug-11	0.977236428	N	M	IM	UD	PEC/OTO
INUG	INUG-02	17-Aug-11	1.059352795	N	F	MA	RS	PEC/OTO
INUG	INUG-02	17-Aug-11	0.939549927	N	M	IM	UD	PEC/OTO
INUG	INUG-02	17-Aug-11	0.976857524	N	F	MA	RS	PEC/OTO
INUG	INUG-02	17-Aug-11	0.862403428	Y				PEC
INUG	INUG-02	17-Aug-11	1.085474511	Y				PEC
INUG	INUG-02	17-Aug-11	1.02511299	N	UN	IM	UD	PEC/OTO
INUG	INUG-04	17-Aug-11	1.272979222	Y				PEC
INUG	INUG-04	17-Aug-11	0.999506628	Y				PEC
INUG	INUG-04	17-Aug-11	0.941861153	N	M	IM	UD	PEC/OTO
INUG	INUG-05	17-Aug-11	1.485961005	Y				PEC
INUG	INUG-05	17-Aug-11	0.940111097	Y				PEC
INUG	INUG-05	17-Aug-11	0.998336238	N	M	MA	RS	PEC/OTO
INUG	INUG-05	17-Aug-11	1.138465925	N	M	MA	RS	PEC/OTO
INUG	INUG-05	17-Aug-11	1.122825153	N	M	MA	RS	PEC/OTO
INUG	INUG-05	17-Aug-11	1.03740365	N	M	MA	RS	PEC/OTO
INUG	INUG-06	17-Aug-11	1	Y				PEC
INUG	INUG-07	18-Aug-11	0.948259227	N	M	MA	RS	PEC/OTO
INUG	INUG-07	18-Aug-11	0.789708841	N	M	IM	UD	PEC/OTO
INUG	INUG-07	18-Aug-11	1.007245769	Y				PEC
INUG	INUG-07	18-Aug-11	1.135174619	Y				PEC
INUG	INUG-07	18-Aug-11	0.944904157	N	F	IM	UD	PEC/OTO
INUG	INUG-07	18-Aug-11	1.007253433	Y				PEC
INUG	INUG-07	18-Aug-11	1.184759229	Y				PEC
INUG	INUG-07	18-Aug-11	0.960898742	N	F	IM	UD	PEC/OTO
INUG	INUG-07	18-Aug-11	0.894943275	N	UN	IM	UD	PEC/OTO
INUG	INUG-07	18-Aug-11	1.043924435	Y				PEC
INUG	INUG-07	18-Aug-11	1.009958279	N	F	IM	UD	PEC/OTO
INUG	INUG-07	18-Aug-11	1.361397556	N (± 10)				PEC
INUG	INUG-08	18-Aug-11	1.067772288	Y				PEC
INUG	INUG-08	18-Aug-11	0.979030158	Y				PEC
INUG	INUG-08	18-Aug-11	0.919700944	Y				PEC
INUG	INUG-08	18-Aug-11	1.001542375	N	M	IM	UD	PEC/OTO
INUG	INUG-08	18-Aug-11	1.079200107	Y				PEC
INUG	INUG-08	18-Aug-11	1.070873056	N	UN	IM	UD	PEC/OTO
INUG	INUG-08	18-Aug-11	0.946622532	N	M	MA	RI	PEC/OTO
INUG	INUG-08	18-Aug-11	0.956757945	N	M	MA	RS	PEC/OTO
INUG	INUG-08	18-Aug-11	1.037296188	N	M	MA	RS	PEC/OTO
INUG	INUG-08	18-Aug-11	1.007476393	N	F	MA	RI	PEC/OTO
INUG	INUG-09	18-Aug-11	1.229697422	Y				PEC
INUG	INUG-09	18-Aug-11	0.880699708	Y				PEC
INUG	INUG-09	18-Aug-11	1.037455808	Y				PEC
INUG	INUG-09	18-Aug-11	1.247469859	Y				PEC
INUG	INUG-10	18-Aug-11	1.113005361	Y				PEC

Lake	Set ID	Date	K	Weighed with 25 g scale	Sex	Maturity	Repro Status	Age Structure
INUG	INUG-10	18-Aug-11	1.181558042	N	F	MA	RI	PEC/OTO
INUG	INUG-10	18-Aug-11	0.902513205	N	F	MA	RS	PEC/OTO
INUG	INUG-13	19-Aug-11	0.963262009	Y				PEC
INUG	INUG-13	19-Aug-11	1.161307388	Y				PEC
INUG	INUG-13	19-Aug-11	0.990982304	N	F	MA	RS	PEC/OTO
INUG	INUG-13	19-Aug-11	1.014709072	N	F	MA	RS	PEC/OTO
INUG	INUG-13	19-Aug-11	0.988003355	Y				PEC
INUG	INUG-13	19-Aug-11	1.019548377	N	M	IM	UD	PEC/OTO
INUG	INUG-13	19-Aug-11	0.973495254	Y				PEC
INUG	INUG-13	19-Aug-11	1.160749264	Y				PEC
INUG	INUG-13	19-Aug-11		Y				PEC
INUG	INUG-13	19-Aug-11	1.076498635	Y				PEC
INUG	INUG-13	19-Aug-11	1.113580222	N	F	MA	RI	PEC/OTO
INUG	INUG-14	19-Aug-11	1.050068548	N	F	IM	UD	PEC/OTO
INUG	INUG-14	19-Aug-11	1.076939811	N	M	IM	UD	PEC/OTO
INUG	INUG-14	19-Aug-11	0.979183843	N	F	IM	UD	PEC/OTO
INUG	INUG-14	19-Aug-11	1.185185185	Y				PEC
INUG	INUG-14	19-Aug-11	0.8849556	Y				PEC
INUG	INUG-14	19-Aug-11	1.1812864	N	M	MA	RS	PEC/OTO
INUG	INUG-14	19-Aug-11	1.086490264	N	F	MA	RI	PEC/OTO
INUG	INUG-14	19-Aug-11	0.988733434	N	F	MA	RS	PEC/OTO
INUG	INUG-14	19-Aug-11	1.227349378	Y				PEC
INUG	INUG-14	19-Aug-11	0.94586055	N	M	MA	RS	PEC/OTO
INUG	INUG-14	19-Aug-11	1.037067515	N	F	IM	UD	PEC/OTO
INUG	INUG-01	17-Aug-11	0.93122298	N	M	MA	RI	PEC/OTO
INUG	INUG-01	17-Aug-11	0.977659422	N	F	MA	RI	PEC/OTO
INUG	INUG-01	17-Aug-11	1.003125	N	F	MA	RI	PEC/OTO
INUG	INUG-01	17-Aug-11	1.045953361	N	F	MA	RI	PEC/OTO
INUG	INUG-01	17-Aug-11	0.980161794	N	M	MA	RI	PEC/OTO
INUG	INUG-01	17-Aug-11	1.038902235	N	F	MA	RI	PEC/OTO
INUG	INUG-01	17-Aug-11	1.111851787	N	M	MA	RI	PEC/OTO
INUG	INUG-01	17-Aug-11	1.14109727	N	M	MA	RI	PEC/OTO
INUG	INUG-01	17-Aug-11	1.142524199	N	M	MA	RI	PEC/OTO
INUG	INUG-01	17-Aug-11	0.907362221	N	F	MA	RI	PEC/OTO
INUG	INUG-02	17-Aug-11	0.900159393	N	UN	IM	UD	PEC/OTO
INUG	INUG-02	17-Aug-11	0.896312788	N	F	IM	UD	PEC/OTO
INUG	INUG-02	17-Aug-11	1.069879637	N	F	MA	RI	PEC/OTO
INUG	INUG-03	17-Aug-11	1.142545686	N	F	MA	RI	PEC/OTO
INUG	INUG-05	17-Aug-11	1.076368951	N	F	MA	RI	PEC/OTO
INUG	INUG-05	17-Aug-11	0.873997794	Y				PEC
INUG	INUG-08	18-Aug-11	1.042386433	N	F	MA	RI	PEC/OTO
INUG	INUG-08	18-Aug-11	0.996077315	N	M	MA	RI	PEC/OTO
INUG	INUG-08	18-Aug-11	1.01018835	N	M	MA	RI	PEC/OTO
INUG	INUG-08	18-Aug-11	1.008326671	N	M	MA	RI	PEC/OTO
INUG	INUG-08	18-Aug-11	0.987108365	N	F	MA	RI	PEC/OTO
INUG	INUG-08	18-Aug-11	0.962425807	N	F	MA	RI	PEC/OTO
INUG	INUG-08	18-Aug-11	0.919414202	N	F	IM	UD	PEC/OTO
INUG	INUG-08	18-Aug-11	1.043442526	N	F	IM	UD	PEC/OTO
INUG	INUG-08	18-Aug-11	0.97245316	N	M	MA	RI	PEC/OTO
INUG	INUG-09	18-Aug-11	0.974516791	N	M	MA	RI	PEC/OTO
INUG	INUG-13	19-Aug-11	1.053219432	N	M	MA	RI	PEC/OTO
INUG	INUG-13	19-Aug-11	1.009610193	N	M	MA	RI	PEC/OTO
INUG	INUG-13	19-Aug-11	1.044393286	N	M	MA	RI	PEC/OTO
INUG	INUG-13	19-Aug-11	1.054907312	N	F	MA	RI	PEC/OTO
INUG	INUG-13	19-Aug-11	0.995088573	N	M	MA	RI	PEC/OTO
INUG	INUG-13	19-Aug-11	0.990625	N	M	MA	RI	PEC/OTO
INUG	INUG-13	19-Aug-11	0.970903704	N	F	MA	RI	PEC/OTO
INUG	INUG-14	19-Aug-11	0.972450375	N	F	MA	RI	PEC/OTO
INUG	INUG-14	19-Aug-11	0.951940741	N	M	MA	RI	PEC/OTO
INUG	INUG-14	19-Aug-11	1.071519723	N	M	MA	RI	PEC/OTO
INUG	INUG-14	19-Aug-11	1.080448645	N	M	MA	RI	PEC/OTO
INUG	INUG-14	19-Aug-11	1.00882963	N	M	MA	RI	PEC/OTO
INUG	INUG-14	19-Aug-11	0.998832505	N	M	MA	RI	PEC/OTO

Lake	Set ID	Date	Age.OTO.yr	Age.PEC.yr	Ext. DELT	Ext. Parasites	Tagged?	Tag ID
INUG	INUG-02	17-Aug-11	ns	4	NONE	NONE	N	
INUG	INUG-07	18-Aug-11		14	NONE	NONE	N	
INUG	INUG-07	18-Aug-11	5	2	NONE	NONE	N	
INUG	INUG-08	18-Aug-11		4	damaged caudal fin	NONE	N	
INUG	INUG-14	19-Aug-11		3	NONE	NONE	N	
INUG	INUG-01	17-Aug-11			NONE	NONE	N	
INUG	INUG-01	17-Aug-11		24	NONE	NONE	Y	87908
INUG	INUG-01	17-Aug-11		22	NONE	NONE	Y	87909
INUG	INUG-01	17-Aug-11		27	NONE	NONE	Y	87910
INUG	INUG-01	17-Aug-11		14	NONE	NONE	Y	87911
INUG	INUG-01	17-Aug-11		16	NONE	NONE	Y	87912
INUG	INUG-01	17-Aug-11		17	NONE	NONE	Y	87913
INUG	INUG-01	17-Aug-11	ns	17	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	10	5	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	7	5	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	19	17	Tiny lower lobe of caudal fin	NONE	N	
INUG	INUG-01	17-Aug-11	17	14	NONE	NONE	N	
INUG	INUG-02	17-Aug-11		8	NONE	NONE	N	
INUG	INUG-02	17-Aug-11	8	3	NONE	NONE	N	
INUG	INUG-02	17-Aug-11	13	3	NONE	NONE	N	
INUG	INUG-02	17-Aug-11		15	NONE	NONE	Y	87942
INUG	INUG-02	17-Aug-11		14	NONE	NONE	Y	87915
INUG	INUG-02	17-Aug-11		16	NONE	NONE	Y	87916
INUG	INUG-02	17-Aug-11		22	slight dorsal fin deformity	NONE	Y	87917
INUG	INUG-02	17-Aug-11		17	slight dorsal fin deformity	NONE	Y	87919
INUG	INUG-02	17-Aug-11	12	7	NONE	NONE	N	
INUG	INUG-02	17-Aug-11	21	12	bi-lobed adipose fin; deformed dorsal	NONE	N	
INUG	INUG-02	17-Aug-11	22	16	NONE	NONE	N	
INUG	INUG-02	17-Aug-11	26	13	NONE	NONE	N	
INUG	INUG-02	17-Aug-11		30+	NONE	NONE	Y	87920
INUG	INUG-02	17-Aug-11		16	NONE	NONE	Y	87921
INUG	INUG-02	17-Aug-11	4	2	NONE	NONE	N	
INUG	INUG-04	17-Aug-11		7	NONE	NONE	N	
INUG	INUG-04	17-Aug-11		20	NONE	NONE	Y	87924
INUG	INUG-04	17-Aug-11	9	9	NONE	NONE	N	
INUG	INUG-05	17-Aug-11		19	NONE	NONE	Y	87922
INUG	INUG-05	17-Aug-11		9	NONE	NONE	Y	87923
INUG	INUG-05	17-Aug-11	18	15	NONE	NONE	N	
INUG	INUG-05	17-Aug-11	17	14	NONE	NONE	N	
INUG	INUG-05	17-Aug-11	23	19	NONE	NONE	N	
INUG	INUG-05	17-Aug-11	18	17	NONE	NONE	N	
INUG	INUG-06	17-Aug-11		5	NONE	NONE	N	
INUG	INUG-07	18-Aug-11	18	13	NONE	NONE	N	
INUG	INUG-07	18-Aug-11	17	13	NONE	NONE	N	
INUG	INUG-07	18-Aug-11		15	NONE	NONE	Y	87925
INUG	INUG-07	18-Aug-11		14	NONE	NONE	Y	87438
INUG	INUG-07	18-Aug-11	10	5	NONE	NONE	N	
INUG	INUG-07	18-Aug-11		17	NONE	NONE	Y	87439
INUG	INUG-07	18-Aug-11		4	NONE	NONE	N	
INUG	INUG-07	18-Aug-11	6	1	NONE	NONE	N	
INUG	INUG-07	18-Aug-11	7	1	NONE	NONE	N	
INUG	INUG-07	18-Aug-11		12	NONE	NONE	Y	87440
INUG	INUG-07	18-Aug-11	8	4	NONE	NONE	N	
INUG	INUG-07	18-Aug-11		5	NONE	NONE	N	
INUG	INUG-08	18-Aug-11		15	NONE	NONE	Y	87441
INUG	INUG-08	18-Aug-11		18	NONE	NONE	Y	87442
INUG	INUG-08	18-Aug-11		13	NONE	NONE	Y	87443
INUG	INUG-08	18-Aug-11	8	5	NONE	NONE	N	
INUG	INUG-08	18-Aug-11		15	scar on stomach	NONE	Y	87444
INUG	INUG-08	18-Aug-11	8	3	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	22	16	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	28	20	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	21	12	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	28	13	NONE	NONE	N	
INUG	INUG-09	18-Aug-11		6	NONE	NONE	Y	87445
INUG	INUG-09	18-Aug-11		28	NONE	NONE	Y	87446
INUG	INUG-09	18-Aug-11		19	NONE	NONE	Y	48476
INUG	INUG-09	18-Aug-11		12	NONE	NONE	Y	48477
INUG	INUG-10	18-Aug-11		27	missing adipose fin	NONE	Y	48478

Lake	Set ID	Date	Age.OTO.yr	Age.PEC.yr	Ext. DELT	Ext. Parasites	Tagged?	Tag ID
INUG	INUG-10	18-Aug-11	22	15	NONE	NONE	N	
INUG	INUG-10	18-Aug-11	23	19	NONE	NONE	N	
INUG	INUG-13	19-Aug-11		13	NONE	NONE	Y	48479
INUG	INUG-13	19-Aug-11		17	NONE	NONE	Y	48480
INUG	INUG-13	19-Aug-11	40	25	NONE	NONE	N	
INUG	INUG-13	19-Aug-11	17	14	NONE	NONE	N	
INUG	INUG-13	19-Aug-11		4	NONE	NONE	N	
INUG	INUG-13	19-Aug-11	11	6	NONE	NONE	N	
INUG	INUG-13	19-Aug-11		14	NONE	NONE	Y	48481
INUG	INUG-13	19-Aug-11		4	NONE	NONE	N	
INUG	INUG-13	19-Aug-11		3	NONE	NONE	N	
INUG	INUG-13	19-Aug-11		3	NONE	NONE	N	
INUG	INUG-13	19-Aug-11	20	14	NONE	NONE	N	
INUG	INUG-14	19-Aug-11	10	5	NONE	NONE	N	
INUG	INUG-14	19-Aug-11	13	9	NONE	NONE	N	
INUG	INUG-14	19-Aug-11	9	4	NONE	NONE	N	
INUG	INUG-14	19-Aug-11		8	NONE	NONE	Y	48482
INUG	INUG-14	19-Aug-11		20	NONE	NONE	Y	48483
INUG	INUG-14	19-Aug-11	24	21	NONE	NONE	N	
INUG	INUG-14	19-Aug-11	20	11	NONE	NONE	N	
INUG	INUG-14	19-Aug-11	14	14	NONE	NONE	N	
INUG	INUG-14	19-Aug-11		14	NONE	NONE	Y	48484
INUG	INUG-14	19-Aug-11	21	16	NONE	NONE	N	
INUG	INUG-14	19-Aug-11	9	7	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	24	14	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	16	10	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	15	12	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	9	8	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	9	6	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	19	13	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	14	16	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	16	13	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	14	14	NONE	NONE	N	
INUG	INUG-01	17-Aug-11	13	9	NONE	NONE	N	
INUG	INUG-02	17-Aug-11	6	4	NONE	NONE	N	
INUG	INUG-02	17-Aug-11	11	9	NONE	NONE	N	
INUG	INUG-02	17-Aug-11	13	10	NONE	NONE	N	
INUG	INUG-03	17-Aug-11	9	11	NONE	NONE	N	
INUG	INUG-05	17-Aug-11	15	14	NONE	NONE	N	
INUG	INUG-05	17-Aug-11		8	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	16	13	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	15	15	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	17	12	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	16	11	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	20	12	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	18	14	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	7	5	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	8	7	NONE	NONE	N	
INUG	INUG-08	18-Aug-11	13	12	NONE	NONE	N	
INUG	INUG-09	18-Aug-11	19	14	NONE	NONE	N	
INUG	INUG-13	19-Aug-11	15	14	NONE	NONE	N	
INUG	INUG-13	19-Aug-11	17	14	NONE	NONE	N	
INUG	INUG-13	19-Aug-11	22	14	NONE	NONE	N	
INUG	INUG-13	19-Aug-11	15	14	NONE	NONE	N	
INUG	INUG-13	19-Aug-11	15	12	NONE	NONE	N	
INUG	INUG-13	19-Aug-11	19	14	NONE	NONE	N	
INUG	INUG-13	19-Aug-11	17	13	NONE	NONE	N	
INUG	INUG-14	19-Aug-11	12	12	NONE	NONE	N	
INUG	INUG-14	19-Aug-11	13	12	NONE	NONE	N	
INUG	INUG-14	19-Aug-11	18	13	NONE	NONE	N	
INUG	INUG-14	19-Aug-11	10	8	NONE	NONE	N	
INUG	INUG-14	19-Aug-11	17	12	NONE	NONE	N	
INUG	INUG-14	19-Aug-11	12	12	NONE	NONE	N	

Lake	Set ID	Date	RECAP?	Gonad weight (g)	Gonad appearance	Weight of 400 eggs (g)	Liver weight (g)
INUG	INUG-02	17-Aug-11	N	0.36	NORMAL		2.15
INUG	INUG-07	18-Aug-11	N				
INUG	INUG-07	18-Aug-11	N		NORMAL		0.46
INUG	INUG-08	18-Aug-11	N				
INUG	INUG-14	19-Aug-11	N				
INUG	INUG-01	17-Aug-11	N				
INUG	INUG-01	17-Aug-11	N				
INUG	INUG-01	17-Aug-11	N				
INUG	INUG-01	17-Aug-11	N				
INUG	INUG-01	17-Aug-11	N				
INUG	INUG-01	17-Aug-11	N				
INUG	INUG-01	17-Aug-11	N		NORMAL		13.75
INUG	INUG-01	17-Aug-11	N		NORMAL		2
INUG	INUG-01	17-Aug-11	N		NORMAL		3.26
INUG	INUG-01	17-Aug-11	N		NORMAL		11.44
INUG	INUG-01	17-Aug-11	N	31.24	NORMAL		11.39
INUG	INUG-02	17-Aug-11	N				
INUG	INUG-02	17-Aug-11	N		NORMAL		0.86
INUG	INUG-02	17-Aug-11	N		NORMAL		1.45
INUG	INUG-02	17-Aug-11	N				
INUG	INUG-02	17-Aug-11	N				
INUG	INUG-02	17-Aug-11	N				
INUG	INUG-02	17-Aug-11	N				
INUG	INUG-02	17-Aug-11	N	0.42	NORMAL		5.06
INUG	INUG-02	17-Aug-11	N	35.53	NORMAL		59.68
INUG	INUG-02	17-Aug-11	N	1.69	NORMAL		14.49
INUG	INUG-02	17-Aug-11	N	12.1	NORMAL		20.44
INUG	INUG-02	17-Aug-11	N				
INUG	INUG-02	17-Aug-11	N		NORMAL		0.45
INUG	INUG-04	17-Aug-11	N				
INUG	INUG-04	17-Aug-11	N				
INUG	INUG-04	17-Aug-11	N		NORMAL		4.15
INUG	INUG-05	17-Aug-11	N				
INUG	INUG-05	17-Aug-11	N				
INUG	INUG-05	17-Aug-11	N		NORMAL		14.84
INUG	INUG-05	17-Aug-11	N		NORMAL		20.73
INUG	INUG-05	17-Aug-11	N		NORMAL		68.46
INUG	INUG-05	17-Aug-11	N		NORMAL		13
INUG	INUG-06	17-Aug-11	N				
INUG	INUG-07	18-Aug-11	N		NORMAL		14.24
INUG	INUG-07	18-Aug-11	N		NORMAL		8.63
INUG	INUG-07	18-Aug-11	N				
INUG	INUG-07	18-Aug-11	N				
INUG	INUG-07	18-Aug-11	N		NORMAL		1.56
INUG	INUG-07	18-Aug-11	N				
INUG	INUG-07	18-Aug-11	N		NORMAL		2.18
INUG	INUG-07	18-Aug-11	N		NORMAL		1.24
INUG	INUG-07	18-Aug-11	N				
INUG	INUG-07	18-Aug-11	N		NORMAL		1.32
INUG	INUG-07	18-Aug-11	N				
INUG	INUG-08	18-Aug-11	N				
INUG	INUG-08	18-Aug-11	N				
INUG	INUG-08	18-Aug-11	N		NORMAL		1.93
INUG	INUG-08	18-Aug-11	N				
INUG	INUG-08	18-Aug-11	N		NORMAL		1.49
INUG	INUG-08	18-Aug-11	N	42.79	NORMAL		11.2
INUG	INUG-08	18-Aug-11	N		NORMAL		19.25
INUG	INUG-08	18-Aug-11	N		NORMAL		18.81
INUG	INUG-08	18-Aug-11	N	194.43	NORMAL	35.08	34.39
INUG	INUG-09	18-Aug-11	N				
INUG	INUG-09	18-Aug-11	N				
INUG	INUG-09	18-Aug-11	N				
INUG	INUG-09	18-Aug-11	N				
INUG	INUG-10	18-Aug-11	N				

Lake	Set ID	Date	RECAP?	Gonad weight (g)	Gonad appearance	Weight of 400 eggs (g)	Liver weight (g)
INUG	INUG-10	18-Aug-11	N	191.21	NORMAL	29.07	40.93
INUG	INUG-10	18-Aug-11	N		NORMAL		16.49
INUG	INUG-13	19-Aug-11	N				
INUG	INUG-13	19-Aug-11	N				
INUG	INUG-13	19-Aug-11	N		NORMAL		41.68
INUG	INUG-13	19-Aug-11	N		NORMAL		16.56
INUG	INUG-13	19-Aug-11	N				
INUG	INUG-13	19-Aug-11	N		NORMAL		4.54
INUG	INUG-13	19-Aug-11	N				
INUG	INUG-13	19-Aug-11	N				
INUG	INUG-13	19-Aug-11	N				
INUG	INUG-13	19-Aug-11	N	117.88	NORMAL	22.98	21.07
INUG	INUG-14	19-Aug-11	N		NORMAL		2.54
INUG	INUG-14	19-Aug-11	N		NORMAL		5.37
INUG	INUG-14	19-Aug-11	N		NORMAL		1.34
INUG	INUG-14	19-Aug-11	N				
INUG	INUG-14	19-Aug-11	N				
INUG	INUG-14	19-Aug-11	N		NORMAL		44.09
INUG	INUG-14	19-Aug-11	N	226	NORMAL	29.57	42.75
INUG	INUG-14	19-Aug-11	N		NORMAL		16.11
INUG	INUG-14	19-Aug-11	N				
INUG	INUG-14	19-Aug-11	N		NORMAL		17.06
INUG	INUG-14	19-Aug-11	N		NORMAL		8.68
INUG	INUG-01	17-Aug-11	N	23.72	NORMAL		5.13
INUG	INUG-01	17-Aug-11	N	24.64	NORMAL	0.89	7.25
INUG	INUG-01	17-Aug-11	N	22.2	NORMAL	0.94	8.73
INUG	INUG-01	17-Aug-11	N	11.13	NORMAL	0.97	4.38
INUG	INUG-01	17-Aug-11	N	3.28	NORMAL		1.6
INUG	INUG-01	17-Aug-11	N	30.91	NORMAL	0.99	9.98
INUG	INUG-01	17-Aug-11	N	32.28	NORMAL		8.14
INUG	INUG-01	17-Aug-11	N	22.03	NORMAL		2.21
INUG	INUG-01	17-Aug-11	N	25.02	NORMAL		2.58
INUG	INUG-01	17-Aug-11	N	16.86	NORMAL	0.97	2.65
INUG	INUG-02	17-Aug-11	N	0.07	NORMAL		0.79
INUG	INUG-02	17-Aug-11	N	1.15	NORMAL		2.26
INUG	INUG-02	17-Aug-11	N	33.69	NORMAL	1.25	10.06
INUG	INUG-03	17-Aug-11	N	19.23	NORMAL	0.91	6.85
INUG	INUG-05	17-Aug-11	N	23.09	NORMAL	0.92	8.23
INUG	INUG-05	17-Aug-11	N				
INUG	INUG-08	18-Aug-11	N	20.21	NORMAL	1.06	6.54
INUG	INUG-08	18-Aug-11	N	21.4	NORMAL		3.39
INUG	INUG-08	18-Aug-11	N	15.71	NORMAL		4.07
INUG	INUG-08	18-Aug-11	N	14.39	NORMAL		4.36
INUG	INUG-08	18-Aug-11	N	15.85	NORMAL	0.79	4.19
INUG	INUG-08	18-Aug-11	N	14.11	NORMAL	0.73	6.88
INUG	INUG-08	18-Aug-11	N		NORMAL		1.1
INUG	INUG-08	18-Aug-11	N		NORMAL		1.81
INUG	INUG-08	18-Aug-11	N	4.06	NORMAL		3.65
INUG	INUG-09	18-Aug-11	N	27.82	NORMAL		4.16
INUG	INUG-13	19-Aug-11	N	20.68	NORMAL		4.81
INUG	INUG-13	19-Aug-11	N	28.8	NORMAL		3
INUG	INUG-13	19-Aug-11	N	20.66	NORMAL		4.7
INUG	INUG-13	19-Aug-11	N	22.18	NORMAL	0.47	5.53
INUG	INUG-13	19-Aug-11	N	12.68	NORMAL		4.27
INUG	INUG-13	19-Aug-11	N	23.62	NORMAL		4.62
INUG	INUG-13	19-Aug-11	N	17	NORMAL	0.85	5.37
INUG	INUG-14	19-Aug-11	N	27.85	NORMAL	1.81	10.01
INUG	INUG-14	19-Aug-11	N	12.54	NORMAL		3.86
INUG	INUG-14	19-Aug-11	N	16.42	NORMAL		5.46
INUG	INUG-14	19-Aug-11	N	7.99	NORMAL		3.25
INUG	INUG-14	19-Aug-11	N	15.11	NORMAL		3.34
INUG	INUG-14	19-Aug-11	N	14.75	NORMAL		3.32

Lake	Set ID	Date	Liver appearance	Int. DELT	Int. Parasites
INUG	INUG-02	17-Aug-11	NORMAL	NONE	LOW
INUG	INUG-07	18-Aug-11			
INUG	INUG-07	18-Aug-11	NORMAL	NONE	HIGH
INUG	INUG-08	18-Aug-11			
INUG	INUG-14	19-Aug-11			
INUG	INUG-01	17-Aug-11			
INUG	INUG-01	17-Aug-11			
INUG	INUG-01	17-Aug-11			
INUG	INUG-01	17-Aug-11			
INUG	INUG-01	17-Aug-11			
INUG	INUG-01	17-Aug-11	NORMAL	NONE	LOW
INUG	INUG-01	17-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-01	17-Aug-11	NORMAL	NONE	LOW
INUG	INUG-01	17-Aug-11	NORMAL	NONE	HIGH
INUG	INUG-01	17-Aug-11	NORMAL	NONE	LOW
INUG	INUG-02	17-Aug-11			
INUG	INUG-02	17-Aug-11	2 PARASITE CYSTS	NONE	LOW
INUG	INUG-02	17-Aug-11	NORMAL	NONE	LOW
INUG	INUG-02	17-Aug-11			
INUG	INUG-02	17-Aug-11			
INUG	INUG-02	17-Aug-11			
INUG	INUG-02	17-Aug-11			
INUG	INUG-02	17-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-02	17-Aug-11	NORMAL	NONE	LOW
INUG	INUG-02	17-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-02	17-Aug-11	NORMAL	NONE	LOW
INUG	INUG-02	17-Aug-11			
INUG	INUG-02	17-Aug-11			
INUG	INUG-02	17-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-04	17-Aug-11			
INUG	INUG-04	17-Aug-11			
INUG	INUG-04	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-05	17-Aug-11			
INUG	INUG-05	17-Aug-11			
INUG	INUG-05	17-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-05	17-Aug-11	SLIGHTLY LIGHT COLOURED	NONE	NONE
INUG	INUG-05	17-Aug-11	DISCOLOURED & BLOTCHY	NONE	MODERATE
INUG	INUG-05	17-Aug-11	NORMAL	NONE	HIGH
INUG	INUG-06	17-Aug-11			
INUG	INUG-07	18-Aug-11	NORMAL	NONE	LOW
INUG	INUG-07	18-Aug-11	NORMAL	NONE	LOW
INUG	INUG-07	18-Aug-11			
INUG	INUG-07	18-Aug-11			
INUG	INUG-07	18-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-07	18-Aug-11			
INUG	INUG-07	18-Aug-11	NORMAL	NONE	LOW
INUG	INUG-07	18-Aug-11	NORMAL	NONE	LOW
INUG	INUG-07	18-Aug-11			
INUG	INUG-07	18-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-07	18-Aug-11			
INUG	INUG-08	18-Aug-11			
INUG	INUG-08	18-Aug-11			
INUG	INUG-08	18-Aug-11			
INUG	INUG-08	18-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-08	18-Aug-11			
INUG	INUG-08	18-Aug-11	NORMAL	NONE	LOW
INUG	INUG-08	18-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-08	18-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-08	18-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-08	18-Aug-11	NORMAL	NONE	LOW
INUG	INUG-09	18-Aug-11			
INUG	INUG-09	18-Aug-11			
INUG	INUG-09	18-Aug-11			
INUG	INUG-09	18-Aug-11			
INUG	INUG-10	18-Aug-11			

Lake	Set ID	Date	Liver appearance	Int. DELT	Int. Parasites
INUG	INUG-10	18-Aug-11	NORMAL	NONE	LOW
INUG	INUG-10	18-Aug-11	NORMAL	NONE	LOW
INUG	INUG-13	19-Aug-11			
INUG	INUG-13	19-Aug-11			
INUG	INUG-13	19-Aug-11	NORMAL	NONE	LOW
INUG	INUG-13	19-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-13	19-Aug-11			
INUG	INUG-13	19-Aug-11	NORMAL	NONE	HIGH
INUG	INUG-13	19-Aug-11			
INUG	INUG-13	19-Aug-11			
INUG	INUG-13	19-Aug-11			
INUG	INUG-13	19-Aug-11	NORMAL	NONE	MODERATE
INUG	INUG-14	19-Aug-11	NORMAL	NONE	MODERATE (with cestodes in gut)
INUG	INUG-14	19-Aug-11	NORMAL	NONE	LOW
INUG	INUG-14	19-Aug-11	NORMAL	NONE	LOW
INUG	INUG-14	19-Aug-11			
INUG	INUG-14	19-Aug-11			
INUG	INUG-14	19-Aug-11	NORMAL	NONE	HIGH
INUG	INUG-14	19-Aug-11	NORMAL	NONE	LOW
INUG	INUG-14	19-Aug-11	NORMAL	NONE	LOW
INUG	INUG-14	19-Aug-11			
INUG	INUG-14	19-Aug-11	NORMAL	NONE	LOW
INUG	INUG-14	19-Aug-11	NORMAL	NONE	LOW
INUG	INUG-01	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-01	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-01	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-01	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-01	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-01	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-01	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-01	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-01	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-01	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-02	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-02	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-02	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-03	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-05	17-Aug-11	NORMAL	NONE	NONE
INUG	INUG-05	17-Aug-11			
INUG	INUG-08	18-Aug-11	NORMAL	NONE	NONE
INUG	INUG-08	18-Aug-11	NORMAL	NONE	NONE
INUG	INUG-08	18-Aug-11	NORMAL	NONE	NONE
INUG	INUG-08	18-Aug-11	NORMAL	NONE	NONE
INUG	INUG-08	18-Aug-11	NORMAL	NONE	NONE
INUG	INUG-08	18-Aug-11	NORMAL	NONE	NONE
INUG	INUG-08	18-Aug-11	NORMAL	NONE	NONE
INUG	INUG-08	18-Aug-11	NORMAL	NONE	NONE
INUG	INUG-08	18-Aug-11	NORMAL	NONE	NONE
INUG	INUG-09	18-Aug-11	NORMAL	NONE	NONE
INUG	INUG-13	19-Aug-11	NORMAL	NONE	NONE
INUG	INUG-13	19-Aug-11	NORMAL	NONE	NONE
INUG	INUG-13	19-Aug-11	NORMAL	NONE	NONE
INUG	INUG-13	19-Aug-11	NORMAL	NONE	NONE
INUG	INUG-13	19-Aug-11	NORMAL	NONE	NONE
INUG	INUG-13	19-Aug-11	NORMAL	NONE	NONE
INUG	INUG-13	19-Aug-11	NORMAL	NONE	LOW
INUG	INUG-14	19-Aug-11	NORMAL	NONE	LOW
INUG	INUG-14	19-Aug-11	NORMAL	NONE	NONE
INUG	INUG-14	19-Aug-11	NORMAL	NONE	NONE
INUG	INUG-14	19-Aug-11	NORMAL	NONE	NONE
INUG	INUG-14	19-Aug-11	NORMAL	NONE	NONE

Lake	Set ID	Date	Stomach contents
INUG	INUG-02	17-Aug-11	40% ZOOPLANKTON
INUG	INUG-07	18-Aug-11	
INUG	INUG-07	18-Aug-11	20% ZOOPLANKTON
INUG	INUG-08	18-Aug-11	
INUG	INUG-14	19-Aug-11	
INUG	INUG-01	17-Aug-11	
INUG	INUG-01	17-Aug-11	
INUG	INUG-01	17-Aug-11	
INUG	INUG-01	17-Aug-11	
INUG	INUG-01	17-Aug-11	
INUG	INUG-01	17-Aug-11	
INUG	INUG-01	17-Aug-11	MT
INUG	INUG-01	17-Aug-11	MT
INUG	INUG-01	17-Aug-11	80% CLAM SHRIMP
INUG	INUG-01	17-Aug-11	MT
INUG	INUG-01	17-Aug-11	70% CLAM SHRIMP
INUG	INUG-02	17-Aug-11	
INUG	INUG-02	17-Aug-11	MT
INUG	INUG-02	17-Aug-11	50% FISH REMAINS, CLAM SHRIMP
INUG	INUG-02	17-Aug-11	
INUG	INUG-02	17-Aug-11	
INUG	INUG-02	17-Aug-11	
INUG	INUG-02	17-Aug-11	
INUG	INUG-02	17-Aug-11	
INUG	INUG-02	17-Aug-11	10% SALVELINUS REMAINS
INUG	INUG-02	17-Aug-11	90% LKTR & OTHER UNID FISH
INUG	INUG-02	17-Aug-11	40% FISH REMAINS
INUG	INUG-02	17-Aug-11	100% CLAM SHRIMP, FISH REMAINS, EPHEMEROPTERA, OSTRACODS
INUG	INUG-02	17-Aug-11	
INUG	INUG-02	17-Aug-11	
INUG	INUG-02	17-Aug-11	70% CHIRONOMIDS, SCULPIN
INUG	INUG-04	17-Aug-11	
INUG	INUG-04	17-Aug-11	
INUG	INUG-04	17-Aug-11	20% FISH REMAINS
INUG	INUG-05	17-Aug-11	
INUG	INUG-05	17-Aug-11	
INUG	INUG-05	17-Aug-11	MT
INUG	INUG-05	17-Aug-11	30% FISH REMAINS, ADULT HYMENOPTERA
INUG	INUG-05	17-Aug-11	80% FISH REMAINS
INUG	INUG-05	17-Aug-11	20% CLAM SHRIMP
INUG	INUG-06	17-Aug-11	
INUG	INUG-07	18-Aug-11	50% CLAM SHRIMP
INUG	INUG-07	18-Aug-11	100% MAMMAL (VOLE?)
INUG	INUG-07	18-Aug-11	
INUG	INUG-07	18-Aug-11	
INUG	INUG-07	18-Aug-11	90% CLAM SHRIMP
INUG	INUG-07	18-Aug-11	
INUG	INUG-07	18-Aug-11	
INUG	INUG-07	18-Aug-11	MT
INUG	INUG-07	18-Aug-11	MT
INUG	INUG-07	18-Aug-11	
INUG	INUG-07	18-Aug-11	MT
INUG	INUG-07	18-Aug-11	
INUG	INUG-08	18-Aug-11	
INUG	INUG-08	18-Aug-11	
INUG	INUG-08	18-Aug-11	
INUG	INUG-08	18-Aug-11	MT
INUG	INUG-08	18-Aug-11	
INUG	INUG-08	18-Aug-11	30% FISH REMAINS
INUG	INUG-08	18-Aug-11	50% CLAM SHRIMP, OSTRACODS
INUG	INUG-08	18-Aug-11	40% LKTR
INUG	INUG-08	18-Aug-11	30% CLAM SHRIMP
INUG	INUG-08	18-Aug-11	MT
INUG	INUG-09	18-Aug-11	
INUG	INUG-09	18-Aug-11	
INUG	INUG-09	18-Aug-11	
INUG	INUG-09	18-Aug-11	
INUG	INUG-10	18-Aug-11	

Lake	Set ID	Date	Stomach contents
INUG	INUG-10	18-Aug-11	20% CLAM SHRIMP
INUG	INUG-10	18-Aug-11	MT
INUG	INUG-13	19-Aug-11	
INUG	INUG-13	19-Aug-11	
INUG	INUG-13	19-Aug-11	MT
INUG	INUG-13	19-Aug-11	30% CLAM SHRIMP
INUG	INUG-13	19-Aug-11	
INUG	INUG-13	19-Aug-11	30% CLAM SHRIMP
INUG	INUG-13	19-Aug-11	
INUG	INUG-13	19-Aug-11	
INUG	INUG-13	19-Aug-11	
INUG	INUG-13	19-Aug-11	40% CLAM SHRIMP
INUG	INUG-14	19-Aug-11	20% FISH REMAINS, CESTODES, ZOOPLANKTON
INUG	INUG-14	19-Aug-11	20% FISH REMAINS (SCULPIN)
INUG	INUG-14	19-Aug-11	10% FISH REMAINS
INUG	INUG-14	19-Aug-11	
INUG	INUG-14	19-Aug-11	
INUG	INUG-14	19-Aug-11	MT
INUG	INUG-14	19-Aug-11	50% CLAM SHRIMP
INUG	INUG-14	19-Aug-11	60% CLAM SHRIMP
INUG	INUG-14	19-Aug-11	
INUG	INUG-14	19-Aug-11	40% BURB REMAINS
INUG	INUG-14	19-Aug-11	50% CLAM SHRIMP
INUG	INUG-01	17-Aug-11	70% CLAM SHRIMP, ZOOPLANKTON, CHIRONOMIDS, OSTRACODS
INUG	INUG-01	17-Aug-11	100% CLAM SHRIMP, CHIRONOMID, ZOOPLANKTON, OSTRACODS
INUG	INUG-01	17-Aug-11	80% CLAM SHRIMP, CHIRONOMID, ZOOPLANKTON, OSTRACODS
INUG	INUG-01	17-Aug-11	100% CLAM SHRIMP, CHIRONOMID, ZOOPLANKTON, OSTRACODS
INUG	INUG-01	17-Aug-11	60% CLAM SHRIMP, CHIRONOMID, ZOOPLANKTON, OSTRACODS
INUG	INUG-01	17-Aug-11	MT
INUG	INUG-01	17-Aug-11	10% CLAM SHRIMP
INUG	INUG-01	17-Aug-11	50% CLAM SHRIMP, OSTRACODS
INUG	INUG-01	17-Aug-11	MT
INUG	INUG-01	17-Aug-11	50% CLAM SHRIMP
INUG	INUG-02	17-Aug-11	80% CLAM SHRIMP
INUG	INUG-02	17-Aug-11	80% CLAMS, CHIRONOMIDS, CLAM SHRIMP
INUG	INUG-02	17-Aug-11	100% CLAM SHRIMP, CHIRONOMID, CLAMS
INUG	INUG-03	17-Aug-11	30% CLADOCERA, CLAM SHRIMP
INUG	INUG-05	17-Aug-11	50% CLAM SHRIMP
INUG	INUG-05	17-Aug-11	
INUG	INUG-08	18-Aug-11	40% CLAM SHRIMP, CHIRONOMIDS
INUG	INUG-08	18-Aug-11	60% CLAM SHRIMP, OSTRACODS, CHIRONOMIDS
INUG	INUG-08	18-Aug-11	60% CLAM SHRIMP, OSTRACODS, CHIRONOMIDS
INUG	INUG-08	18-Aug-11	30% CLAM SHRIMP, OSTRACODS, CHIRONOMIDS
INUG	INUG-08	18-Aug-11	30% CLAM SHRIMP, OSTRACODS, CHIRONOMIDS
INUG	INUG-08	18-Aug-11	10% CLAM SHRIMP
INUG	INUG-08	18-Aug-11	10% CHIRONOMIDS
INUG	INUG-08	18-Aug-11	20% CHIRONOMIDS, CLAM SHRIMP
INUG	INUG-08	18-Aug-11	10% CLAM SHRIMP
INUG	INUG-09	18-Aug-11	30% CLAM SHRIMP, OSTRACODS, CHIRONOMIDS
INUG	INUG-13	19-Aug-11	20% CLAM SHRIMP, ZOOPLANKTON, OTHER INVERTS
INUG	INUG-13	19-Aug-11	10% TRICHOPTERANS, CLAM SHRIMP
INUG	INUG-13	19-Aug-11	MT
INUG	INUG-13	19-Aug-11	20% CLAM SHRIMP, OSTRACODS
INUG	INUG-13	19-Aug-11	MT
INUG	INUG-13	19-Aug-11	20% CLAM SHRIMP, OSTRACODS
INUG	INUG-13	19-Aug-11	40% CLAM SHRIMP, ZOOPLANKTON
INUG	INUG-14	19-Aug-11	30% CLAM SHRIMP, OSTRACODS
INUG	INUG-14	19-Aug-11	20% TRICHOPTERA, ZOOPLANKTON
INUG	INUG-14	19-Aug-11	30% CLAM SHRIMP, OSTRACODS
INUG	INUG-14	19-Aug-11	40% CLAM SHRIMP, OSTRACODS, TRICHOPTERA
INUG	INUG-14	19-Aug-11	70% CHIRONOMIDS
INUG	INUG-14	19-Aug-11	10% CLAM SHRIMP, CHIRONOMIDS

Lake	Set ID	Date	Mesh Size (mm)	Day/Night	Species	Sample ID	Mortality
PDL	PD-01	17-Aug-11	38	NIGHT	ARCH	PD-01.14	Y
PDL	PD-01	17-Aug-11	38	NIGHT	ARCH	PD-01.15	Y
PDL	PD-01	17-Aug-11	38	NIGHT	ARCH	PD-01.18	N
PDL	PD-01	17-Aug-11	25	NIGHT	ARCH	PD-01.20	N
PDL	PD-03B	18-Aug-11	51	NIGHT	ARCH	PD-03.06	Y
PDL	PD-03B	18-Aug-11	51	NIGHT	ARCH	PD-03.12	Y
PDL	PD-04B	18-Aug-11	25	NIGHT	ARCH	PD-04.16	N
PDL	PD-08B	19-Aug-11	102	NIGHT	ARCH	PD-08.01	Y
PDL	PD-01	17-Aug-11	126	NIGHT	LKTR	PD-01.01	N
PDL	PD-01	17-Aug-11	102	NIGHT	LKTR	PD-01.02	N
PDL	PD-01	17-Aug-11	76	NIGHT	LKTR	PD-01.05	N
PDL	PD-01	17-Aug-11	76	NIGHT	LKTR	PD-01.06	Y
PDL	PD-01	17-Aug-11	76	NIGHT	LKTR	PD-01.09	Y
PDL	PD-01	17-Aug-11	76	NIGHT	LKTR	PD-01.10	Y
PDL	PD-01	17-Aug-11	76	NIGHT	LKTR	PD-01.11	Y
PDL	PD-01	17-Aug-11	38	NIGHT	LKTR	PD-01.12	N
PDL	PD-01	17-Aug-11	38	NIGHT	LKTR	PD-01.13	N
PDL	PD-01	17-Aug-11	38	NIGHT	LKTR	PD-01.16	Y
PDL	PD-01	17-Aug-11	38	NIGHT	LKTR	PD-01.17	Y
PDL	PD-01	17-Aug-11	25	NIGHT	LKTR	PD-01.19	N
PDL	PD-01	17-Aug-11	25	NIGHT	LKTR	PD-01.21	N
PDL	PD-02	17-Aug-11	51	NIGHT	LKTR	PD-02.01	Y
PDL	PD-02	17-Aug-11	51	NIGHT	LKTR	PD-02.02	Y
PDL	PD-03A	17-Aug-11	51	DAY	LKTR	PD-03.01	N
PDL	PD-03B	18-Aug-11	51	NIGHT	LKTR	PD-03.02	Y
PDL	PD-03B	18-Aug-11	51	NIGHT	LKTR	PD-03.03	Y
PDL	PD-03B	18-Aug-11	51	NIGHT	LKTR	PD-03.04	Y
PDL	PD-03B	18-Aug-11	51	NIGHT	LKTR	PD-03.05	Y
PDL	PD-03B	18-Aug-11	76	NIGHT	LKTR	PD-03.09	Y
PDL	PD-03B	18-Aug-11	51	NIGHT	LKTR	PD-03.10	Y
PDL	PD-03B	18-Aug-11	51	NIGHT	LKTR	PD-03.11	N
PDL	PD-03B	18-Aug-11	51	NIGHT	LKTR	PD-03.13	Y
PDL	PD-03B	18-Aug-11	51	NIGHT	LKTR	PD-03.14	Y
PDL	PD-03B	18-Aug-11	76	NIGHT	LKTR	PD-03.15	Y
PDL	PD-03B	18-Aug-11	76	NIGHT	LKTR	PD-03.16	Y
PDL	PD-03B	18-Aug-11	76	NIGHT	LKTR	PD-03.17	Y
PDL	PD-03B	18-Aug-11	76	NIGHT	LKTR	PD-03.19	N
PDL	PD-03B	18-Aug-11	76	NIGHT	LKTR	PD-03.20	Y
PDL	PD-04B	18-Aug-11	126	NIGHT	LKTR	PD-04.01	N
PDL	PD-04B	18-Aug-11	102	NIGHT	LKTR	PD-04.02	N
PDL	PD-04B	18-Aug-11	102	NIGHT	LKTR	PD-04.03	N
PDL	PD-04B	18-Aug-11	102	NIGHT	LKTR	PD-04.04	Y
PDL	PD-04B	18-Aug-11	76	NIGHT	LKTR	PD-04.05	Y
PDL	PD-04B	18-Aug-11	76	NIGHT	LKTR	PD-04.06	Y
PDL	PD-04B	18-Aug-11	76	NIGHT	LKTR	PD-04.07	N
PDL	PD-04B	18-Aug-11	51	NIGHT	LKTR	PD-04.08	Y
PDL	PD-04B	18-Aug-11	51	NIGHT	LKTR	PD-04.09	Y
PDL	PD-04B	18-Aug-11	51	NIGHT	LKTR	PD-04.10	N
PDL	PD-04B	18-Aug-11	38	NIGHT	LKTR	PD-04.11	Y
PDL	PD-04B	18-Aug-11	38	NIGHT	LKTR	PD-04.12	Y
PDL	PD-04B	18-Aug-11	38	NIGHT	LKTR	PD-04.13	Y
PDL	PD-04B	18-Aug-11	25	NIGHT	LKTR	PD-04.14	Y
PDL	PD-04B	18-Aug-11	25	NIGHT	LKTR	PD-04.15	N
PDL	PD-05A	18-Aug-11	76	DAY	LKTR	PD-05.01	N
PDL	PD-06A	18-Aug-11	126	DAY	LKTR	PD-06.01	N
PDL	PD-06A	18-Aug-11	102	DAY	LKTR	PD-06.02	N
PDL	PD-07B	19-Aug-11	76	NIGHT	LKTR	PD-07.01	N
PDL	PD-07B	19-Aug-11	76	NIGHT	LKTR	PD-07.02	N
PDL	PD-07B	19-Aug-11	51	NIGHT	LKTR	PD-07.04	N
PDL	PD-08A	18-Aug-11	102	DAY	LKTR	PD-08.01A	Y
PDL	PD-08B	19-Aug-11	76	NIGHT	LKTR	PD-08.02	N
PDL	PD-08B	19-Aug-11	76	NIGHT	LKTR	PD-08.03	N
PDL	PD-08B	19-Aug-11	51	NIGHT	LKTR	PD-08.04	N
PDL	PD-08B	19-Aug-11	38	NIGHT	LKTR	PD-08.05	Y
PDL	PD-08B	19-Aug-11	38	NIGHT	LKTR	PD-08.06	Y
PDL	PD-08B	19-Aug-11	38	NIGHT	LKTR	PD-08.07	N
PDL	PD-01	17-Aug-11	126	NIGHT	RNWH	PD-01.03	Y
PDL	PD-01	17-Aug-11	102	NIGHT	RNWH	PD-01.04	Y
PDL	PD-01	17-Aug-11	76	NIGHT	RNWH	PD-01.07	Y
PDL	PD-01	17-Aug-11	76	NIGHT	RNWH	PD-01.08	Y
PDL	PD-03B	18-Aug-11	51	NIGHT	RNWH	PD-03.08	Y
PDL	PD-03B	18-Aug-11	76	NIGHT	RNWH	PD-03.18	Y
PDL	PD-07B	19-Aug-11	51	NIGHT	RNWH	PD-07.03	Y

Lake	Set ID	Date	Length.mm	Weight.g	K	Weighed with 25 g scale	Sex
PDL	PD-01	17-Aug-11	243	152	1.059314135	N	M
PDL	PD-01	17-Aug-11	255	172	1.037308426	N	F
PDL	PD-01	17-Aug-11	205	100	1.160749264	N (± 10)	
PDL	PD-01	17-Aug-11	154	48	1.314253296	N (± 1)	
PDL	PD-03B	18-Aug-11	246	161	1.081485136	N	M
PDL	PD-03B	18-Aug-11	272	213	1.0584562	N	U
PDL	PD-04B	18-Aug-11	114	18	1.214948729	N (± 1)	
PDL	PD-08B	19-Aug-11	255	174	1.049370152	N	U
PDL	PD-01	17-Aug-11	621	2600	1.085671775	Y	
PDL	PD-01	17-Aug-11	869	7600	1.15812187	Y	
PDL	PD-01	17-Aug-11	820	6000	1.088202435	Y	
PDL	PD-01	17-Aug-11	344	396	0.972791704	N	M
PDL	PD-01	17-Aug-11	282	220	0.98101434	N	F
PDL	PD-01	17-Aug-11	283	228	1.005948021	N	F
PDL	PD-01	17-Aug-11	278	212	0.986737136	N	M
PDL	PD-01	17-Aug-11	804	6100	1.173712389	Y	
PDL	PD-01	17-Aug-11	811	4450	0.834252369	Y	
PDL	PD-01	17-Aug-11	212	108	1.133486032	N	F
PDL	PD-01	17-Aug-11	250	146	0.9344	N	F
PDL	PD-01	17-Aug-11	135	36	1.463191587	N (± 1)	
PDL	PD-01	17-Aug-11	142	42	1.46684511	N (± 1)	
PDL	PD-02	17-Aug-11	295	272	1.059504623	N	M
PDL	PD-02	17-Aug-11	268	184	0.955902156	N	M
PDL	PD-03A	17-Aug-11	403	640	0.977833328	Y	
PDL	PD-03B	18-Aug-11	280	211	0.961188047	N	M
PDL	PD-03B	18-Aug-11	281	247	1.113212293	N	M
PDL	PD-03B	18-Aug-11	482	1090	0.973386645	N	F
PDL	PD-03B	18-Aug-11	496	1418	1.162067088	N	F
PDL	PD-03B	18-Aug-11	522	1316	0.925218654	N	F
PDL	PD-03B	18-Aug-11	710	4100	1.145536181	Y	
PDL	PD-03B	18-Aug-11	269	236	1.212425782	N (± 10)	
PDL	PD-03B	18-Aug-11	253	150	0.926253145	N	F
PDL	PD-03B	18-Aug-11	220	120	1.126972201	N	U
PDL	PD-03B	18-Aug-11	577	2126	1.106715062	N	F
PDL	PD-03B	18-Aug-11	534	1518	0.996891747	N	F
PDL	PD-03B	18-Aug-11	565	1740	0.964725826	N	M
PDL	PD-03B	18-Aug-11	189	66	0.97759399	N (± 10)	
PDL	PD-03B	18-Aug-11	508	1470	1.121311298	N	M
PDL	PD-04B	18-Aug-11	304	236	0.840023145	Y	
PDL	PD-04B	18-Aug-11	534	1600	1.050742289	Y	
PDL	PD-04B	18-Aug-11	818	6300	1.151014086	Y	
PDL	PD-04B	18-Aug-11	584	2188	1.098522044	N	M
PDL	PD-04B	18-Aug-11	521	1598	1.129961392	N	M
PDL	PD-04B	18-Aug-11	566	1966	1.084261956	N	F
PDL	PD-04B	18-Aug-11	532	1700	1.129052208	Y	
PDL	PD-04B	18-Aug-11	638	2552	0.982694746	N	M
PDL	PD-04B	18-Aug-11	390	554	0.933933478	N	F
PDL	PD-04B	18-Aug-11	697	3900	1.151771354	Y	
PDL	PD-04B	18-Aug-11	184	61	0.979211186	N	F
PDL	PD-04B	18-Aug-11	203	80	0.956316994	N	M
PDL	PD-04B	18-Aug-11	193	78	1.084982356	N	F
PDL	PD-04B	18-Aug-11	136	23	0.91434714	N	F
PDL	PD-04B	18-Aug-11	132	30	1.304365974	N (± 1)	
PDL	PD-05A	18-Aug-11	620	2175	0.912607835	Y	
PDL	PD-06A	18-Aug-11	584	1850	0.928823483	Y	
PDL	PD-06A	18-Aug-11	548	1675	1.017825052	Y	
PDL	PD-07B	19-Aug-11	251	150	0.948571549	N (± 10)	
PDL	PD-07B	19-Aug-11	237	140	1.051678505	N (± 10)	
PDL	PD-07B	19-Aug-11	151	55	1.597466824	N (± 1)	
PDL	PD-08A	18-Aug-11	629	2746	1.103439678	N	F
PDL	PD-08B	19-Aug-11	559	1675	0.9589134	Y	
PDL	PD-08B	19-Aug-11	686	3400	1.053189619	Y	
PDL	PD-08B	19-Aug-11	282	225	1.003310121	N (± 10)	
PDL	PD-08B	19-Aug-11	235	136	1.047937355	N	M
PDL	PD-08B	19-Aug-11	202	80	0.970590148	N	F
PDL	PD-08B	19-Aug-11	168	52	1.096668826	N (± 1)	
PDL	PD-01	17-Aug-11	390	748	1.260978776	N	F
PDL	PD-01	17-Aug-11	386	742	1.290155301	N	F
PDL	PD-01	17-Aug-11	372	558	1.08394034	N	F
PDL	PD-01	17-Aug-11	247	158	1.04849467	N	M
PDL	PD-03B	18-Aug-11	377	564	1.052579854	N	M
PDL	PD-03B	18-Aug-11	412	838	1.19826361	N	M
PDL	PD-07B	19-Aug-11	322	388	1.162155148	N	F

Lake	Set ID	Date	Maturity	Repro Status	Age Structure	Age.OTO.yr	Age.PEC.yr
PDL	PD-01	17-Aug-11	IM	UD	PEC/OTO	5	
PDL	PD-01	17-Aug-11	IM	UD	PEC/OTO	10	
PDL	PD-01	17-Aug-11					
PDL	PD-01	17-Aug-11					
PDL	PD-03B	18-Aug-11	IM	UD	PEC/OTO	6	
PDL	PD-03B	18-Aug-11	IM	UD	PEC/OTO	11	
PDL	PD-04B	18-Aug-11					
PDL	PD-08B	19-Aug-11	IM	UD	PEC/OTO	6	
PDL	PD-01	17-Aug-11			PEC		20
PDL	PD-01	17-Aug-11			PEC		26
PDL	PD-01	17-Aug-11			PEC		21
PDL	PD-01	17-Aug-11	IM	UD	PEC/OTO	15	10
PDL	PD-01	17-Aug-11	IM	UD	PEC/OTO	10	7
PDL	PD-01	17-Aug-11	IM	UD	PEC/OTO	12	8
PDL	PD-01	17-Aug-11	IM	UD	PEC/OTO	10	8
PDL	PD-01	17-Aug-11			PEC		30
PDL	PD-01	17-Aug-11			PEC		ua
PDL	PD-01	17-Aug-11	IM	UD	PEC/OTO	11	5
PDL	PD-01	17-Aug-11	IM	UD	PEC/OTO	12	5
PDL	PD-01	17-Aug-11					
PDL	PD-02	17-Aug-11	IM	UD	PEC/OTO	10	8
PDL	PD-02	17-Aug-11	IM	UD	PEC/OTO	8	7
PDL	PD-03A	17-Aug-11			PEC		14
PDL	PD-03B	18-Aug-11	IM	UD	PEC/OTO	8	8
PDL	PD-03B	18-Aug-11	IM	UD	PEC/OTO	8	5
PDL	PD-03B	18-Aug-11	MA	RS	PEC/OTO	15	12
PDL	PD-03B	18-Aug-11	MA	RI	PEC/OTO	24	13
PDL	PD-03B	18-Aug-11	MA	RS	PEC/OTO	15	10
PDL	PD-03B	18-Aug-11			PEC		18
PDL	PD-03B	18-Aug-11			PEC		5
PDL	PD-03B	18-Aug-11	IM	UD	PEC/OTO	10	6
PDL	PD-03B	18-Aug-11	IM	UD	PEC/OTO	15	4
PDL	PD-03B	18-Aug-11	MA	RI	PEC/OTO	22	12
PDL	PD-03B	18-Aug-11	MA	RS	PEC/OTO	18	11
PDL	PD-03B	18-Aug-11	MA	RI	PEC/OTO	24	14
PDL	PD-03B	18-Aug-11			PEC		ns
PDL	PD-03B	18-Aug-11	MA	RI	PEC/OTO	14	10
PDL	PD-04B	18-Aug-11			PEC		9
PDL	PD-04B	18-Aug-11			PEC		20
PDL	PD-04B	18-Aug-11			PEC		28
PDL	PD-04B	18-Aug-11	MA	RS	PEC/OTO	29	16
PDL	PD-04B	18-Aug-11	MA	RI	PEC/OTO	23	10
PDL	PD-04B	18-Aug-11	MA	RI	PEC/OTO	20	12
PDL	PD-04B	18-Aug-11			PEC		17
PDL	PD-04B	18-Aug-11	IM	UD	PEC/OTO	16	6
PDL	PD-04B	18-Aug-11	IM	UD	PEC/OTO	18	10
PDL	PD-04B	18-Aug-11			PEC		24
PDL	PD-04B	18-Aug-11	IM	UD	PEC/OTO	10	4
PDL	PD-04B	18-Aug-11	IM	UD	PEC/OTO	6	4
PDL	PD-04B	18-Aug-11	IM	UD	PEC/OTO	10	4
PDL	PD-04B	18-Aug-11	IM	UD	PEC/OTO	4	3
PDL	PD-04B	18-Aug-11					
PDL	PD-05A	18-Aug-11			PEC		22
PDL	PD-06A	18-Aug-11			PEC		16
PDL	PD-06A	18-Aug-11			PEC		17
PDL	PD-07B	19-Aug-11			PEC		4
PDL	PD-07B	19-Aug-11			PEC		3
PDL	PD-07B	19-Aug-11			PEC		4
PDL	PD-08A	18-Aug-11	MA	RI	PEC/OTO	20	18
PDL	PD-08B	19-Aug-11			PEC		20
PDL	PD-08B	19-Aug-11			PEC		22
PDL	PD-08B	19-Aug-11			PEC		6
PDL	PD-08B	19-Aug-11	IM	UD	PEC/OTO	10	4
PDL	PD-08B	19-Aug-11	IM	UD	PEC/OTO	10	4
PDL	PD-08B	19-Aug-11			PEC		2
PDL	PD-01	17-Aug-11	MA	RI	PEC/OTO	12	12
PDL	PD-01	17-Aug-11	MA	RI	PEC/OTO	22	14
PDL	PD-01	17-Aug-11	MA	RI	PEC/OTO	14	8
PDL	PD-01	17-Aug-11	IM	UD	PEC/OTO	12	5
PDL	PD-03B	18-Aug-11	MA	RI	PEC/OTO	16	14
PDL	PD-03B	18-Aug-11	MA	RI	PEC/OTO	16	12
PDL	PD-07B	19-Aug-11	MA	RI	PEC/OTO	9	6

Lake	Set ID	Date	Ext. DELT	Ext. Parasites	Tagged?	Tag ID
PDL	PD-01	17-Aug-11	NONE	NONE	N	
PDL	PD-01	17-Aug-11	NONE	NONE	N	
PDL	PD-01	17-Aug-11	NONE	NONE	N	
PDL	PD-01	17-Aug-11	TORN LEFT OPERCULUM	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-04B	18-Aug-11	NONE	NONE	N	
PDL	PD-08B	19-Aug-11	NONE	NONE	N	
PDL	PD-01	17-Aug-11	NONE	NONE	Y	77152
PDL	PD-01	17-Aug-11	DORSAL FIN DEFORM	NONE	Y	77153
PDL	PD-01	17-Aug-11	SMALL SCRATCH DORSAL	NONE	Y	77154
PDL	PD-01	17-Aug-11	NONE	NONE	N	
PDL	PD-01	17-Aug-11	NONE	NONE	N	
PDL	PD-01	17-Aug-11	NONE	NONE	N	
PDL	PD-01	17-Aug-11	RED SPOTS BW PECS - DORSAL DAMAGE	NONE	Y	77155
PDL	PD-01	17-Aug-11	UNUSUALLY SKINNY - FLACID BELLY	NONE	Y	77156
PDL	PD-01	17-Aug-11	NONE	NONE	N	
PDL	PD-01	17-Aug-11	NONE	NONE	N	
PDL	PD-01	17-Aug-11	NONE	NONE	N	
PDL	PD-01	17-Aug-11	NONE	NONE	N	
PDL	PD-02	17-Aug-11	NONE	NONE	N	
PDL	PD-02	17-Aug-11	NONE	NONE	N	
PDL	PD-03A	17-Aug-11	NONE	NONE	Y	77157
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	Y	77158
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-03B	18-Aug-11	NONE	NONE	N	
PDL	PD-0					

Lake	Set ID	Date	RECAP?	Gonad weight (g)	Gonad appearance
PDL	PD-01	17-Aug-11	N		NORMAL
PDL	PD-01	17-Aug-11	N		NORMAL
PDL	PD-01	17-Aug-11	N		
PDL	PD-01	17-Aug-11	N		
PDL	PD-03B	18-Aug-11	N		NORMAL
PDL	PD-03B	18-Aug-11	N		NORMAL
PDL	PD-04B	18-Aug-11	N		
PDL	PD-08B	19-Aug-11	N		NORMAL
PDL	PD-01	17-Aug-11	N		
PDL	PD-01	17-Aug-11	N		
PDL	PD-01	17-Aug-11	N		NORMAL
PDL	PD-01	17-Aug-11	N		NORMAL
PDL	PD-01	17-Aug-11	N		NORMAL
PDL	PD-01	17-Aug-11	N		NORMAL
PDL	PD-01	17-Aug-11	N		
PDL	PD-01	17-Aug-11	N		
PDL	PD-01	17-Aug-11	N		NORMAL
PDL	PD-01	17-Aug-11	N		NORMAL
PDL	PD-01	17-Aug-11	N		
PDL	PD-01	17-Aug-11	N		
PDL	PD-02	17-Aug-11	N		NORMAL
PDL	PD-02	17-Aug-11	N		NORMAL
PDL	PD-03A	17-Aug-11	N		
PDL	PD-03B	18-Aug-11	N		NORMAL
PDL	PD-03B	18-Aug-11	N		NORMAL
PDL	PD-03B	18-Aug-11	N		NORMAL
PDL	PD-03B	18-Aug-11	N	71.31	NORMAL
PDL	PD-03B	18-Aug-11	N		NORMAL
PDL	PD-03B	18-Aug-11	N		
PDL	PD-03B	18-Aug-11	N		
PDL	PD-03B	18-Aug-11	N		NORMAL
PDL	PD-03B	18-Aug-11	N		NORMAL
PDL	PD-03B	18-Aug-11	N	230	NORMAL
PDL	PD-03B	18-Aug-11	N		NORMAL
PDL	PD-03B	18-Aug-11	N	76.63	NORMAL
PDL	PD-03B	18-Aug-11	N		
PDL	PD-03B	18-Aug-11	N	62.71	NORMAL
PDL	PD-04B	18-Aug-11	N		
PDL	PD-04B	18-Aug-11	N		
PDL	PD-04B	18-Aug-11	N		
PDL	PD-04B	18-Aug-11	N		NORMAL
PDL	PD-04B	18-Aug-11	N	75.14	NORMAL
PDL	PD-04B	18-Aug-11	N	236	NORMAL
PDL	PD-04B	18-Aug-11	N		
PDL	PD-04B	18-Aug-11	N		NORMAL
PDL	PD-04B	18-Aug-11	N		NORMAL
PDL	PD-04B	18-Aug-11	N		
PDL	PD-04B	18-Aug-11	N		NORMAL
PDL	PD-04B	18-Aug-11	N		NORMAL
PDL	PD-04B	18-Aug-11	N		NORMAL
PDL	PD-04B	18-Aug-11	N		
PDL	PD-05A	18-Aug-11	N		
PDL	PD-06A	18-Aug-11	N		
PDL	PD-06A	18-Aug-11	N		
PDL	PD-07B	19-Aug-11	N		
PDL	PD-07B	19-Aug-11	N		
PDL	PD-07B	19-Aug-11	N		
PDL	PD-08A	18-Aug-11	N	390	NORMAL
PDL	PD-08B	19-Aug-11	N		
PDL	PD-08B	19-Aug-11	N		
PDL	PD-08B	19-Aug-11	N		
PDL	PD-08B	19-Aug-11	N		NORMAL
PDL	PD-08B	19-Aug-11	N		NORMAL
PDL	PD-08B	19-Aug-11	N		
PDL	PD-01	17-Aug-11	N	28.77	NORMAL
PDL	PD-01	17-Aug-11	N	29.98	NORMAL
PDL	PD-01	17-Aug-11	N	17.76	NORMAL
PDL	PD-01	17-Aug-11	N		NORMAL
PDL	PD-03B	18-Aug-11	N	24.87	NORMAL
PDL	PD-03B	18-Aug-11	N	48.78	NORMAL
PDL	PD-07B	19-Aug-11	N	10.27	NORMAL

Lake	Set ID	Date	Weight of 400 eggs (g)	Liver weight (g)
PDL	PD-01	17-Aug-11		1.89
PDL	PD-01	17-Aug-11		2.15
PDL	PD-01	17-Aug-11		
PDL	PD-01	17-Aug-11		
PDL	PD-03B	18-Aug-11		1.79
PDL	PD-03B	18-Aug-11		2.22
PDL	PD-04B	18-Aug-11		
PDL	PD-08B	19-Aug-11		2.02
PDL	PD-01	17-Aug-11		
PDL	PD-01	17-Aug-11		
PDL	PD-01	17-Aug-11		4.52
PDL	PD-01	17-Aug-11		1.7
PDL	PD-01	17-Aug-11		1.93
PDL	PD-01	17-Aug-11		1.79
PDL	PD-01	17-Aug-11		
PDL	PD-01	17-Aug-11		
PDL	PD-01	17-Aug-11		0.93
PDL	PD-01	17-Aug-11		1.14
PDL	PD-01	17-Aug-11		
PDL	PD-01	17-Aug-11		
PDL	PD-02	17-Aug-11		2.79
PDL	PD-02	17-Aug-11		1.62
PDL	PD-03A	17-Aug-11		
PDL	PD-03B	18-Aug-11		1.79
PDL	PD-03B	18-Aug-11		2.33
PDL	PD-03B	18-Aug-11		9.91
PDL	PD-03B	18-Aug-11	11.26	28.85
PDL	PD-03B	18-Aug-11		14.21
PDL	PD-03B	18-Aug-11		
PDL	PD-03B	18-Aug-11		
PDL	PD-03B	18-Aug-11		1.13
PDL	PD-03B	18-Aug-11		1.05
PDL	PD-03B	18-Aug-11	27.33	49.74
PDL	PD-03B	18-Aug-11		16.05
PDL	PD-03B	18-Aug-11		15.97
PDL	PD-03B	18-Aug-11		
PDL	PD-03B	18-Aug-11		19.23
PDL	PD-04B	18-Aug-11		
PDL	PD-04B	18-Aug-11		
PDL	PD-04B	18-Aug-11		
PDL	PD-04B	18-Aug-11		20.51
PDL	PD-04B	18-Aug-11		16.11
PDL	PD-04B	18-Aug-11	30.52	45.07
PDL	PD-04B	18-Aug-11		
PDL	PD-04B	18-Aug-11		3.22
PDL	PD-04B	18-Aug-11		5.84
PDL	PD-04B	18-Aug-11		
PDL	PD-04B	18-Aug-11		0.5
PDL	PD-04B	18-Aug-11		0.68
PDL	PD-04B	18-Aug-11		0.83
PDL	PD-04B	18-Aug-11		0.22
PDL	PD-04B	18-Aug-11		
PDL	PD-05A	18-Aug-11		
PDL	PD-06A	18-Aug-11		
PDL	PD-06A	18-Aug-11		
PDL	PD-07B	19-Aug-11		
PDL	PD-07B	19-Aug-11		
PDL	PD-07B	19-Aug-11		
PDL	PD-08A	18-Aug-11	35.19	68.74
PDL	PD-08B	19-Aug-11		
PDL	PD-08B	19-Aug-11		
PDL	PD-08B	19-Aug-11		
PDL	PD-08B	19-Aug-11		1.23
PDL	PD-08B	19-Aug-11		0.72
PDL	PD-08B	19-Aug-11		
PDL	PD-01	17-Aug-11	0.84	11.82
PDL	PD-01	17-Aug-11	0.91	8.39
PDL	PD-01	17-Aug-11	0.76	8.84
PDL	PD-01	17-Aug-11		1.19
PDL	PD-03B	18-Aug-11		4.62
PDL	PD-03B	18-Aug-11		5.93
PDL	PD-07B	19-Aug-11	0.92	4.82

Lake	Set ID	Date	Liver appearance	Int. DELT	Int. Parasites
PDL	PD-01	17-Aug-11	NORMAL	NONE	NONE
PDL	PD-01	17-Aug-11	CYST IN LIVER	NONE	LOW
PDL	PD-01	17-Aug-11			
PDL	PD-01	17-Aug-11			
PDL	PD-03B	18-Aug-11	NORMAL	NONE	LOW
PDL	PD-03B	18-Aug-11	NORMAL	NONE	LOW
PDL	PD-04B	18-Aug-11			
PDL	PD-08B	19-Aug-11	NORMAL	NONE	LOW
PDL	PD-01	17-Aug-11			
PDL	PD-01	17-Aug-11			
PDL	PD-01	17-Aug-11	NORMAL	NONE	LOW
PDL	PD-01	17-Aug-11	NORMAL	NONE	MODERATE (cestodes)
PDL	PD-01	17-Aug-11	CYST IN LIVER	NONE	LOW
PDL	PD-01	17-Aug-11	NORMAL	NONE	LOW
PDL	PD-01	17-Aug-11			
PDL	PD-01	17-Aug-11			
PDL	PD-01	17-Aug-11	NORMAL	NONE	LOW
PDL	PD-01	17-Aug-11	NORMAL	NONE	MODERATE
PDL	PD-01	17-Aug-11			
PDL	PD-01	17-Aug-11			
PDL	PD-02	17-Aug-11	NORMAL	NONE	LOW
PDL	PD-02	17-Aug-11	NORMAL	NONE	LOW
PDL	PD-03A	17-Aug-11			
PDL	PD-03B	18-Aug-11	NORMAL	NONE	LOW
PDL	PD-03B	18-Aug-11	NORMAL	NONE	LOW (cestodes and cysts)
PDL	PD-03B	18-Aug-11	CYST IN LIVER	NONE	LOW
PDL	PD-03B	18-Aug-11	NORMAL	NONE	LOW
PDL	PD-03B	18-Aug-11	NORMAL	NONE	LOW
PDL	PD-03B	18-Aug-11			
PDL	PD-03B	18-Aug-11			
PDL	PD-03B	18-Aug-11	NORMAL	NONE	LOW
PDL	PD-03B	18-Aug-11	NORMAL	NONE	MODERATE
PDL	PD-03B	18-Aug-11	NORMAL	NONE	MODERATE
PDL	PD-03B	18-Aug-11	CYST IN LIVER	NONE	LOW
PDL	PD-03B	18-Aug-11	CYST IN LIVER	NONE	NONE
PDL	PD-03B	18-Aug-11			
PDL	PD-03B	18-Aug-11	NORMAL	NONE	LOW
PDL	PD-04B	18-Aug-11			
PDL	PD-04B	18-Aug-11			
PDL	PD-04B	18-Aug-11			
PDL	PD-04B	18-Aug-11	NORMAL	NONE	LOW
PDL	PD-04B	18-Aug-11	NORMAL	NONE	LOW
PDL	PD-04B	18-Aug-11	NORMAL	NONE	LOW
PDL	PD-04B	18-Aug-11			
PDL	PD-04B	18-Aug-11	NORMAL	NONE	MODERATE
PDL	PD-04B	18-Aug-11	NORMAL	NONE	LOW
PDL	PD-04B	18-Aug-11			
PDL	PD-04B	18-Aug-11	NORMAL	NONE	LOW
PDL	PD-04B	18-Aug-11	CYST ON LIVER	CYST ON LIVER	LOW
PDL	PD-04B	18-Aug-11	NORMAL	NONE	MODERATE
PDL	PD-04B	18-Aug-11	NORMAL	NONE	NONE
PDL	PD-04B	18-Aug-11			
PDL	PD-05A	18-Aug-11			
PDL	PD-06A	18-Aug-11			
PDL	PD-06A	18-Aug-11			
PDL	PD-07B	19-Aug-11			
PDL	PD-07B	19-Aug-11			
PDL	PD-07B	19-Aug-11			
PDL	PD-08A	18-Aug-11	NORMAL	NONE	MODERATE
PDL	PD-08B	19-Aug-11			
PDL	PD-08B	19-Aug-11			
PDL	PD-08B	19-Aug-11			
PDL	PD-08B	19-Aug-11	NORMAL	NONE	LOW
PDL	PD-08B	19-Aug-11	NORMAL	NONE	LOW
PDL	PD-08B	19-Aug-11			
PDL	PD-01	17-Aug-11	NORMAL	NONE	NONE
PDL	PD-01	17-Aug-11	NORMAL	NONE	NONE
PDL	PD-01	17-Aug-11	NORMAL	NONE	NONE
PDL	PD-01	17-Aug-11	NORMAL	NONE	NONE
PDL	PD-03B	18-Aug-11	NORMAL	NONE	NONE
PDL	PD-03B	18-Aug-11	NORMAL	NONE	NONE
PDL	PD-07B	19-Aug-11	CYST IN LIVER	NONE	LOW

Lake	Set ID	Date	Stomach contents
PDL	PD-01	17-Aug-11	80% ZOOPLANKTON
PDL	PD-01	17-Aug-11	70% ZOOPLANKTON
PDL	PD-01	17-Aug-11	
PDL	PD-01	17-Aug-11	
PDL	PD-03B	18-Aug-11	10% ZOOPLANKTON
PDL	PD-03B	18-Aug-11	30% ZOOPLANKTON
PDL	PD-04B	18-Aug-11	
PDL	PD-08B	19-Aug-11	30% ZOOPLANKTON
PDL	PD-01	17-Aug-11	
PDL	PD-01	17-Aug-11	
PDL	PD-01	17-Aug-11	MT
PDL	PD-01	17-Aug-11	5% cestodes
PDL	PD-01	17-Aug-11	20% UNID REMAINS
PDL	PD-01	17-Aug-11	MT
PDL	PD-01	17-Aug-11	
PDL	PD-01	17-Aug-11	
PDL	PD-01	17-Aug-11	90% FISH REMAINS
PDL	PD-01	17-Aug-11	90% CHIRONOMIDS, FISH REMAINS
PDL	PD-01	17-Aug-11	
PDL	PD-01	17-Aug-11	
PDL	PD-02	17-Aug-11	MT
PDL	PD-02	17-Aug-11	MT
PDL	PD-03A	17-Aug-11	
PDL	PD-03B	18-Aug-11	60% FISH REMAINS
PDL	PD-03B	18-Aug-11	50% OSTRACODS, CESTODES
PDL	PD-03B	18-Aug-11	MT
PDL	PD-03B	18-Aug-11	MT
PDL	PD-03B	18-Aug-11	MT
PDL	PD-03B	18-Aug-11	
PDL	PD-03B	18-Aug-11	
PDL	PD-03B	18-Aug-11	MT
PDL	PD-03B	18-Aug-11	10% DIPTERA
PDL	PD-03B	18-Aug-11	MT
PDL	PD-03B	18-Aug-11	5% DIPTERA
PDL	PD-03B	18-Aug-11	MT
PDL	PD-03B	18-Aug-11	
PDL	PD-03B	18-Aug-11	10% FISH REMAINS
PDL	PD-04B	18-Aug-11	
PDL	PD-04B	18-Aug-11	
PDL	PD-04B	18-Aug-11	
PDL	PD-04B	18-Aug-11	10% FISH REMAINS (SCULPIN)
PDL	PD-04B	18-Aug-11	20% INVERTS (DIPTERA), FISH REMAINS
PDL	PD-04B	18-Aug-11	10% DIPTERA
PDL	PD-04B	18-Aug-11	
PDL	PD-04B	18-Aug-11	100% LKTR REMAINS
PDL	PD-04B	18-Aug-11	80% FISH REMAINS (SCULPIN)
PDL	PD-04B	18-Aug-11	
PDL	PD-04B	18-Aug-11	20% CHIRONOMIDS, OTHER UNID INVERTS
PDL	PD-04B	18-Aug-11	MT
PDL	PD-04B	18-Aug-11	MT
PDL	PD-04B	18-Aug-11	MT
PDL	PD-04B	18-Aug-11	
PDL	PD-05A	18-Aug-11	
PDL	PD-06A	18-Aug-11	
PDL	PD-06A	18-Aug-11	
PDL	PD-07B	19-Aug-11	
PDL	PD-07B	19-Aug-11	
PDL	PD-07B	19-Aug-11	
PDL	PD-08A	18-Aug-11	MT
PDL	PD-08B	19-Aug-11	
PDL	PD-08B	19-Aug-11	
PDL	PD-08B	19-Aug-11	
PDL	PD-08B	19-Aug-11	10% ZOOPLANKTON
PDL	PD-08B	19-Aug-11	30% FISH REMAINS (SCULPIN)
PDL	PD-08B	19-Aug-11	
PDL	PD-01	17-Aug-11	80% CHIRONOMIDS, ZOOPLANKTON
PDL	PD-01	17-Aug-11	90% CHIRONOMIDS, TRICHOPTERA
PDL	PD-01	17-Aug-11	90% TRICHOPTERA, CHIRONOMIDS
PDL	PD-01	17-Aug-11	MT
PDL	PD-03B	18-Aug-11	20% TRICHOPTERA, DIPTERA
PDL	PD-03B	18-Aug-11	10% TRICHOPTERA
PDL	PD-07B	19-Aug-11	60% TRICHOPTERA, DIPTERA

APPENDIX I

NORTH/SOUTH CONSULTANTS FISH AGING LABORATORY REPORT



NSC Ageing ID: 237
Company Name: Azimuth
Project Name: Meadowbank
Client Contact: R.Baker/G.Mann
& Type of Structures: 205 OT
Work to be Completed: Set/cut

<u>Location</u>	<u>Date</u>	<u>Structures</u>	<u>Total</u>	<u>Species</u>	<u>Site #</u>	<u>Fish #</u>	<u>Final</u>	<u>Comments</u>
INUG	Aug-11	OT	1	ARCH	2	13	ns	
INUG	Aug-11	OT	1	ARCH	7	15	5	
INUG	Aug-11	OT	1	LKTR	1	1	7	
INUG	Aug-11	OT	1	LKTR	1	8	ns	
INUG	Aug-11	OT	1	LKTR	1	9	10	
INUG	Aug-11	OT	1	LKTR	1	16	19	
INUG	Aug-11	OT	1	LKTR	1	17	17	
INUG	Aug-11	OT	1	LKTR	2	5	8	
INUG	Aug-11	OT	1	LKTR	2	6	13	
INUG	Aug-11	OT	1	LKTR	2	15	12	
INUG	Aug-11	OT	1	LKTR	2	16	21	
INUG	Aug-11	OT	1	LKTR	2	17	22	
INUG	Aug-11	OT	1	LKTR	2	18	26	
INUG	Aug-11	OT	1	LKTR	2	21	4	
INUG	Aug-11	OT	1	LKTR	4	3	9	
INUG	Aug-11	OT	1	LKTR	5	5	18	
INUG	Aug-11	OT	1	LKTR	5	6	17	
INUG	Aug-11	OT	1	LKTR	5	7	23	
INUG	Aug-11	OT	1	LKTR	5	8	18	
INUG	Aug-11	OT	1	LKTR	7	1	18	
INUG	Aug-11	OT	1	LKTR	7	1	7	
INUG	Aug-11	OT	1	LKTR	7	3	17	
INUG	Aug-11	OT	1	LKTR	7	6	10	
INUG	Aug-11	OT	1	LKTR	7	9	6	
INUG	Aug-11	OT	1	LKTR	7	12	8	
INUG	Aug-11	OT	1	LKTR	8	2	28	
INUG	Aug-11	OT	1	LKTR	8	6	8	
INUG	Aug-11	OT	1	LKTR	8	15	8	
INUG	Aug-11	OT	1	LKTR	8	17	22	
INUG	Aug-11	OT	1	LKTR	8	18	28	
INUG	Aug-11	OT	1	LKTR	8	19	21	
INUG	Aug-11	OT	1	LKTR	10	2	22	
INUG	Aug-11	OT	1	LKTR	10	4	23	
INUG	Aug-11	OT	1	LKTR	13	1	11	
INUG	Aug-11	OT	1	LKTR	13	5	40	
INUG	Aug-11	OT	1	LKTR	13	7	17	
INUG	Aug-11	OT	1	LKTR	13	17	20	
INUG	Aug-11	OT	1	LKTR	14	1	10	
INUG	Aug-11	OT	1	LKTR	14	3	13	
INUG	Aug-11	OT	1	LKTR	14	5	9	
INUG	Aug-11	OT	1	LKTR	14	13	24	
INUG	Aug-11	OT	1	LKTR	14	15	20	
INUG	Aug-11	OT	1	LKTR	14	16	14	
INUG	Aug-11	OT	1	LKTR	14	18	21	
INUG	Aug-11	OT	1	LKTR	14	19	9	
INUG	Aug-11	OT	1	RNWH	1	2	16	
INUG	Aug-11	OT	1	RNWH	1	11	24	
INUG	Aug-11	OT	1	RNWH	1	12	16	

<u>Location</u>	<u>Date</u>	<u>Structures</u>	<u>Total</u>	<u>Species</u>	<u>Site #</u>	<u>Fish #</u>	<u>Final</u>	<u>Comments</u>
INUG	Aug-11	OT	1	RNWH	1	13	15	
INUG	Aug-11	OT	1	RNWH	1	14	9	
INUG	Aug-11	OT	1	RNWH	1	15	9	
INUG	Aug-11	OT	1	RNWH	1	18	19	
INUG	Aug-11	OT	1	RNWH	1	19	14	
INUG	Aug-11	OT	1	RNWH	1	21	14	
INUG	Aug-11	OT	1	RNWH	1	22	13	
INUG	Aug-11	OT	1	RNWH	2	4	6	
INUG	Aug-11	OT	1	RNWH	2	8	11	
INUG	Aug-11	OT	1	RNWH	2	14	13	
INUG	Aug-11	OT	1	RNWH	3	1	9	03.01 on slide
INUG	Aug-11	OT	1	RNWH	5	1	15	
INUG	Aug-11	OT	1	RNWH	8	4	16	
INUG	Aug-11	OT	1	RNWH	8	5	15	
INUG	Aug-11	OT	1	RNWH	8	8	17	
INUG	Aug-11	OT	1	RNWH	8	9	16	
INUG	Aug-11	OT	1	RNWH	8	11	20	
INUG	Aug-11	OT	1	RNWH	8	12	18	
INUG	Aug-11	OT	1	RNWH	8	13	7	
INUG	Aug-11	OT	1	RNWH	8	14	8	
INUG	Aug-11	OT	1	RNWH	8	16	13	
INUG	Aug-11	OT	1	RNWH	9	4	19	
INUG	Aug-11	OT	1	RNWH	13	2	15	
INUG	Aug-11	OT	1	RNWH	13	4	17	
INUG	Aug-11	OT	1	RNWH	13	9	22	
INUG	Aug-11	OT	1	RNWH	13	12	15	
INUG	Aug-11	OT	1	RNWH	13	14	15	
INUG	Aug-11	OT	1	RNWH	13	18	19	
INUG	Aug-11	OT	1	RNWH	13	19	17	
INUG	Aug-11	OT	1	RNWH	14	1	10	
INUG	Aug-11	OT	1	RNWH	14	2	12	
INUG	Aug-11	OT	1	RNWH	14	8	13	
INUG	Aug-11	OT	1	RNWH	14	9	18	
INUG	Aug-11	OT	1	RNWH	14	11	17	
INUG	Aug-11	OT	1	RNWH	14	14	12	
TPN	Aug-11	OT	1	ARCH	10	12	6	
TPN	Aug-11	OT	1	ARCH	10	15	5	
TPN	Aug-11	OT	1	ARCH	10	16	4	
TPN	Aug-11	OT	1	ARCH	23	1	11	
TPN	Aug-11	OT	1	ARCH	25	3	5	
TPN	Aug-11	OT	1	ARCH	25	7	4	
TPN	Aug-11	OT	1	ARCH	5	7	5	
TPN	Aug-11	OT	1	ARCH	5	9	5	
TPN	Aug-11	OT	1	ARCH	5	11	2	
TPN	Aug-11	OT	1	ARCH	6	8	8	
TPN	Aug-11	OT	1	ARCH	6	11	9	
TPN	Aug-11	OT	1	ARCH	6	17	3	
TPN	Aug-11	OT	1	ARCH	6	19	4	
TPN	Aug-11	OT	1	ARCH	6	2	5	
TPN	Aug-11	OT	1	ARCH	10	3	3	
TPN	Aug-11	OT	1	ARCH	10	4	4	
TPN	Aug-11	OT	1	ARCH	10	5	3	
TPN	Aug-11	OT	1	LKTR	1	1	15	
TPN	Aug-11	OT	1	LKTR	5	1	21	
TPN	Aug-11	OT	1	LKTR	5	2	19	

<u>Location</u>	<u>Date</u>	<u>Structures</u>	<u>Total</u>	<u>Species</u>	<u>Site #</u>	<u>Fish #</u>	<u>Final</u>	<u>Comments</u>
TPN	Aug-11	OT	1	LKTR	5	3	14	
TPN	Aug-11	OT	1	LKTR	5	12	6	
TPN	Aug-11	OT	1	LKTR	5	13	5	
TPN	Aug-11	OT	1	LKTR	5	15	3	
TPN	Aug-11	OT	1	LKTR	5	21	15	
TPN	Aug-11	OT	1	LKTR	6	2	26	
TPN	Aug-11	OT	1	LKTR	6	3	18	
TPN	Aug-11	OT	1	LKTR	6	5	19	
TPN	Aug-11	OT	1	LKTR	6	14	12	
TPN	Aug-11	OT	1	LKTR	6	15	5	
TPN	Aug-11	OT	1	LKTR	6	18	6	
TPN	Aug-11	OT	1	LKTR	6	21	6	
TPN	Aug-11	OT	1	LKTR	6	23	6	
TPN	Aug-11	OT	1	LKTR	7	1	13	
TPN	Aug-11	OT	1	LKTR	8	2	19	
TPN	Aug-11	OT	1	LKTR	8	3	17	
TPN	Aug-11	OT	1	LKTR	8	6	22	
TPN	Aug-11	OT	1	LKTR	8	7	22	
TPN	Aug-11	OT	1	LKTR	10	9	7	
TPN	Aug-11	OT	1	LKTR	10	1	5	
TPN	Aug-11	OT	1	LKTR	10	13	16	
TPN	Aug-11	OT	1	LKTR	10	14	13	
TPN	Aug-11	OT	1	LKTR	10	19	6	
TPN	Aug-11	OT	1	LKTR	10	2	5	
TPN	Aug-11	OT	1	LKTR	10	21	4	
TPN	Aug-11	OT	1	LKTR	10	22	18	
TPN	Aug-11	OT	1	LKTR	10	23	23	
TPN	Aug-11	OT	1	LKTR	10	25	17	
TPN	Aug-11	OT	1	LKTR	10	26	40	
TPN	Aug-11	OT	1	LKTR	11	1	8	
TPN	Aug-11	OT	1	LKTR	11	2	21	
TPN	Aug-11	OT	1	LKTR	11	3	19	
TPN	Aug-11	OT	1	LKTR	11	5	26	
TPN	Aug-11	OT	1	LKTR	11	8	20	
TPN	Aug-11	OT	1	LKTR	11	9	15	
TPN	Aug-11	OT	1	LKTR	16	4	25	
TPN	Aug-11	OT	1	LKTR	17	1	16	
TPN	Aug-11	OT	1	LKTR	11	1	7	
TPN	Aug-11	OT	1	LKTR	16	6	8	
TPN	Aug-11	OT	1	LKTR	16	9	18	
TPN	Aug-11	OT	1	LKTR	16	12	22	
TPN	Aug-11	OT	1	LKTR	16	13	20	
TPN	Aug-11	OT	1	LKTR	16	14	15	
TPN	Aug-11	OT	1	LKTR	16	17	21	
TPN	Aug-11	OT	1	LKTR	16	21	19	
TPN	Aug-11	OT	1	LKTR	16	22	15	

<u>Location</u>	<u>Date</u>	<u>Structures</u>	<u>Total</u>	<u>Species</u>	<u>Site #</u>	<u>Fish #</u>	<u>Final</u>	<u>Comments</u>
TPN	Aug-11	OT	1	LKTR	16	25	28	
TPN	Aug-11	OT	1	LKTR	16	26	17	
TPN	Aug-11	OT	1	LKTR	16	27	34	
TPN	Aug-11	OT	1	LKTR	16	28	37	
TPN	Aug-11	OT	1	LKTR	16	29	34	
TPN	Aug-11	OT	1	LKTR	22	2	20	
TPN	Aug-11	OT	1	LKTR	22	3	18	
TPN	Aug-11	OT	1	LKTR	24	4	9	
TPN	Aug-11	OT	1	LKTR	24	5	29	
TPN	Aug-11	OT	1	LKTR	25	1	13	
TPN	Aug-11	OT	1	LKTR	25	2	14	
TPN	Aug-11	OT	1	RNWH	10	8	14	
TPN	Aug-11	OT	1	RNWH	11	6	14	
PDL	Aug-11	OT	1	ARCH	1	14	5	
PDL	Aug-11	OT	1	ARCH	1	15	10	
PDL	Aug-11	OT	1	ARCH	3	6	6	
PDL	Aug-11	OT	1	ARCH	3	12	11	
PDL	Aug-11	OT	1	ARCH	8	1	6	
PDL	Aug-11	OT	1	LKTR	1	6	15	
PDL	Aug-11	OT	1	LKTR	1	9	10	
PDL	Aug-11	OT	1	LKTR	1	1	12	
PDL	Aug-11	OT	1	LKTR	1	11	10	
PDL	Aug-11	OT	1	LKTR	1	16	11	
PDL	Aug-11	OT	1	LKTR	1	17	12	
PDL	Aug-11	OT	1	LKTR	2	1	10	
PDL	Aug-11	OT	1	LKTR	2	2	8	
PDL	Aug-11	OT	1	LKTR	3	2	8	
PDL	Aug-11	OT	1	LKTR	3	3	8	
PDL	Aug-11	OT	1	LKTR	3	4	15	
PDL	Aug-11	OT	1	LKTR	3	5	24	
PDL	Aug-11	OT	1	LKTR	3	9	15	
PDL	Aug-11	OT	1	LKTR	3	13	10	
PDL	Aug-11	OT	1	LKTR	3	14	15	
PDL	Aug-11	OT	1	LKTR	3	15	22	
PDL	Aug-11	OT	1	LKTR	3	16	18	
PDL	Aug-11	OT	1	LKTR	3	17	24	
PDL	Aug-11	OT	1	LKTR	3	2	14	
PDL	Aug-11	OT	1	LKTR	4	4	29	
PDL	Aug-11	OT	1	LKTR	4	5	23	
PDL	Aug-11	OT	1	LKTR	4	6	20	
PDL	Aug-11	OT	1	LKTR	4	8	16	
PDL	Aug-11	OT	1	LKTR	4	9	18	
PDL	Aug-11	OT	1	LKTR	4	11	10	
PDL	Aug-11	OT	1	LKTR	4	12	6	
PDL	Aug-11	OT	1	LKTR	4	13	10	
PDL	Aug-11	OT	1	LKTR	4	14	4	
PDL	Aug-11	OT	1	LKTR	8	01A	20	
PDL	Aug-11	OT	1	LKTR	8	5	10	
PDL	Aug-11	OT	1	LKTR	8	6	10	
PDL	Aug-11	OT	1	RNWH	1	3	12	
PDL	Aug-11	OT	1	RNWH	1	4	22	
PDL	Aug-11	OT	1	RNWH	1	7	14	
PDL	Aug-11	OT	1	RNWH	1	8	12	
PDL	Aug-11	OT	1	RNWH	3	8	16	
PDL	Aug-11	OT	1	RNWH	3	18	16	
PDL	Aug-11	OT	1	RNWH	7	3	9	

NSC Ageing ID: 237
Company Name: Azimuth
Project Name: Meadowbank
Client Contact: R.Baker/G.Mann
& Type of Structures: 336 FR
Work to be Completed: Set/cut

<u>Location</u>	<u>Date</u>	<u>Structures</u>	<u>Total</u>	<u>Species</u>	<u>Site #</u>	<u>Fish #</u>	<u>Final</u>	<u>Comments</u>
INUG	Aug-11	FR	1	LKTR	1	8	17	
INUG	Aug-11	FR	1	LKTR	1	9	5	
INUG	Aug-11	FR	1	LKTR	1	10	5	
INUG	Aug-11	FR	1	LKTR	1	16	17	
INUG	Aug-11	FR	1	LKTR	1	17	14	
INUG	Aug-11	FR	1	LKTR	2	5	3	
INUG	Aug-11	FR	1	LKTR	2	6	3	
INUG	Aug-11	FR	1	LKTR	2	15	7	
INUG	Aug-11	FR	1	LKTR	2	16	12	
INUG	Aug-11	FR	1	LKTR	2	17	16	
INUG	Aug-11	FR	1	LKTR	2	18	13	
INUG	Aug-11	FR	1	LKTR	2	21	2	
INUG	Aug-11	FR	1	LKTR	4	3	9	
INUG	Aug-11	FR	1	LKTR	5	5	15	
INUG	Aug-11	FR	1	LKTR	5	6	14	
INUG	Aug-11	FR	1	LKTR	5	7	19	
INUG	Aug-11	FR	1	LKTR	5	8	17	
INUG	Aug-11	FR	1	LKTR	7	1	13	
INUG	Aug-11	FR	1	LKTR	7	3	13	
INUG	Aug-11	FR	1	LKTR	7	6	5	
INUG	Aug-11	FR	1	LKTR	7	9	1	
INUG	Aug-11	FR	1	LKTR	7	10	1	
INUG	Aug-11	FR	1	LKTR	7	12	4	
INUG	Aug-11	FR	1	LKTR	8	6	5	
INUG	Aug-11	FR	1	LKTR	8	15	3	
INUG	Aug-11	FR	1	LKTR	8	17	16	
INUG	Aug-11	FR	1	LKTR	8	18	20	
INUG	Aug-11	FR	1	LKTR	8	19	12	
INUG	Aug-11	FR	1	LKTR	8	20	13	
INUG	Aug-11	FR	1	LKTR	10	2	15	
INUG	Aug-11	FR	1	LKTR	10	4	19	
INUG	Aug-11	FR	1	LKTR	13	5	25	
INUG	Aug-11	FR	1	LKTR	13	7	14	
INUG	Aug-11	FR	1	LKTR	13	10	6	
INUG	Aug-11	FR	1	LKTR	13	17	14	
INUG	Aug-11	FR	1	LKTR	14	1	5	
INUG	Aug-11	FR	1	LKTR	14	3	9	
INUG	Aug-11	FR	1	LKTR	14	5	4	
INUG	Aug-11	FR	1	LKTR	14	13	21	
INUG	Aug-11	FR	1	LKTR	14	15	11	
INUG	Aug-11	FR	1	LKTR	14	16	14	
INUG	Aug-11	FR	1	LKTR	14	18	16	
INUG	Aug-11	FR	1	LKTR	14	19	7	
INUG	Aug-11	FR	1	RNWH	1	2	13	
INUG	Aug-11	FR	1	RNWH	1	11	14	
INUG	Aug-11	FR	1	RNWH	1	12	10	
INUG	Aug-11	FR	1	RNWH	1	13	12	

<u>Location</u>	<u>Date</u>	<u>Structures</u>	<u>Total</u>	<u>Species</u>	<u>Site #</u>	<u>Fish #</u>	<u>Final</u>	<u>Comments</u>
INUG	Aug-11	FR	1	RNWH	1	14	8	
INUG	Aug-11	FR	1	RNWH	1	15	6	
INUG	Aug-11	FR	1	RNWH	1	18	13	
INUG	Aug-11	FR	1	RNWH	1	19	16	
INUG	Aug-11	FR	1	RNWH	1	21	14	
INUG	Aug-11	FR	1	RNWH	1	22	9	
INUG	Aug-11	FR	1	RNWH	2	4	4	
INUG	Aug-11	FR	1	RNWH	2	8	9	
INUG	Aug-11	FR	1	RNWH	2	14	10	
INUG	Aug-11	FR	1	RNWH	3	1	11	
INUG	Aug-11	FR	1	RNWH	5	1	14	
INUG	Aug-11	FR	1	RNWH	5	2	8	
INUG	Aug-11	FR	1	RNWH	8	4	13	
INUG	Aug-11	FR	1	RNWH	8	5	15	
INUG	Aug-11	FR	1	RNWH	8	8	12	
INUG	Aug-11	FR	1	RNWH	8	9	11	
INUG	Aug-11	FR	1	RNWH	8	11	12	
INUG	Aug-11	FR	1	RNWH	8	12	14	
INUG	Aug-11	FR	1	RNWH	8	13	5	
INUG	Aug-11	FR	1	RNWH	8	14	7	
INUG	Aug-11	FR	1	RNWH	8	16	12	
INUG	Aug-11	FR	1	RNWH	9	4	14	
INUG	Aug-11	FR	1	RNWH	13	2	14	
INUG	Aug-11	FR	1	RNWH	13	4	14	
INUG	Aug-11	FR	1	RNWH	13	9	14	
INUG	Aug-11	FR	1	RNWH	13	12	14	
INUG	Aug-11	FR	1	RNWH	13	14	12	
INUG	Aug-11	FR	1	RNWH	13	18	14	
INUG	Aug-11	FR	1	RNWH	13	19	13	
INUG	Aug-11	FR	1	RNWH	14	2	12	
INUG	Aug-11	FR	1	RNWH	14	8	12	
INUG	Aug-11	FR	1	RNWH	14	9	13	
INUG	Aug-11	FR	1	RNWH	14	10	8	
INUG	Aug-11	FR	1	RNWH	14	11	12	
INUG	Aug-11	FR	1	RNWH	14	14	12	
TPN	Aug-11	FR	1	ARCH	6	4	10	
TPN	Aug-11	FR	1	ARCH	6	6	9	
TPN	Aug-11	FR	1	ARCH	10	7	10	
TPN	Aug-11	FR	1	ARCH	10	17	6	
TPN	Aug-11	FR	1	ARCH	11	7	5	
TPN	Aug-11	FR	1	ARCH	16	1	9	
TPN	Aug-11	FR	1	ARCH	16	19	7	
TPN	Aug-11	FR	1	ARCH	24	3	4	
TPN	Aug-11	FR	1	ARCH	25	6	5	
TPN	Aug-11	FR	1	ARCH	25	9	10	
TPN	Aug-11	FR	1	ARCH	25	11	8	
TPN	Aug-11	FR	1	ARCH	5	7	5	
TPN	Aug-11	FR	1	ARCH	5	9	4	
TPN	Aug-11	FR	1	ARCH	5	11	7	
TPN	Aug-11	FR	1	ARCH	6	8	7	
TPN	Aug-11	FR	1	ARCH	6	11	8	
TPN	Aug-11	FR	1	ARCH	6	17	2	
TPN	Aug-11	FR	1	ARCH	6	19	2	
TPN	Aug-11	FR	1	ARCH	6	20	3	
TPN	Aug-11	FR	1	ARCH	10	3	1	
TPN	Aug-11	FR	1	ARCH	10	4	2	
TPN	Aug-11	FR	1	ARCH	10	5	2	
TPN	Aug-11	FR	1	LKTR	1	2	13	
TPN	Aug-11	FR	1	LKTR	1	3	10	
TPN	Aug-11	FR	1	LKTR	2	2	11	

<u>Location</u>	<u>Date</u>	<u>Structures</u>	<u>Total</u>	<u>Species</u>	<u>Site #</u>	<u>Fish #</u>	<u>Final</u>	<u>Comments</u>
TPN	Aug-11	FR	1	LKTR	3	1	14	
TPN	Aug-11	FR	1	LKTR	3	2	13	
TPN	Aug-11	FR	1	LKTR	5	4	16	
TPN	Aug-11	FR	1	LKTR	5	5	17	
TPN	Aug-11	FR	1	LKTR	5	6	6	
TPN	Aug-11	FR	1	LKTR	5	8	3	
TPN	Aug-11	FR	1	LKTR	5	14	3	
TPN	Aug-11	FR	1	LKTR	5	17	2	
TPN	Aug-11	FR	1	LKTR	5	19	25	
TPN	Aug-11	FR	1	LKTR	5	2	21	
TPN	Aug-11	FR	1	LKTR	5	23	17	
TPN	Aug-11	FR	1	LKTR	6	1	17	
TPN	Aug-11	FR	1	LKTR	6	7	17	
TPN	Aug-11	FR	1	LKTR	6	9	12	
TPN	Aug-11	FR	1	LKTR	6	1	10	
TPN	Aug-11	FR	1	LKTR	6	12	18	
TPN	Aug-11	FR	1	LKTR	6	13	15	
TPN	Aug-11	FR	1	LKTR	6	16	18	
TPN	Aug-11	FR	1	LKTR	6	22	15	
TPN	Aug-11	FR	1	LKTR	6	29	14	
TPN	Aug-11	FR	1	LKTR	6	3	17	
TPN	Aug-11	FR	1	LKTR	8	4	12	
TPN	Aug-11	FR	1	LKTR	8	5	15	
TPN	Aug-11	FR	1	LKTR	8	8	26	
TPN	Aug-11	FR	1	LKTR	8	9	14	
TPN	Aug-11	FR	1	LKTR	10	1	22	
TPN	Aug-11	FR	1	LKTR	10	6	14	
TPN	Aug-11	FR	1	LKTR	10	11	11	
TPN	Aug-11	FR	1	LKTR	10	18	8	
TPN	Aug-11	FR	1	LKTR	10	24	14	
TPN	Aug-11	FR	1	LKTR	11	4	5	
TPN	Aug-11	FR	1	LKTR	14	1	14	
TPN	Aug-11	FR	1	LKTR	16	2	14	
TPN	Aug-11	FR	1	LKTR	16	3	17	
TPN	Aug-11	FR	1	LKTR	11	11	20	
TPN	Aug-11	FR	1	LKTR	11	12	17	
TPN	Aug-11	FR	1	LKTR	16	7	6	
TPN	Aug-11	FR	1	LKTR	16	8	18	
TPN	Aug-11	FR	1	LKTR	16	1	16	
TPN	Aug-11	FR	1	LKTR	16	11	15	
TPN	Aug-11	FR	1	LKTR	16	15	18	
TPN	Aug-11	FR	1	LKTR	16	18	26	
TPN	Aug-11	FR	1	LKTR	16	2	19	
TPN	Aug-11	FR	1	LKTR	16	23	18	
TPN	Aug-11	FR	1	LKTR	16	24	15	
TPN	Aug-11	FR	1	LKTR	20	1	6	
TPN	Aug-11	FR	1	LKTR	22	1	18	
TPN	Aug-11	FR	1	LKTR	22	4	16	
TPN	Aug-11	FR	1	LKTR	22	5	25	
TPN	Aug-11	FR	1	LKTR	24	1	6	
TPN	Aug-11	FR	1	LKTR	24	2	10	
TPN	Aug-11	FR	1	LKTR	25	4	22	
TPN	Aug-11	FR	1	LKTR	25	5	18	
TPN	Aug-11	FR	1	LKTR	25	8	19	
TPN	Aug-11	FR	1	LKTR	25	10	21	
TPN	Aug-11	FR	1	LKTR	1	1	17	
TPN	Aug-11	FR	1	LKTR	5	1	10	
TPN	Aug-11	FR	1	LKTR	5	2	15	
TPN	Aug-11	FR	1	LKTR	5	3	11	
TPN	Aug-11	FR	1	LKTR	5	12	4	

<u>Location</u>	<u>Date</u>	<u>Structures</u>	<u>Total</u>	<u>Species</u>	<u>Site #</u>	<u>Fish #</u>	<u>Final</u>	<u>Comments</u>
TPN	Aug-11	FR	1	LKTR	5	13	5	
TPN	Aug-11	FR	1	LKTR	5	15	4	
TPN	Aug-11	FR	1	LKTR	5	21	15	
TPN	Aug-11	FR	1	LKTR	6	2	15	
TPN	Aug-11	FR	1	LKTR	6	3	14	
TPN	Aug-11	FR	1	LKTR	6	5	14	
TPN	Aug-11	FR	1	LKTR	6	14	8	
TPN	Aug-11	FR	1	LKTR	6	15	4	
TPN	Aug-11	FR	1	LKTR	6	18	ua	
TPN	Aug-11	FR	1	LKTR	6	21	3	
TPN	Aug-11	FR	1	LKTR	6	23	4	
TPN	Aug-11	FR	1	LKTR	7	1	13	
TPN	Aug-11	FR	1	LKTR	8	2	15	
TPN	Aug-11	FR	1	LKTR	8	3	14	
TPN	Aug-11	FR	1	LKTR	8	6	11	
TPN	Aug-11	FR	1	LKTR	8	7	14	
TPN	Aug-11	FR	1	LKTR	10	9	4	
TPN	Aug-11	FR	1	LKTR	10	1	4	
TPN	Aug-11	FR	1	LKTR	10	13	9	
TPN	Aug-11	FR	1	LKTR	10	14	6	
TPN	Aug-11	FR	1	LKTR	10	19	4	
TPN	Aug-11	FR	1	LKTR	10	20	4	
TPN	Aug-11	FR	1	LKTR	10	21	3	
TPN	Aug-11	FR	1	LKTR	10	22	9	
TPN	Aug-11	FR	1	LKTR	10	23	17	
TPN	Aug-11	FR	1	LKTR	10	25	16	
TPN	Aug-11	FR	1	LKTR	10	26	15	
TPN	Aug-11	FR	1	LKTR	11	1	5	
TPN	Aug-11	FR	1	LKTR	11	2	14	
TPN	Aug-11	FR	1	LKTR	11	3	14	
TPN	Aug-11	FR	1	LKTR	11	5	13	
TPN	Aug-11	FR	1	LKTR	11	8	13	
TPN	Aug-11	FR	1	LKTR	11	9	11	
TPN	Aug-11	FR	1	LKTR	16	4	21	
TPN	Aug-11	FR	1	LKTR	17	1	12	
TPN	Aug-11	FR	1	LKTR	11	10	7	
TPN	Aug-11	FR	1	LKTR	16	6	3	
TPN	Aug-11	FR	1	LKTR	16	9	14	
TPN	Aug-11	FR	1	LKTR	16	12	14	
TPN	Aug-11	FR	1	LKTR	16	13	11	
TPN	Aug-11	FR	1	LKTR	16	14	11	
TPN	Aug-11	FR	1	LKTR	16	17	19	
TPN	Aug-11	FR	1	LKTR	16	21	17	
TPN	Aug-11	FR	1	LKTR	16	22	15	
TPN	Aug-11	FR	1	LKTR	16	25	26	
TPN	Aug-11	FR	1	LKTR	16	26	17	
TPN	Aug-11	FR	1	LKTR	16	27	25	
TPN	Aug-11	FR	1	LKTR	16	28	ua	cut too high
TPN	Aug-11	FR	1	LKTR	16	29	20	
TPN	Aug-11	FR	1	LKTR	22	2	15	
TPN	Aug-11	FR	1	LKTR	22	3	13	
TPN	Aug-11	FR	1	LKTR	24	4	5	
TPN	Aug-11	FR	1	LKTR	24	5	22	
TPN	Aug-11	FR	1	LKTR	25	1	9	
TPN	Aug-11	FR	1	LKTR	25	2	7	
TPN	Aug-11	FR	1	RNWH	10	8	14	
TPN	Aug-11	FR	1	RNWH	11	6	14	
PDL	Aug-11	FR	1	LKTR	1	1	20	
PDL	Aug-11	FR	1	LKTR	1	2	26	
PDL	Aug-11	FR	1	LKTR	1	5	21	

<u>Location</u>	<u>Date</u>	<u>Structures</u>	<u>Total</u>	<u>Species</u>	<u>Site #</u>	<u>Fish #</u>	<u>Final</u>	<u>Comments</u>
PDL	Aug-11	FR	1	LKTR	1	12	30	too high
PDL	Aug-11	FR	1	LKTR	1	13	ua	
PDL	Aug-11	FR	1	LKTR	3	1	14	
PDL	Aug-11	FR	1	LKTR	3	1	18	
PDL	Aug-11	FR	1	LKTR	3	11	5	
PDL	Aug-11	FR	1	LKTR	3	19	ns	
PDL	Aug-11	FR	1	LKTR	4	1	9	
PDL	Aug-11	FR	1	LKTR	4	2	20	
PDL	Aug-11	FR	1	LKTR	4	3	28	
PDL	Aug-11	FR	1	LKTR	4	7	17	
PDL	Aug-11	FR	1	LKTR	4	1	24	
PDL	Aug-11	FR	1	LKTR	5	1	22	
PDL	Aug-11	FR	1	LKTR	6	1	16	
PDL	Aug-11	FR	1	LKTR	6	2	17	
PDL	Aug-11	FR	1	LKTR	7	1	4	
PDL	Aug-11	FR	1	LKTR	7	2	3	
PDL	Aug-11	FR	1	LKTR	7	4	4	
PDL	Aug-11	FR	1	LKTR	8	2	20	
PDL	Aug-11	FR	1	LKTR	8	3	22	
PDL	Aug-11	FR	1	LKTR	8	4	6	
PDL	Aug-11	FR	1	LKTR	8	7	2	
PDL	Aug-11	FR	1	LKTR	1	6	10	
PDL	Aug-11	FR	1	LKTR	1	9	7	
PDL	Aug-11	FR	1	LKTR	1	1	8	
PDL	Aug-11	FR	1	LKTR	1	11	8	
PDL	Aug-11	FR	1	LKTR	1	16	5	
PDL	Aug-11	FR	1	LKTR	1	17	5	
PDL	Aug-11	FR	1	LKTR	2	1	8	
PDL	Aug-11	FR	1	LKTR	2	2	7	
PDL	Aug-11	FR	1	LKTR	3	2	8	
PDL	Aug-11	FR	1	LKTR	3	3	5	
PDL	Aug-11	FR	1	LKTR	3	4	12	
PDL	Aug-11	FR	1	LKTR	3	5	13	
PDL	Aug-11	FR	1	LKTR	3	9	10	
PDL	Aug-11	FR	1	LKTR	3	13	6	
PDL	Aug-11	FR	1	LKTR	3	14	4	
PDL	Aug-11	FR	1	LKTR	3	15	12	
PDL	Aug-11	FR	1	LKTR	3	16	11	
PDL	Aug-11	FR	1	LKTR	3	17	14	
PDL	Aug-11	FR	1	LKTR	3	2	10	
PDL	Aug-11	FR	1	LKTR	4	4	16	
PDL	Aug-11	FR	1	LKTR	4	5	10	
PDL	Aug-11	FR	1	LKTR	4	6	12	
PDL	Aug-11	FR	1	LKTR	4	8	6	
PDL	Aug-11	FR	1	LKTR	4	9	10	
PDL	Aug-11	FR	1	LKTR	4	11	4	
PDL	Aug-11	FR	1	LKTR	4	12	4	
PDL	Aug-11	FR	1	LKTR	4	13	4	
PDL	Aug-11	FR	1	LKTR	4	14	3	
PDL	Aug-11	FR	1	LKTR	8	01A	18	
PDL	Aug-11	FR	1	LKTR	8	5	4	
PDL	Aug-11	FR	1	LKTR	8	6	4	
PDL	Aug-11	FR	1	RNWH	1	3	12	
PDL	Aug-11	FR	1	RNWH	1	4	14	
PDL	Aug-11	FR	1	RNWH	1	7	8	
PDL	Aug-11	FR	1	RNWH	1	8	5	
PDL	Aug-11	FR	1	RNWH	3	8	14	
PDL	Aug-11	FR	1	RNWH	3	18	12	
PDL	Aug-11	FR	1	RNWH	7	3	6	
INUG	Aug-11	FR	1	ARCH	2	13	4	

<u>Location</u>	<u>Date</u>	<u>Structures</u>	<u>Total</u>	<u>Species</u>	<u>Site #</u>	<u>Fish #</u>	<u>Final</u>	<u>Comments</u>
INUG	Aug-11	FR	1	ARCH	7	2	14	
INUG	Aug-11	FR	1	ARCH	7	15	2	
INUG	Aug-11	FR	1	ARCH	8	1	4	
INUG	Aug-11	FR	1	ARCH	14	7	3	
INUG	Aug-11	FR	1	ARGR	13	6	6	
INUG	Aug-11	FR	1	LKTR	1	2	24	
INUG	Aug-11	FR	1	LKTR	1	3	22	
INUG	Aug-11	FR	1	LKTR	1	4	27	
INUG	Aug-11	FR	1	LKTR	1	5	14	
INUG	Aug-11	FR	1	LKTR	1	6	16	
INUG	Aug-11	FR	1	LKTR	1	7	17	
INUG	Aug-11	FR	1	LKTR	2	1	8	
INUG	Aug-11	FR	1	LKTR	2	1	16	
INUG	Aug-11	FR	1	LKTR	2	2	16	
INUG	Aug-11	FR	1	LKTR	2	7	15	
INUG	Aug-11	FR	1	LKTR	2	9	14	
INUG	Aug-11	FR	1	LKTR	2	11	22	
INUG	Aug-11	FR	1	LKTR	2	12	17	
INUG	Aug-11	FR	1	LKTR	2	19	30+	
INUG	Aug-11	FR	1	LKTR	4	1	7	
INUG	Aug-11	FR	1	LKTR	4	2	20	
INUG	Aug-11	FR	1	LKTR	5	3	19	
INUG	Aug-11	FR	1	LKTR	5	4	9	
INUG	Aug-11	FR	1	LKTR	6	1	5	
INUG	Aug-11	FR	1	LKTR	7	4	15	
INUG	Aug-11	FR	1	LKTR	7	5	14	
INUG	Aug-11	FR	1	LKTR	7	7	17	
INUG	Aug-11	FR	1	LKTR	7	8	4	
INUG	Aug-11	FR	1	LKTR	7	11	12	
INUG	Aug-11	FR	1	LKTR	7	13	5	
INUG	Aug-11	FR	1	LKTR	8	1	15	
INUG	Aug-11	FR	1	LKTR	8	2	18	
INUG	Aug-11	FR	1	LKTR	8	3	13	
INUG	Aug-11	FR	1	LKTR	8	7	15	
INUG	Aug-11	FR	1	LKTR	9	1	6	
INUG	Aug-11	FR	1	LKTR	9	2	28	
INUG	Aug-11	FR	1	LKTR	9	3	19	
INUG	Aug-11	FR	1	LKTR	9	5	12	
INUG	Aug-11	FR	1	LKTR	10	1	27	
INUG	Aug-11	FR	1	LKTR	13	1	13	
INUG	Aug-11	FR	1	LKTR	13	3	17	
INUG	Aug-11	FR	1	LKTR	13	8	4	
INUG	Aug-11	FR	1	LKTR	13	11	14	
INUG	Aug-11	FR	1	LKTR	13	13	4	
INUG	Aug-11	FR	1	LKTR	13	15	3	
INUG	Aug-11	FR	1	LKTR	13	16	3	
INUG	Aug-11	FR	1	LKTR	14	6	8	
INUG	Aug-11	FR	1	LKTR	14	12	20	
INUG	Aug-11	FR	1	LKTR	14	17	14	

APPENDIX J

SUMMARY STATISTICS FOR OPPORTUNISTICALLY- COLLECTED LETHAL PARAMETERS



Species	Area	Gonad Weight (g)**							Egg Weight (g/400 eggs)**						
		n	mean	median	SD	SE	min	max	n	mean	median	SD	SE	min	max
LKTR	TPN	0	--	--	--	--	--	--	0	--	--	--	--	--	--
LKTR	INUG	8	106.4	80.3	87.0	30.8	12.1	226.0	4	29.18	29.32	4.95	2.47	22.98	35.08
LKTR	PDL	7	163.1	76.6	125.8	47.6	62.7	390.0	4	26.08	28.93	10.39	5.20	11.26	35.19
RNWH*	TPN	2	24.6	24.6	7.1	5.1	19.5	29.6	1	1.02	1.02	--	--	1.02	1.02
RNWH*	INUG	34	19.5	20.4	7.4	1.3	3.3	33.7	14	0.97	0.93	0.30	0.08	0.47	1.81
RNWH*	PDL	6	26.7	26.8	13.1	5.3	10.3	48.8	4	0.86	0.88	0.07	0.04	0.76	0.92
ARCH*	TPN	0	--	--	--	--	--	--	0	--	--	--	--	--	--
ARCH*	INUG	0	--	--	--	--	--	--	0	--	--	--	--	--	--
ARCH*	PDL	0	--	--	--	--	--	--	0	--	--	--	--	--	--

Species	Area	Fecundity (number of eggs)**							Liver Weight (g)						
		n	mean	median	SD	SE	min	max	n	mean	median	SD	SE	min	max
LKTR	TPN	0	--	--	--	--	--	--	0	--	--	--	--	--	--
LKTR	INUG	4	2489	2424	450	225	2052	3057	43	15.37	11.44	16.35	2.49	0.45	68.46
LKTR	PDL	4	3356	3230	797	399	2533	4433	31	10.98	2.33	16.62	2.99	0.22	68.74
RNWH*	TPN	1	11608	11608	NA	NA	11608	11608	1	8.20	8.20	--	--	8.20	8.20
RNWH*	INUG	14	9303	8239	3449	922	4590	18877	38	4.80	4.32	2.48	0.40	0.79	10.06
RNWH*	PDL	4	10173	11263	4271	2136	4465	13700	7	6.52	5.93	3.47	1.31	1.19	11.82
ARCH*	TPN	0	--	--	--	--	--	--	0	--	--	--	--	--	--
ARCH*	INUG	0	--	--	--	--	--	--	2	1.31	1.31	1.20	0.85	0.46	2.15
ARCH*	PDL	0	--	--	--	--	--	--	5	2.01	2.02	0.18	0.08	1.79	2.22

Notes: n = sample size; SD = standard deviation; SE = standard error; min = minimum; max = maximum; -- = not applicable.

* = non-sentinel species.

** = mature fish only.

APPENDIX K

**ZEAS BENTHIC INVERTEBRATE TAXONOMIC
IDENTIFICATION RAW DATA**



TABLE 1: BENTHIC MACROINVERTEBRATES COLLECTED FROM NUNAVUT MINE LAKES - EEM (2011).

Project Number Station Replicate	AE-11-01--EEM					PDL					TPN				
	1-A	2-A	3-A	4-A	5-A	1-A	2-A	3-A	4-A	5-A	1-A	2-A	3-A	4-A	5-A
ROUNDWORMS															
P. Nemata	-	2	1	-	1	-	2	-	1	-	1	-	1	-	-
ANNELIDS															
P. Annelida															
WORMS															
Cl. Oligochaeta															
F. Tubificidae															
<i>Tasserkidrilus americanus</i>	-	-	-	-	-	-	-	-	-	2	1	-	-	-	-
immatures with hair chaetae	-	-	-	-	-	-	-	-	-	-	-	2	-	-	-
F. Lumbriculidae															
<i>Lumbriculus</i>	-	2	-	1	-	-	-	-	-	3	-	1	-	-	-
ARTHROPODS															
P. Arthropoda															
MITES															
Cl. Arachnida															
O. Acarina	-	-	-	1	-	1	1	-	1	-	2	-	-	-	-
SEED SHRIMPS															
Cl. Ostracoda	-	-	-	-	-	-	2	-	-	-	-	-	1	-	-
INSECTS															
Cl. Insecta															
CADDISFLIES															
O. Trichoptera															
F. Limnephilidae															
<i>Grensia praeterita</i>	-	-	-	-	-	-	-	1	-	-	-	-	-	-	-
TRUE FLIES															
O. Diptera															
MIDGES															
F. Chironomidae															
chironomid pupae	-	-	1	-	-	-	-	2	-	-	-	1	-	1	1
S.F. Chironominae															
<i>Corynocera</i>	-	-	-	-	1	-	-	-	-	-	-	-	-	-	-
<i>Microsetra</i>	-	-	-	2	-	-	-	-	-	-	2	-	-	-	-
<i>Microtendipes</i>	-	-	-	-	2	-	-	-	-	-	-	-	-	-	-
<i>Paratanytarsus</i>	-	-	-	-	2	-	-	-	-	-	-	-	-	-	-
<i>Stictochironomus</i>	-	-	1	-	1	-	13	14	-	1	4	-	-	-	-
<i>Tanytarsus</i>	-	-	2	1	1	-	2	-	-	-	2	-	-	-	-
S.F. Diamesinae															
<i>Protanypus</i>	-	-	2	-	-	-	-	-	-	-	-	-	-	-	-
<i>Pseudodiamesa</i>	-	-	-	-	-	-	-	-	-	1	-	-	-	-	-
S.F. Orthocladiinae															
<i>Heterotrissocladius</i>	-	-	-	1	1	-	-	-	1	-	-	-	1	-	-
<i>Paracladius</i>	-	-	1	-	-	-	-	-	-	-	-	-	-	-	-
<i>Psectrocladius</i>	1	-	-	1	-	-	2	-	1	-	-	-	-	-	1
<i>Zalutschia</i>	-	-	-	-	-	-	-	1	1	1	-	-	-	-	-
indeterminate	-	-	-	-	-	-	-	-	-	1	-	-	-	-	-
S.F. Prodiamesinae															
<i>Monodiamesa</i>	-	-	3	-	1	-	-	-	-	-	-	-	-	-	-
S.F. Tanypodinae															
<i>Ablabesmyia</i>	-	-	-	1	-	-	-	-	-	-	-	-	-	-	-
<i>Procladius</i>	-	3	5	3	6	-	1	1	2	6	1	3	-	1	-
<i>Thienemannimyia</i> complex	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1
MOLLUSCS															
P. Mollusca															
CLAMS															
Cl. Bivalvia															
F. Sphaeriidae															
<i>Cyclocalyx/Neopisidium</i>	-	9	10	11	4	1	3	10	7	1	15	14	5	2	-
<i>Cyclocalyx</i>	-	-	4	-	-	-	-	-	-	5	-	-	-	-	-
<i>Sphaerium nitidum</i>	-	-	-	-	1	-	-	-	-	-	-	-	-	-	-
TOTAL NUMBER OF ORGANISMS	1	16	30	22	21	2	26	29	14	21	28	21	8	4	3
TOTAL NUMBER OF TAXA ^a	1	4	9	9	11	2	8	5	7	9	8	4	4	2	2

^a Bold entries excluded from taxa count

APPENDIX L

**BENTHIC INVERTEBRATE SUPPORTING TABLES:
FAMILY DENSITY, FAMILY PROPORTION, FAMILY
PRESENCE/ABSENCE MATRIX, RAW FAMILY DATA,
AND RAW LOWEST PRACTICAL LEVEL DATA**



Family Density (#/m ²)								
	F. Tubificidae	F. Lumbriculidae	O. Acarina	F. Triopsidae	F. Limnephilidae	F. Chironomidae	F. Sphaeriidae	
INUG								
1	14	14	0	0	0	174	116	
2	0	58	14	0	0	304	406	
3	0	43	29	0	0	377	522	
4	0	72	29	72	0	797	812	
5	0	43	14	0	0	464	261	
Mean	2.9	46	17	14	0	423	423	
SE	2.9	9.6	5.4	14	0	105	119	
PDL								
1	43	0	58	0	0	420	217	
2	0	14	14	0	14	580	275	
3	43	58	43	0	14	623	420	
4	0	0	29	0	0	304	304	
5	58	43	14	0	14	609	188	
Mean	29	23	32	0	8.7	507	281	
SE	12	12	8.5	0	3.5	62	40	
TPN								
1	14	14	29	0	0	377	464	
2	58	14	0	0	0	391	391	
3	0	0	14	0	0	159	87	
4	0	0	29	0	0	290	217	
5	0	0	0	0	14	130	72	
Mean	14	5.8	14	0	2.9	270	246	
SE	11	3.5	6.5	0	2.9	54	79	

Family Proportion (%)								
	F. Tubificidae	F. Lumbriculidae	O. Acarina	F. Triopsidae	F. Limnephilidae	F. Chironomidae	F. Sphaeriidae	
INUG								
1	4.5	4.5	0	0	0	55	36	
2	0	7.4	1.9	0	0	39	52	
3	0	4.5	3.0	0	0	39	54	
4	0	4.1	1.6	4.1	0	45	46	
5	0	5.6	1.9	0	0	59	33	
Mean	0.9	5.2	1.7	0.8	0	47	44	
SE	0.9	0.6	0.5	0.8	0	4.2	4.1	
PDL								
1	5.9	0	7.8	0	0	57	29	
2	0	1.6	1.6	0	1.6	65	31	
3	3.6	4.8	3.6	0	1.2	52	35	
4	0	0	4.5	0	0	48	48	
5	6.2	4.7	1.6	0	1.6	66	20	
Mean	3.1	2.2	3.8	0	0.9	57	33	
SE	1.4	1.1	1.2	0	0	3.5	4.5	
TPN								
1	1.6	1.6	3.2	0	0	42	52	
2	6.8	1.7	0	0	0	46	46	
3	0	0	5.6	0	0	61	33	
4	0	0	5.4	0	0	54	41	
5	0	0	0	0	6.7	60	33	
Mean	1.7	0.7	2.8	0	1	53	41	
SE	1.3	0.4	1.2	0	1	3.8	3.6	

Presence (+) / Absence (-) matrix of benthic invertebrate families for each replicate station.

Area ID	INUG	INUG	INUG	INUG	INUG
Replicate ID	1	2	3	4	5
Depth (m)	6.9	6.8	7.6	8.5	9.2
# Subsamples/replicate	3	3	3	3	3
Mesh Size (µm)	500	500	500	500	500
Collection Date	14-Aug-11	14-Aug-11	14-Aug-11	14-Aug-11	14-Aug-11
<u>ANNELIDS</u> (P. Annelida)					
Cl. Oligochaeta (Worms)					
F. Tubificidae	+	-	-	-	-
F. Lumbriculidae	+	+	+	+	+
<u>ARTHROPODS</u> (P. Arthropoda)					
Cl. Arachnida					
O. Acarina (Mites)	-	+	+	+	+
Cl. Branchiopoda (Fairy shrimp)					
O. Notostraca					
F. Triopsidae	-	-	-	+	-
<u>INSECTS</u> (Cl. Insecta)					
O. Trichoptera (Caddisflies)					
F. Limnephilidae	-	-	-	-	-
O. Diptera (True flies)					
F. Chironomidae (Midges)	+	+	+	+	+
<u>MOLLUSCS</u> (P. Mollusca)					
Cl. Bivalvia (Bivalves)					
F. Sphaeriidae (Fingernail clams)	+	+	+	+	+

Presence (+) / Absence (-) matrix of benthic invertebrate families for each replicate station.

Area ID	PDL	PDL	PDL	PDL	PDL
Replicate ID	1	2	3	4	5
Depth (m)	6.5	6.8	6.9	7.6	6.7
# Subsamples/replicate	3	3	3	3	3
Mesh Size (µm)	500	500	500	500	500
Collection Date	13-Aug-11	13-Aug-11	13-Aug-11	13-Aug-11	13-Aug-11
<u>ANNELIDS</u> (P. Annelida)					
Cl. Oligochaeta (Worms)					
F. Tubificidae	+	-	+	-	+
F. Lumbriculidae	-	+	+	-	+
<u>ARTHROPODS</u> (P. Arthropoda)					
Cl. Arachnida					
O. Acarina (Mites)	+	+	+	+	+
Cl. Branchiopoda (Fairy shrimp)					
O. Notostraca					
F. Triopsidae	-	-	-	-	-
<u>INSECTS</u> (Cl. Insecta)					
O. Trichoptera (Caddisflies)					
F. Limnephilidae	-	+	+	-	+
O. Diptera (True flies)					
F. Chironomidae (Midges)	+	+	+	+	+
<u>MOLLUSCS</u> (P. Mollusca)					
Cl. Bivalvia (Bivalves)					
F. Sphaeriidae (Fingernail clams)	+	+	+	+	+

Presence (+) / Absence (-) matrix of benthic invertebrate families for each replicate station.

Area ID	TPN	TPN	TPN	TPN	TPN
Replicate ID	1	2	3	4	5
Depth (m)	7.0	9.0	9.2	8.6	8.4
# Subsamples/replicate	3	3	3	3	3
Mesh Size (µm)	500	500	500	500	500
Collection Date	19-Aug-11	19-Aug-11	19-Aug-11	19-Aug-11	19-Aug-11
<u>ANNELIDS</u> (P. Annelida)					
Cl. Oligochaeta (Worms)					
F. Tubificidae	+	+	-	-	-
F. Lumbriculidae	+	+	-	-	-
<u>ARTHROPODS</u> (P. Arthropoda)					
Cl. Arachnida					
O. Acarina (Mites)	+	-	+	+	-
Cl. Branchiopoda (Fairy shrimp)					
O. Notostraca					
F. Triopsidae	-	-	-	-	-
<u>INSECTS</u> (Cl. Insecta)					
O. Trichoptera (Caddisflies)					
F. Limnephilidae	-	-	-	-	+
O. Diptera (True flies)					
F. Chironomidae (Midges)	+	+	+	+	+
<u>MOLLUSCS</u> (P. Mollusca)					
Cl. Bivalvia (Bivalves)					
F. Sphaeriidae (Fingernail clams)	+	+	+	+	+

Raw benthic invertebrate family data for each replicate station.

Area ID	INUG	INUG	INUG	INUG	INUG
Replicate ID	1	2	3	4	5
Depth (m)	6.9	6.8	7.6	8.5	9.2
# Subsamples/replicate	3	3	3	3	3
Mesh Size (µm)	500	500	500	500	500
Collection Date	14-Aug-11	14-Aug-11	14-Aug-11	14-Aug-11	14-Aug-11
<u>ROUNDWORMS</u> (P. Nemata) ¹		3	2	2	4
<u>ANNELIDS</u> (P. Annelida)					
Cl. Oligochaeta (Worms)					
F. Tubificidae	1				
F. Lumbriculidae	1	4	3	5	3
<u>ARTHROPODS</u> (P. Arthropoda)					
Cl. Arachnida					
O. Acarina (Mites)		1	2	2	1
Cl. Ostracoda (Seed shrimp) ¹	2	1	3	5	
Cl. Branchiopoda (Fairy shrimp)					
O. Notostraca					
F. Triopsidae				5	
<u>INSECTS</u> (Cl. Insecta)					
O. Trichoptera (Caddisflies)					
F. Limnephilidae					
O. Diptera (True flies)					
F. Chironomidae (Midges)	12	21	26	55	32
<u>MOLLUSCS</u> (P. Mollusca)					
Cl. Bivalvia (Bivalves)					
F. Sphaeriidae (Fingernail clams)	8	28	36	56	18

Raw benthic invertebrate family data for each replicate station.

Area ID	PDL	PDL	PDL	PDL	PDL
Replicate ID	1	2	3	4	5
Depth (m)	6.5	6.8	6.9	7.6	6.7
# Subsamples/replicate	3	3	3	3	3
Mesh Size (µm)	500	500	500	500	500
Collection Date	13-Aug-11	13-Aug-11	13-Aug-11	13-Aug-11	13-Aug-11
<u>ROUNDWORMS</u> (P. Nemata) ¹	1	2		2	2
<u>ANNELIDS</u> (P. Annelida)					
Cl. Oligochaeta (Worms)					
F. Tubificidae	3		3		4
F. Lumbriculidae		1	4		3
<u>ARTHROPODS</u> (P. Arthropoda)					
Cl. Arachnida					
O. Acarina (Mites)	4	1	3	2	1
Cl. Ostracoda (Seed shrimp) ¹	5	4			
Cl. Branchiopoda (Fairy shrimp)					
O. Notostraca					
F. Triopsidae					
<u>INSECTS</u> (Cl. Insecta)					
O. Trichoptera (Caddisflies)					
F. Limnephilidae		1	1		1
O. Diptera (True flies)					
F. Chironomidae (Midges)	29	40	43	21	42
<u>MOLLUSCS</u> (P. Mollusca)					
Cl. Bivalvia (Bivalves)					
F. Sphaeriidae (Fingernail clams)	15	19	29	21	13

Raw benthic invertebrate family data for each replicate station.

Area ID	TPN	TPN	TPN	TPN	TPN
Replicate ID	1	2	3	4	5
Depth (m)	7.0	9.0	9.2	8.6	8.4
# Subsamples/replicate	3	3	3	3	3
Mesh Size (µm)	500	500	500	500	500
Collection Date	19-Aug-11	19-Aug-11	19-Aug-11	19-Aug-11	19-Aug-11
<u>ROUNDWORMS</u> (P. Nemata) ¹	4		2		
<u>ANNELIDS</u> (P. Annelida)					
Cl. Oligochaeta (Worms)					
F. Tubificidae	1	4			
F. Lumbriculidae	1	1			
<u>ARTHROPODS</u> (P. Arthropoda)					
Cl. Arachnida					
O. Acarina (Mites)	2		1	2	
Cl. Ostracoda (Seed shrimp) ¹	1	1	2	2	6
Cl. Branchiopoda (Fairy shrimp)					
O. Notostraca					
F. Triopsidae					
<u>INSECTS</u> (Cl. Insecta)					
O. Trichoptera (Caddisflies)					
F. Limnephilidae					1
O. Diptera (True flies)					
F. Chironomidae (Midges)	26	27	11	20	9
<u>MOLLUSCS</u> (P. Mollusca)					
Cl. Bivalvia (Bivalves)					
F. Sphaeriidae (Fingernail clams)	32	27	6	15	5

Raw benthic invertebrate data identified to the lowest practical level.

Area ID	INUG	INUG	INUG	INUG	INUG
Replicate ID	1	2	3	4	5
Depth (m)	6.9	6.8	7.6	8.5	9.2
# Subsamples/replicate	3	3	3	3	3
Mesh Size (µm)	500	500	500	500	500
Collection Date	14-Aug-11	14-Aug-11	14-Aug-11	14-Aug-11	14-Aug-11
ROUNDWORMS (P. Nematoda) ¹		3	2	2	4
ANNELIDS (P. Annelida)					
Cl. Oligochaeta (Worms)					
F. Tubificidae					
<i>Tasserkidrilus americanus</i>					
immatures with hair chaetae ²	1				
F. Lumbriculidae					
<i>Lumbriculus</i>	1	4	3	5	3
ARTHROPODS (P. Arthropoda)					
Cl. Arachnida					
O. Acarina (Mites)		1	2	2	1
Cl. Ostracoda (Seed shrimp) ¹	2	1	3	5	
Cl. Branchiopoda (Fairy shrimp)					
O. Notostraca					
F. Triopsidae					
<i>Lepidurus arcticus</i>				5	
INSECTS (Cl. Insecta)					
O. Trichoptera (Caddisflies)					
F. Limnephilidae					
<i>Grensia praeterita</i>					
O. Diptera (True flies)					
F. Chironomidae (Midges)					
chironomid pupae ²			1		1
S.F. Chironominae					
<i>Corynocera</i>					1
<i>Microspectra</i>		1	2	5	1
<i>Microtendipes</i>		1		3	3
<i>Paratanytarsus</i>				2	4
<i>Stictochironomus</i>	2	9	3	1	2
<i>Tanytarsus</i>			4	3	1
S.F. Diamesinae					
<i>Protanypus</i>	1		2		
<i>Pseudodiamesa</i>					
S.F. Orthocladiinae					
<i>Abiskomyia</i>	2				
<i>Cricotopus</i>					
<i>Heterotrissocladius</i>				2	3
<i>Hydrobaenus</i>					
<i>Paracladius</i>			1		
<i>Psectrocladius</i>	1			1	
<i>Psectrocladius</i>	1			5	
<i>Zalutschia</i>				1	
indeterminate ²					
S.F. Prodiamesinae					
<i>Monodiamesa</i>	1		3	2	1
S.F. Tanypodinae					
<i>Ablabesmyia</i>				1	
<i>Procladius</i>	4	10	10	29	15
<i>Thienemannimyia</i> complex					
MOLLUSCS (P. Mollusca)					
Cl. Bivalvia (Bivalves)					
F. Sphaeriidae (Fingernail clams)					
<i>Cyclocalyx/Neopisidium</i>	8	22	27	51	13
<i>Cyclocalyx</i>		5	5	1	2
<i>Cyclocalyx nitidum</i>					1
<i>Sphaerium nitidum</i>		1	4	4	2

Raw benthic invertebrate data identified to the lowest practical level.

Area ID	PDL	PDL	PDL	PDL	PDL
Replicate ID	1	2	3	4	5
Depth (m)	6.5	6.8	6.9	7.6	6.7
# Subsamples/replicate	3	3	3	3	3
Mesh Size (µm)	500	500	500	500	500
Collection Date	13-Aug-11	13-Aug-11	13-Aug-11	13-Aug-11	13-Aug-11
ROUNDWORMS (P. Nemata) ¹	1	2		2	2
ANNELIDS (P. Annelida)					
Cl. Oligochaeta (Worms)					
F. Tubificidae					
<i>Tasserkidrilus americanus</i>			3		3
immatures with hair chaetae ²	3				1
F. Lumbriculidae					
<i>Lumbriculus</i>		1	4		3
ARTHROPODS (P. Arthropoda)					
Cl. Arachnida					
O. Acarina (Mites)	4	1	3	2	1
Cl. Ostracoda (Seed shrimp) ¹	5	4			
Cl. Branchiopoda (Fairy shrimp)					
O. Notostraca					
F. Triopsidae					
<i>Lepidurus arcticus</i>					
INSECTS (Cl. Insecta)					
O. Trichoptera (Caddisflies)					
F. Limnephilidae					
<i>Grensia praeterita</i>		1	1		1
O. Diptera (True flies)					
F. Chironomidae (Midges)					
chironomid pupae ²		1	2	1	1
S.F. Chironominae					
<i>Corynocera</i>					
<i>Micropsectra</i>	1				
<i>Microtendipes</i>					
<i>Paratanytarsus</i>					2
<i>Stictochironomus</i>	15	21	31	5	17
<i>Tanytarsus</i>		3			2
S.F. Diamesinae					
<i>Protanypus</i>	2	1	1		
<i>Pseudodiamesa</i>					1
S.F. Orthocladiinae					
<i>Abiskomyia</i>	1			2	
<i>Cricotopus</i>	2				
<i>Heterotrissocladius</i>		1	4	2	2
<i>Hydrobaenus</i>	1				
<i>Paracladius</i>					
<i>Psectrocladius</i>		2		1	
<i>Psectrocladius</i>	1	6			2
<i>Zalutschia</i>			1	1	1
indeterminate ²					1
S.F. Prodiamesinae					
<i>Monodiamesa</i>	1				
S.F. Tanypodinae					
<i>Ablabesmyia</i>					
<i>Procladius</i>	5	5	4	9	13
<i>Thienemannimyia</i> complex					
MOLLUSCS (P. Mollusca)					
Cl. Bivalvia (Bivalves)					
F. Sphaeriidae (Fingernail clams)					
<i>Cyclocalyx/Neopisidium</i>	13	16	29	21	3
<i>Cyclocalyx</i>	2	3			10
<i>Cyclocalyx nitidum</i>					
<i>Sphaerium nitidum</i>					

Raw benthic invertebrate data identified to the lowest practical level.

Area ID	TPN	TPN	TPN	TPN	TPN
Replicate ID	1	2	3	4	5
Depth (m)	7.0	9.0	9.2	8.6	8.4
# Subsamples/replicate	3	3	3	3	3
Mesh Size (µm)	500	500	500	500	500
Collection Date	19-Aug-11	19-Aug-11	19-Aug-11	19-Aug-11	19-Aug-11
ROUNDWORMS (P. Nemata) ¹	4		2		
ANNELIDS (P. Annelida)					
Cl. Oligochaeta (Worms)					
F. Tubificidae					
<i>Tasserkidrilus americanus</i>	1	1			
immatures with hair chaetae ²		3			
F. Lumbriculidae					
<i>Lumbriculus</i>	1	1			
ARTHROPODS (P. Arthropoda)					
Cl. Arachnida					
O. Acarina (Mites)	2		1	2	
Cl. Ostracoda (Seed shrimp) ¹	1	1	2	2	6
Cl. Branchiopoda (Fairy shrimp)					
O. Notostraca					
F. Triopsidae					
<i>Lepidurus arcticus</i>					
INSECTS (Cl. Insecta)					
O. Trichoptera (Caddisflies)					
F. Limnephilidae					
<i>Grensia praeterita</i>					1
O. Diptera (True flies)					
F. Chironomidae (Midges)					
chironomid pupae ²		5	3	1	2
S.F. Chironominae					
<i>Corynocera</i>					
<i>Micropsectra</i>	4	2		1	2
<i>Microtendipes</i>					
<i>Paratanytarsus</i>	1			1	
<i>Stictochironomus</i>	10	1		1	
<i>Tanytarsus</i>	4	2		2	
S.F. Diamesinae					
<i>Protanypus</i>				1	
<i>Pseudodiamesa</i>					
S.F. Orthocladiinae					
<i>Abiskomyia</i>					
<i>Cricotopus</i>					
<i>Heterotrissocladius</i>	2	3	4	7	1
<i>Hydrobaenus</i>					
<i>Paracladius</i>					
<i>Psectrocladius</i>					1
<i>Psectrocladius</i>				2	1
<i>Zalutschia</i>		3	1		
indeterminate ²					
S.F. Prodiamesinae					
<i>Monodiamesa</i>					
S.F. Tanypodinae					
<i>Ablabesmyia</i>					
<i>Procladius</i>	5	9	2	2	1
<i>Thienemannimyia</i> complex		2	1	2	1
MOLLUSCS (P. Mollusca)					
Cl. Bivalvia (Bivalves)					
F. Sphaeriidae (Fingernail clams)					
<i>Cyclocalyx/Neopisidium</i>	32	27	6	15	5
<i>Cyclocalyx</i>					
<i>Cyclocalyx nitidium</i>					
<i>Sphaerium nitidum</i>					